

Historic, archived document

Do not assume content reflects current scientific knowledge, policies, or practices.

UNITED STATES
DEPARTMENT OF AGRICULTURE
LIBRARY



BOOK NUMBER

748482

1
Ag84Bi
No. 9-11
Dec. 1947-
Dec. 1949

643i 8 of 10
UNITED STATES DEPARTMENT OF AGRICULTURE

BIBLIOGRAPHICAL BULLETIN No. 9

Washington, D. C.

Issued December 1947

3928

REVIEW OF LITERATURE ON CINCHONA DISEASES, INJURIES, AND FUNGI

By

FRANCES F. LOMBARD

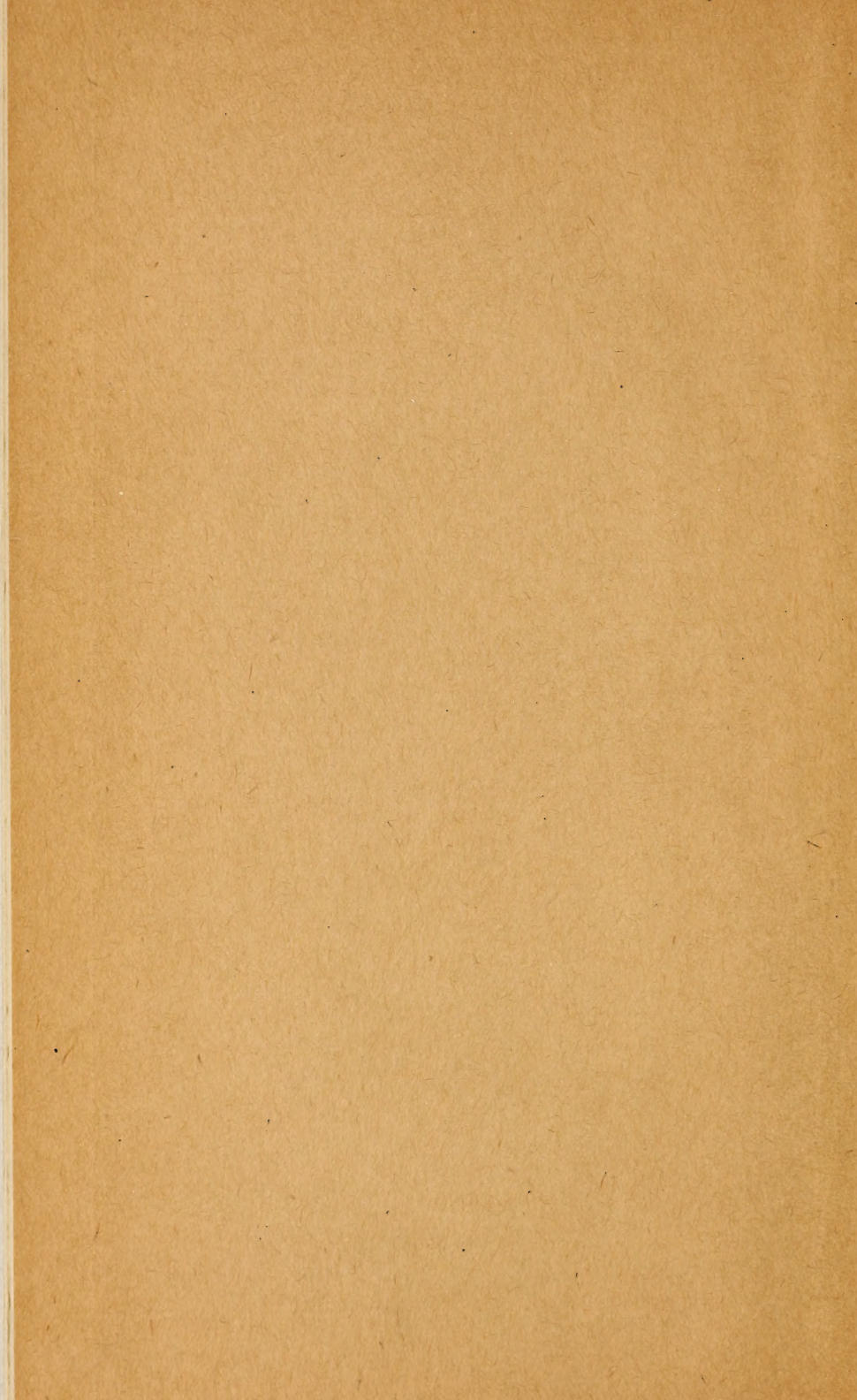
Assistant Pathologist

Division of Forest Pathology

Bureau of Plant Industry, Soils, and Agricultural Engineering

Agricultural Research Administration





UNITED STATES DEPARTMENT OF AGRICULTURE

BIBLIOGRAPHICAL BULLETIN No. 9

Washington, D. C.

Issued December 1947

**REVIEW OF LITERATURE ON
CINCHONA DISEASES,
INJURIES, AND FUNGI**

By

FRANCES F. LOMBARD

Assistant Pathologist

Division of Forest Pathology

Bureau of Plant Industry, Soils, and Agricultural Engineering

Agricultural Research Administration



CONTENTS

	Page
Seedbed diseases	2
Damping-off	2
Mechanical injury	8
Transplant bed diseases	9
Leaf spots	9
Other leaf diseases	11
Stem blights	12
Stem cankers	17
Root rots	18
Nematode root infections	20
Plantation diseases	21
Leaf spots	21
Other leaf diseases	25
Bark diseases	27
Systemic diseases	29
Trunk and branch cankers	29
Root and collar rots	36
Injuries attributed to environmental factors other than soil conditions .	44
Drought	44
Frost or freezing	44
Light	44
Lightning	45
Rain and hail	45
Shade	45
Smoke	45
Wind	45
Miscellaneous fungi and lichens	45
Mycorrhizae	48
Literature cited	49
Index	66

REVIEW OF LITERATURE ON CINCHONA DISEASES, INJURIES, AND FUNGI

By

FRANCES F. LOMBARD

Assistant pathologist, Division of Forest Pathology, Bureau of Plant Industry, Soils, and Agricultural Engineering, Agricultural Research Administration¹

CINCHONA, the natural source of quinine, is a native tree of the slopes of the Andes in South America from Venezuela to Bolivia and in a comparatively limited area in Central America. Quinine and related alkaloids, which are extracted from cinchona bark, are the well-known medicaments for the treatment of malaria. Following the discovery of the medicinal value of the bark during the seventeenth century and the subsequent wholesale harvesting of the wild barks for the European market, efforts were made to introduce cinchona into other parts of the world. In 1865 Charles Ledger sent seeds of a high-alkaloid-yielding variety of *Cinchona officinalis* to England (185).² Part of this shipment of seeds was sold to Dutch planters in Java, who through careful selection and breeding over a period of years were able to develop stock with a quinine content of 3 to 13 percent (99). Aided by the possession of this superior variety, known as *C. officinalis* var. *ledgeriana*, planters of the Netherlands Indies became the foremost producers of commercial bark. When the Netherlands Indies fell to the Japanese during World War II, it was producing more than nine-tenths of the world's supply of cinchona bark (99).

Malaria occurs intermittently in northern and southern Africa, the Mediterranean countries, most of southern Asia and Asia Minor, all of the South and Central American countries north of the Rio Plata, and in most of the Pacific tropics (283). It is endemic in 17 of the Southern and Southwestern States of the United States of America (283). The quinine requirements for human consumption alone in the United States were estimated at 3,000,000 to 4,000,000 ounces per year before the war, of which approximately 60 percent was used by malarial sufferers (190). Following the outbreak of war, deaths from malaria throughout the world were estimated to have increased from 4 to 6 million (283).

¹ Publication is made possible by funds provided through the Interdepartmental Committee on Scientific and Cultural Cooperation, Department of State, in cooperation with the Office of Foreign Agricultural Relations, Department of Agriculture. Valuable assistance was rendered by May Coult, translator, Office of Foreign Agricultural Relations, who translated most of the foreign literature used in the preparation of this manuscript. Thanks are due also to John A. Stevenson, principal mycologist, and Edith K. Cash, associate mycologist, Division of Mycology and Disease Survey, Bureau of Plant Industry, Soils, and Agricultural Engineering, for checking mycological details during the course of the preparation of this publication.

² Italic numbers in parentheses refer to Literature Cited, p. 49.

During the war the important commercial sources of quinine were controlled by the Japanese. To meet the wartime needs of the Allied armed forces and the future needs of the civilian populations, the United States and other countries in the Western Hemisphere cooperated in developing the cinchona industry in this hemisphere. At the same time that cinchona production on existing and new plantations in the Western Hemisphere was being stimulated, bark was being harvested from the regenerated natural stands in Colombia, Ecuador, Peru, and Bolivia. About 18,000 tons were harvested from these wild stands in South America during a $2\frac{1}{2}$ -year period (81). Until a completely satisfactory synthetic product is available, prudence demands that all possible encouragement be given to the establishment of a cinchona culture sufficiently expansive to provide for any exigencies that may arise in the Western Hemisphere.

This publication has been prepared in an effort to bring together the world literature on the diseases of cinchona. With the introduction of cinchona into the Netherlands Indies, India, and other countries, the problem of developing the proper planting techniques for an exotic species and also the problem of preventing and controlling diseases confronted the planters. Many of the early workers recognized the threat to the new industry of uncontrolled diseases in the plantations and the advisability of combating them. When the plants could be brought to harvesting age, however, the incidence of disease assumed a relatively unimportant aspect, particularly where high-alkaloid-yielding varieties were used in large-scale plantings and a consequent good economic return could be realized despite the diseases.

It is not surprising therefore that the main efforts of the early workers were directed toward establishing the plant rather than investigating the disease-producing organisms and that some of the reports and descriptions of the various diseases are so general as to contribute little to the discussions based on the more accurate investigations of later workers.

Present-day pathologists and mycologists will recognize as saprophytic or weakly parasitic many of the fungi that were reported by early writers as occurring on cinchona plants or causing their diseases or death. Since the purpose of this publication is to review the pathological literature on cinchona, most of these earlier reports are included for the benefit of investigators who may not have access to them and who may wish to have the information contained in these older publications at hand.

SEEDBED DISEASES

DAMPING-OFF

The term "damping-off" is used here for any fungus infection that causes the rapid decay or death of young succulent seedlings (119). The disease is of world-wide distribution, affecting many hosts in nurseries and greenhouses (19). The fungi that cause damping-off occur commonly in the soil; and under favorable conditions of moisture and temperature the fungus hyphae attack

the host plants by directly penetrating the host tissues (19). Some fungus species that cause damping-off attack a great variety of hosts, whereas others have a rather limited host range (119). Among the fungi that have been reported to cause or that are associated with damping-off are: *Phytophthora cactorum* (L. and C.) Schroet., *P. cinnamomi* Rands, *P. parasitica* Dast., *Pythium* spp., *Botrytis cinerea* Pers., *Fusarium* spp., *Pestalozzia funerea* Desm., and *Rhizoctonia* sp. (*Corticium vagum* type) (19). A number of fungi that cause damping-off in seedbeds may continue to cause losses in transplant beds³ or may cause other diseases on the mature plants (119). The so-called "late damping-off" of seedlings that have developed woody supporting tissues is usually considered an early stage of root rot (50).

Several diseases that have been reported on cinchona can be classed as damping-off diseases, although the authors may not have used that name specifically in reference to them. For example, Loble (174) stated that in cinchona seedbeds fungus infection is most likely to occur just after the seeds have begun to germinate. Following is a discussion of fungi that have been reported to be associated with or to cause damping-off.

Cladosporium

Losses in cinchona seedbeds caused by a fungus that attacks the radicle before it has penetrated the soil have been reported at Mulungu and Tshibinda, Belgian Congo, by Hendrickx (122), Stoffels (262), and Feilden and Garner (78). The fungus destroys the positive geotropism of the rootcap (122), with the result that the radicle does not enter the soil and the seedling is left lying flat on the ground (78). Correct watering and good ventilation during the period of germination retard the development of the fungus (78). Spraying with a copper fungicide at the time the seeds begin to swell also achieves some control, according to Feilden and Garner (78). Hendrickx (122) stated that an *Alternaria* was isolated from the diseased material but that inoculation tests had not been made. Later, Stoffels (262) wrote that the causal organism has been found to be a physiologic race of *Cladosporium herbarum* (Pers.) Lk.

Cladosporium herbarum (Pers.) Lk.—"Caespitulis dense aggregatis, confluentibus, stratum velutinum flavo-olivaceum dein atro-olivaceum constituentibus; hyphis erectis vel adscendentibus, brunneis vel olivaceis, paullum ramosis, septatis, 5-7 μ crassis; conidiis prope hypharum apicem nascentibus, non vel parce concatenatis, dilute brunneis vel olivaceis, forma et magnitudine variabilissima, oblongis, ovoideis, simplicibus vel oblongo-ellipticis cylindraceisve, 1-3-septatis, ad seipimenta constrictis, levibus." (242, v. 4, pp. 350-351.)

³ The term "transplant beds," as used in the organization of this publication, includes all nursery beds in which the cinchona plants are grown from the time they are moved from the original seedbeds until they are placed in permanent field locations. The terminology used in the text follows as closely as possible that used in the original articles cited. Scientific names of fungi given within parentheses and indicated by sign of equality (=) are added to bring the nomenclature to date.

Clitocybe

In 1940 Heim (120) reported a destructive seedling wilt of *Cinchona ledgeriana*⁴ and *C. succirubra* in French Guinea. The seedlings wilted and died in patches in the seedbeds when the cotyledons were about 4 to 5 mm. long. Numerous white rhizomorphs of a tropical species of *Clitocybe* were found in the soil of the affected patches; and on contact with the cinchona rootlets the rhizomorphs produced mycelial threads that penetrated the host tissues. The methods recommended by Heim for preventing and combating the trouble were: Presterilization of seedbed soil by using formaldehyde (2 to 4 percent), aromatic nitrated phenols (ortho-2-4-dinitrophenol), or steam; the removal of the *Clitocybe* mycelial mats from the beds; and careful drying of the seed flat soil in the sun and air.

Clitocybe sp.—Fruiting body abortive, sterile; stipes dirty white, 22 mm. high; caps less than 3 mm. in diameter, with triangular profile, flattened on the top; peridium dark blue gray, highly involute; gills plicate, about 24 in number, thick, strongly decurrent, whitish, with sterile hymenium; flesh whitish, odorless; cap and stipe not separable; rhizomorphs milky white, branching, 250 μ to a few microns in diameter, composed of "constituent hyphae" and "protective hyphae"; constituent hyphae parallel, thin-walled, constricted, septate, colorless, constituting the central part of the rhizomorphs; protective hyphae of irregular contour, slender, not constricted, rarely septate, thick-walled, interlaced, often swollen, constituting the peripheral part of the rhizomorphs (120).

Fusarium

Barat (6) in 1931 reported a *Fusarium* attacking germinating cinchona seedlings in south Indochina. He stated that the seed appeared to have been infected because the *Fusarium* developed on seed that had been sown on sterilized sand.

(See fusarium stem blight, p. 12, and fusarium root rot, p. 38.)

Pestalozzia

A disease on cinchona seedlings at Pala, Sumatra, was reported by Hunger (131) to have been caused by *Pestalozzia myristicae*. The fungus attacked the cotyledons of the seedlings, and the plants died. Removing and burning infected plants immediately was advised.

Pythium

Tschirch (271) stated that according to Breda de Haan a species of *Pythium* attacks cinchona seedlings if the beds are improperly watered. Reinking⁵ reported that a species of *Pythium* was present in cinchona seedbeds in which the seedlings were affected by damping-off in Guatemala.

⁴ The scientific names for the species of *Cinchona* used in the text are those given in the publications cited and cannot be checked for accuracy of identification.

⁵ REINKING, O. A. REPORT ON CINCHONA DISEASES IN GUATEMALA. U. S. Bur. Plant Indus., Soils, and Agr. Engin., Plant Dis. Rptr. 29: 432-439. 1945. [Processed.]

Rhizoctonia

As early as 1882 Moens (189) recorded a disease that occurred in cinchona seedbeds in Java and that resulted in the death of seedlings in patches and was caused by a "cobwebby mycelium." Koorders (154) observed the same disease in Preanger, Java, on cinchona plants that were only a few days old. He found a thin, colorless mycelium in the stems, hypocotyl region, and roots of all dead or dying seedlings. In 1908 and 1915 Rant (220, 228, 229) described the disease under the name "mopo" (usually called "mopog" in the Dutch literature). From his cultural studies he identified the fungus as *Moniliopsis aderholdii* Ruhl. (229), which was shown later to be a synonym of *Rhizoctonia solani* Kuehn (= *Corticium solani* (Prill. and Del.) Bourd. and Galz. = *Pellicularia filamentosa* (Pat.) Rogers). Following Rant's investigations and identification of the fungus, numerous reports of the occurrence of mopog at the governmental cinchona stations and on private estates in Java and Sumatra have been made in the literature (13, 44, 45, 54-58, 61, 62, 64-67, 88-93, 100, 102-108, 110-112, 139-144, 148-150, 157-161, 167, 170, 205, 210, 227, 252, 259, 271). The disease was rather serious on a few private plantations in Java and Sumatra in 1926 and 1930 (157, 160), causing a 10 percent loss in the seedbeds in 1926 (157, 252).

Rant (228) stated that the mopog fungus may attack cinchona seeds either before or after germination. In some cases it attacks seedlings so young that at no time do the seedlings ever stand erect (161); and it also attacks seedlings large enough to transplant (92, 161). In the seedbed bare places containing dead plants and ungerminated seed appear and these are surrounded by diseased plants (13). The infected spots quickly enlarge under favorable moisture conditions, and a large number of seedlings may be lost in a short time (228). The infected seedlings collapse on the top of the ground and remain green for several days, but appear to be swollen (205). A dark spot may be seen on the lower part of the stem, and the seedling will break off at this point if touched (13). Rant (228, 229) stated that a characteristic symptom of mopog is the tendency of the particles of soil, ungerminated seeds, and dead seedlings to hang together by the mycelial filaments of the fungus when the soil is lightly stirred or lifted. If the humidity in the seedbed is high, the mycelium grows over the dead plants and seeds (229). Rant successfully inoculated hybrid cinchona seedlings 4 to 6 inches high with the mopog fungus and obtained the disease. Similar inoculations with *Botrytis cinerea* Pers. failed to give the typical mopog symptoms (229).

Careful adjustment of light and ventilation and moderate watering with a fine spray were recommended by Rant (229) as preventive measures. Keuchenius (147) reported that, although 5 million plants were started in the seed and transplant beds in West Sumatra in 1935, there were no losses from mopog, because the germination houses were built so as to allow proper regulation of the relative humidity. The addition of wood ashes to the soil of infected seedbeds is said to discourage the growth of the fungus,

(229) and Stoffels (262) advised replacing contaminated patches of seedbed soil with wood ashes or sulfur. Keuchenius stated that mopog was successfully controlled by spraying with a Cryptonol solution (149) and that the Bayer fungicides "Nosperit" (75-percent solution) and "Terbolan" (5-percent solution) were being tested (150). Coster (45) reported that *Rhizoctonia solani* could be controlled with "Superol."

Rhizoctonia solani Kuehn.—Hyphae practically colorless when young, vacuolate, more or less irregular, septate with septa at intervals of 100μ to 200μ ; some development of external hyphae on certain hosts; external hyphae somewhat colored, usually yellowish brown, and generally of two types, a purely vegetative type and a second type constituting the short tufted or mealy growth as occurring on certain hosts; vegetative hyphae, 8μ to 12μ , branches when young pointing in the direction of growth and somewhat constricted at point of union with main hyphae, branches when older approximately at right angles to the main hyphae, more deeply colored and more uniform and rigid, constrictions less marked and septa farther apart; hyphae of tufts profusely branched when young, lobed, sometimes botryoid, becoming light brown with maturity, ultimately dividing into short hyphal lengths or single ovate cells, arranged in short chains or elbowed and producing branches in a more or less dichotomous fashion, the single cells somewhat resembling conidia, which may germinate by tube within a few hours under suitable conditions, the denser masses in culture giving rise to sclerotia; hyphae in tissues colorless while in active growth, smaller in diameter but otherwise similar to the young external hyphae; sclerotia of fairly homogenous structure, generally more or less flattened, irregular, deep chestnut brown, generally smooth under natural conditions, ranging from a minute size to 1 to 2 cm. in diameter (51, pp. 439-444).

In 1941 Madarang (182) reported that damping-off is estimated to cause a 20 to 30 percent loss in the Government cinchona plantation at Bukidnon, P. I. The disease is said to be most injurious from the time the seedlings emerge from the soil until they are about 2 inches high. A species of *Rhizoctonia*, similar to or identical with *R. solani* Kuehn, was isolated from infected seedlings. This disease occurs in patches in the seed flats; and when it is allowed to go unchecked, most of the seedlings may be lost. The fungus prevents seed germination by attacking the radicle and cotyledons before they emerge from the seed, finally causing them to deteriorate. Also seedlings that have emerged are attacked at or near the ground line, and when the stems have rotted at the point of infection the seedlings topple over. The infected seedlings continue to decay if the soil and atmosphere are moist or they wilt and shrivel if the atmosphere is dry. Occasionally leaves touching the soil may become infected, while the rest of the plant escapes. Heavy mycelial growth may smother small seedlings.

Soil inoculation tests to prove the pathogenicity of the *Rhizoctonia* were successful on *Cinchona succirubra*, and the causal organism was reisolated. Limited inoculation tests on other host plants—a native species of tomato, Irish potato, and *Pinus insularis* Endl.—showed the cinchona strain of *Rhizoctonia* to have a narrow host range. Sterilization of the soil by steam pressure at 15 pounds for 2 hours or drenching the soil with 40-percent commercial formaldehyde diluted 1 : 50 with tap water and applied

at the rate of 2.5 liters per square foot before seeding effectively checked damping-off of cinchona seedlings. Seed dusted with cuprous oxide at the rate of 2 grams dust to 100 grams seed showed a higher percentage of infection than the controls and a very low percentage of germination. The poor germination of the seed dusted with cuprous oxide was believed to have been due to injurious effects of the dust or to have been caused by premergence infection in the presence of the dust or a combination of the two factors (182).

Sundararaman (265) reported in 1928 that cinchona seedlings at the Government cinchona plantation at Anamalai, Madras, India, were dying in patches from a damping-off caused by *Rhizoctonia* sp.

Rhizoctonia sp. is reported to have caused 100 percent loss in certain cases in the cinchona seedbeds in Belgian Congo (135). Mercuric chloride and formaldehyde when used as soil disinfectants retarded infection but did not suppress it completely (135).

The organisms causing damping-off of cinchona in Guatemala have not been identified; however, Reinking⁶ reported in 1945 that a species of *Rhizoctonia* had been isolated from infected plants and a species of *Pythium* was present also in affected beds. The disease was observed on *C. ledgeriana* and *C. calisaya* and was not serious in well-managed plantings visited by Reinking.⁶ Control measures suggested were the use of new soil in each new seedbed and soaking the soil of severely attacked beds with bordeaux mixture or Yellow Cuprocide.⁶ Popenoe (209) stated that cinchona is rarely attacked by damping-off if the seed is sown at the beginning of the dry season (January or February in Guatemala) and if the seedlings make 2 or 3 months' growth before the heavy rains begin.

(See rhizoctonia root rot, p. 18.)

Undetermined

In 1877 a disease that affected cinchona seedlings immediately after germination was reported in Jamaica (241).

In 1880 seedlings at the Government cinchona plantations in Sikkim, India, were reported to have died, owing to the occasional presence of a fungus mycelium in the seedbeds during very damp weather (153). Gently stirring the soil was said to have some value in checking the disease (153).

A fungus disease that occurs during periods of excessive humidity in the "nurseries" in Peru was reported by Augusto (5) in 1943. Bordeaux mixture is used to control it (5).

Damping-off in Puerto Rico is said to be caused by a basidiomycete (216). Although the causal organism has not been identified, control measures have been investigated. Semesan and Cuprocide dusts used as seed and seedling protectants against damping-off were found to be harmful to the seed and to reduce

⁶ See footnote 5, p. 4.

the stand by 90 percent or more; however, liquid applications after germination were not injurious to the plants and appeared to control the spread of the disease (261).⁷ Untreated seed and seed treated with calcium hypochlorite, when planted in soil disinfected with formaldehyde and pretreated with hot water, germinated equally well, and neither lot of seed was attacked by damping-off organisms (215). Cinchona seeds, which were disinfected with calcium hypochlorite and planted on sterile agar slants for germination counts, were attacked by an unidentified fungus (215). The ridged, uneven surface of the seed coat was thought to be responsible for the inadequate disinfection, permitting the development of the fungus growth (215).

Boecop (15) reported a "root collar" disease in Java that affected cinchona seedlings about 1 to 2 cm. high. The stem at the soil level is infected, while the root remains healthy and the leaves remain green for a considerable time after the plant becomes affected. In the early stages of the disease the plants show a tendency to droop slightly and, when brushed lightly with the hand, do not spring back to an upright position. The seedlings fall flat on the ground finally and break off at the collar when an attempt is made to pull them up. The disease occurs in patches in the seedbed. Boecop was unable to isolate a fungus from either the diseased plants or the soil. He stated that control was obtained by removing the infected plants and soil, leaving the vacant space open to the air and light, and withholding water from the bed for a week.

MECHANICAL INJURY

Coprinus and Marasmius

Mechanical injury to seedlings caused by the lifting of the soil when the fruiting bodies of species of *Coprinus* and *Marasmius* are formed has been reported by Hendrickx (122) in seedbeds at Mulungu and Tshibinda, Belgian Congo.

Stemonitis

The suffocation of cinchona seedlings by the plasmodia of a myxomycete was reported from the Preanger and East Java, Netherlands Indies (154, 167, 193, 227, 271, 286). The affected plants were about 2½ cm. in height, and both sides of the leaves were thickly covered with the fruiting bodies of a species of *Stemonitis* (286). The seedlings had died when the overgrowing plasmodia obstructed the stomata of the leaves and cut off exposure to light (286). Wurth (286) recommended spraying the plasmodia with common salt or saltpeter or, after the formation of sporangia, the removal and burning of the affected plants and disinfection of the soil with unslaked lime or 1-percent carbolic acid.

⁷ STODDARD, D. L. TREATMENT OF CINCHONA SEED WITH FUNGICIDES GAVE POOR RESULTS. Rpt. Activities of Puerto Rico (Mayaguez) [Fed.] Expt. Sta. for months of July, Aug., and Sept., 1943: 8. 1943. [Processed.]

Turf fungus

Cinchona seedlings were reported to have died as the result of an unusual condition in the seedbeds at the Government cinchona station in Java (169, 224, 226, 227). In some spots in the beds the seeds had not germinated, and in other beds there were spots of backward plants among the healthy ones (224). The affected plants were small, and the leaves were red (169, 224). Examination of the affected seedlings failed to show any causal organisms (224). The ground felt peaty and elastic to the touch; and when such a spot was dug out, a brown mass of fungus mycelium was found a few centimeters below the surface (224). The fungus was found growing luxuriantly and forming thick brown rhizomorphs (169, 224). In the absence of fructifications Rant was unable to identify the fungus (224). If the plants succeeded in penetrating the mass of mycelium, they grew normally, although they remained smaller than the unaffected plants (224). In 1936 Goot (92) reported that a comparatively large number of plants were lost on an estate where the soil used in the seedbeds was forest humus and full of the turf fungus. Rant (224) advised replacing the contaminated seedbed soil with soil from another place in the forest or making a new seedbed.

TRANSPLANT BED DISEASES

LEAF SPOTS

Colletotrichum

Koorders (154, 155) described a new species, *Colletotrichum cinchonae*, causing a leaf spot on 1-year-old seedlings of a cinchona hybrid in Java. The seedlings were growing at an altitude of 1,500 meters above sea level (227). The leaf spots were pale and the sori scattered (155, 227). The plants attacked by this fungus had already been attacked by an undetermined fungus and the *Colletotrichum* species did no serious damage (155, 227). *Cinchona calisaya* plants in the Transcaucasian region of Russia (U.S.S.R.) are reported to have been attacked by *Colletotrichum cinchonae*; however, the leaf spots were reddish brown (210).

Colletotrichum cinchonae Koord.—“Maculis pallidis foliiculis; acervulis sparsis, punctiformibus, epidermide erumpentibus, vix 0.1 mm. diam.; sporulis oblongis, subrectis v. curvulis, utrinque obtusis v. rotundatis, 9–18 × 3–5, plerumque 10–12 × 3.5–4, hyalinis; setulis paucis, fuscis, 1-pluri-septatis, subrectis v. curvulis v. flexuosis, 60–75 μ longis, basi 4–7 μ diam.; sporophoris cylindraceis, hyalinis, c. 10 μ longis.” (242, v. 22, p. 1203; also described in 155, 227.)

Gloeosporium

Bugnicourt (24) reported a withertip of cinchona seedlings in South Annam, Indochina. The affected seedlings were from 8 months to 1 year old. The infection started with the terminal bud. The upper three or five internodes withered and shriveled, and the adjacent leaves and diseased parts turned black. The whole top drooped. When the disease was allowed to develop, the infection spread downward and the plant died. A species of *Gloeosporium* was isolated. Bugnicourt suggested that the initial

infection might have been caused by infected seed. He advised disinfection of seed, cutting off the diseased parts, painting the pruning wounds, and spraying with bordeaux mixture.

Bugnicourt (24) also described an anthracnose leaf spot affecting young plants in seedbeds and nurseries in Laos, Indochina. The leaf spots, which were brown at first and then turned yellow, affected the entire thickness of the blade. Later the affected parts of the leaf dropped out. The leaf disease was also caused by a species of *Gloeosporium*, and Bugnicourt considered it a form of the withertip disease in South Annam.

Gloeosporium sp.—Mycelium in culture dense, white, hyphae 2μ to 3μ , forming fine orange droplets; conidia abundant, hyaline, somewhat cylindrical; conidiospores $15\mu \times 4.5\mu$ (24).

(See gloeosporium leaf spot, p. 22.)

Lapp disease

Lapp disease, or "leaf scab," occurs on the leaves of *Ledgeriana* grafts and seedlings in the seedbeds, nurseries, and plantations in Java and Sumatra (61, 109, 126, 132, 144, 147, 148, 167, 168, 227). Grafts are said to suffer considerably less injury than the young seedlings (147). The infected plants have curled or wrinkled brittle leaves that tear easily (167). The leaves are red at the edges, and brown spots with red borders are found between the veins (167). Occasionally only one part of a plant is affected, the rest of the leaves being healthy (227); and the plants or affected parts generally appear to be rather slender (227). The disease may attack widely scattered plants in the plantations or plants that are growing close together in the seedbed (167).

Although the occurrence of the disease has been noted in the reports from the Netherlands Indies for about 30 years (1907–36), the cause is still unknown. Microscopic examination and attempts to isolate a causal organism from affected plants gave negative results (168). Rant (as reported by Leersum, 168) was of the opinion that the type or condition of the soil was responsible for the disease, although he indicated that insects might be the cause of the damage. Hall (109) stated that some plants affected with Lapp disease could always be found in young plantations and nurseries, but as the disease is generally limited to a comparatively small number of plants only a small extent of damage is done. He reported that the disease had broken out so severely in Sumatra in 1922 in both the nurseries and young groves of *Ledgeriana* seedlings, however, that a very large number of plants had been killed (109). Hunger (132) stated that in 1907 no attempt to combat the disease was necessary, as it gradually diminished and finally disappeared.

Phyllostictina

In recent years (1935–1938) a species of *Phyllostictina* has been reported to occur periodically in cinchona nurseries as a leaf disease on young grafts of Clone R. G. 1 in West Sumatra (93, 147, 148, 150). This fungus is believed to be identical with *Phyl-*

losticta cinchonae Koord. (147). The application of lime, nitrogen, and phosphoric acid to the soil and repeated spraying with bordeaux mixture are said to give satisfactory control (147). (See *Phyllosticta cinchonae*, p. 23.)

OTHER LEAF DISEASES

In 1921 and 1922 Vincens (273, 274) reported a disease in the cinchona nurseries at the Honbà station, Annam, Indochina. *Cinchona ledgeriana* was particularly susceptible, although *C. succirubra* and a hybrid (*C. ledgeriana* \times *C. succirubra*) were also attacked (274). The diseased Ledgeriana plants were only 15 to 20 cm. high, while the better developed Ledgeriana plants were about 35 to 40 cm. in height (274). No plant was entirely healthy, heavily spotted and often perforated leaves being found on plants least affected (274). Older leaves, showing comparatively little damage, had circular brown spots a few millimeters in diameter with purplish-blue borders or perforations from which the contents of the leaf at those places had dropped out (274). The spots were more numerous on the younger leaves, often uniting to form checkerboard designs occupying a large part of the leaf area (274). The leaves near the top of the plants were misshapen, wrinkled, and tattered looking, similar in appearance to leaves severely attacked by anthracnose (274). The young green shoots were bent and deformed, sometimes slightly enlarged, and occasionally had abnormal suberose formations with a cankerous appearance (274). Only a few misshapen leaves remained on the weak, dying plants, the majority having already fallen to the ground (274).

The disease showed the same characteristics on the hybrids, except that the shoots of the hybrids looked healthy despite the diseased condition of the leaves (274). Badly infected leaves were comparatively scarce on the *C. succirubra* plants and all the shoots were healthy (274). Vincens (273) stated that according to Dr. Yersin, Director of the Honbà Station, the same disease has resulted in much damage in Java where its cause is unknown.

The disease at Honbà was said to be favored by the frequent mountain mists (274). The use of copper sprays proved both ineffective in the control of the disease and injurious to the young cinchona plants (273, 274). New plantings of Ledgeriana were free from disease, and further studies on control measures were unnecessary (274). In the earlier report Vincens stated that the disease appeared to be caused by a species of *Guignardia* (273); however, in his later report he said that it was impossible to observe any parasite on the majority of the diseased leaves still attached to the plants and that no parasite was found on or near the suberose young shoots (274). Parts of diseased plants later revealed the fruiting bodies of several fungi, however, and these were described by Vincens (274) under the following names: *Dendrophoma cinchonae* Vincens, *Guignardia yersini* Vincens, *Phlyctaena cinchonae* Vincens, *Phoma cinchonae* Vincens, *Phyllosticta cinchonaecola* Vincens, *P. honbaensis* Vincens, *P. yersini*

Vincens, and *Physalospora cinchonae* Vincens. Vincens stated that there was not much doubt that the *Phyllosticta* species were harmful and that the frequent occurrence of the *Phlyctaena* in the abnormally suberose bark of the plants led him to consider it a parasite (274).

Phyllosticta yersini Vincens was reported on the leaves of cinchona plants in a greenhouse in Russia (U.S.S.R.) (18).

(See *Phyllosticta*, p. 22.)

STEM BLIGHTS

Diplodia

Arens (3) reported that *Cinchona ledgeriana* plants, which had been topped and potted preparatory to an entomological investigation in Java, had been killed by *Lasiodiplodia theobromae* Griff. and Maub. The fungus attacked the plants at the wound, the newly formed shoots were killed back, and finally when the roots became infected the plants died (3). According to Nowell (194) *L. theobromae* is a synonym of *Diplodia theobromae* (Pat.) Nowell.

Diplodia theobromae (Pat.) Nowell.—“B. peritheciis gregariis, 200 μ , latis, atris, plus minus villosis, stromate villosis, atro, junctis; basidiis elongatis; hyalinis (50 μ) apicem versus incrassatis; sporulis ellipticis, 1-septatis, hyalinis dein brunneis, 25–35 \times 12–15 μ .” (201, p. 136; also described in 194 and in 242, v. 11, p. 522.)

(See *Diplodia theobromae*, p. 35.)

Roepke (237) reported that cinchona plants being used in entomological investigations in Java were killed by the *Diplodia* fungus.

Fusarium

An epidemic wilt disease that appeared in the Munsong cinchona plantation in Bengal, India, in 1926–27 (30), and later at Mungpoo (32), continued to be quite serious for about 10 years (30–35, 84, 178, 188, 199, 200, 246, 247). A few plants became diseased in October of one year, and by April and May of the following year the disease had become of such epidemic proportions that thousands of plants had died (178). The disease is believed to attack the plants at midnursery age (31), but usually develops just after the seedlings are set out in the field (30). Coppice shoots are said to be immune (31). Plants in 2- or 3-year-old nursery beds are more severely attacked than those in new beds, although the new beds are not entirely free of the disease (178). The disease may appear in nurseries at both high and low elevations (178). On plants 15 months old the leaves turn dingy yellow and then red and finally wilt (178). Slight longitudinal depressions appear in the bark (178), extending from the collar up to the first or second branch (30). The largest lesions are about one-fourth inch wide and several inches long (178). The bark beneath the depressions becomes brown and dry, and the brown area extends beyond the cambium into the wood (178). Mildly attacked plants may throw off the disease after being planted in the open (32); more severely attacked plants die after a period

of varying length (30). Fungal hyphae were found sparingly near the advancing edges of the lesions (178).

During the investigations on the disease, species of *Botryodiplodia*, *Diplodia*, *Fusarium*, *Nectria*, *Sporotrichum*, and *Verticillium* were isolated from a large number of the diseased plants (188). Inoculations with some of these organisms gave negative results (188), but the *Sporotrichum* and *Verticillium* isolates gave positive infections (181). Galloway (84) suggested that the *Diplodia* might be the causal organism. In 1939 Padwick (199) stated that a species of *Fusarium* was regularly isolated from the vascular tissues of the diseased plants; and Mitra (188) reported that the *Fusarium* had been isolated from both the roots and the collar region. One-year-old seedlings were inoculated with the *Fusarium* after damaging the bark, and seed and transplant bed tests were made by infecting the soil with three *Fusarium* isolates (200). In these tests infection had not yet taken place at the end of 3 months (200). Selection of disease-free plants for the plantation at the time of lifting from the nursery beds is difficult because the disease symptoms usually are not evident at that time (34). The continual development of new nursery bed sites (31, 35), disinfection of the nursery soils and sites (31), and transplanting to the plantations at an earlier age (178) have reduced the losses somewhat, but have not proved to be very satisfactory control measures. The effectiveness of watering the beds with weak fungicidal solutions of "Kerol" and malachite green was being tested as a control measure (84).

Fusarium and *Helminthosporium* were reported by Keuchenius (150) to have caused injury to 4-month-old seedlings in the West Sumatra area in 1938. Spraying with a 5-percent Cryptonol solution was recommended.

(See fusarium damping-off, p. 4, and fusarium root rot, p. 38.)

Phytophthora

In 1936 Sawada (245) reported an epidemic disease affecting cinchona seedlings in a plantation in Formosa. The 1-year-old seedlings, which were 10 to 30 cm. in height, were attacked during the rainy season from May to August, and about half of the 20,000 seedlings in the plantation were lost. The fungus isolated from the diseased seedlings was a species of *Phytophthora* to which the author assigned the name *P. cinchonae* Saw. The first symptom to be noticed is a darkening of the stem, followed by the spread of the infection to the leaves, which shrivel and die, and finally by the death of the plant. Positive results were obtained in inoculation tests, and the causal organism was reisolated. Sawada recommended presterilizing the seedbeds with formalin, draining the seedbeds well, burning infected seedlings, and spraying the infected soil in the seedbed with bordeaux mixture.

Phytophthora cinchonae Saw.—Hyphae continuous when young, forming a few "pseudo-septa" in old age, branching profusely on media, thin, membranous, colorless, commonly 5μ to 6μ in diameter on soybean agar, 4μ to 8μ on millet broth agar, 4μ to 15μ on polished rice media, 7μ to 8μ on citrus

agar; aerial hyphae 6μ to 11μ in diameter on millet broth agar, 4μ to 7μ on polished rice media, 6μ to 10μ on citrus agar; sporangiophores about 4μ in diameter; sporangia terminal, colorless, smooth, oval-shaped or sometimes dumbbell-shaped with rounded base and flattened papilla at apex, 18μ to $68\mu \times 13\mu$ to 38μ (average of 48 specimens, $41.12\mu \times 23.0\mu$), secondary sporangia borne on short sporangiophores branching from below the terminal sporangia or by proliferating through the empty terminal ones, germinating to produce free swimming zoospores or by germ tubes; zoospores biciliate, 15μ to $17\mu \times 12\mu$ to 13μ while swimming, 12μ to 16μ in diameter when at rest, germinating after a short while by germ tubes 2μ in diameter to form small sporangia $13\mu \times 10\mu$ (which in turn produce zoospores) or by producing zoospores directly without forming the germ tubes; oospores not formed on artificial media (245).

Thompson (269) in 1940 reported a wilt of cinchona seedlings in the Cameron Highlands, Malaya, caused by *Phytophthora cinnamomi* Rands. *Cinchona ledgeriana*, *C. succirubra*, and a hybrid (*C. succirubra* \times *C. ledgeriana*) were affected. The plants were attacked at the collar by a canker and bark rot and the stems split to a height of 3 to 9 inches. The leaves of the diseased seedlings had previously suffered repeated insect injury. The fungus also caused a root rot and dieback of mature trees.

Phytophthora cinnamomi Rands.—“Maculis in cortice Cinnamomi burmanni perpendicularibus, elongatis, linearibus vel irregularibus, 1–5 cm. latis, saepe supra terram usque ad 10 M. in altitudinem extensis insita; cortice mortuo radiatum zonato. Mycelio in cortice et ligno exteriore, irregulari; hyphis aëris in agare avenae cultis tenuibus, 5–7 μ crassis, primo continuis, deinde septatis; haustoria absunt. Chlamydosporis et inter et intra cellulas matricis formatis, tenuibus, globosis vel pyriformibus, plerumque 31–50 μ in diametrum, in apice ramorum brevium nascentibus, copiosis in culturis artificialibus, saepe 3–10 in racemis coalitis; hyphis 3–11 germinantibus. Conidiophoris non a mycelio distinctis, simplicibus vel sympodio-ramosis. Conidiis neque in natura neque in culturis artificialibus visis, sed copiose nascentibus in mycelio e solutione nutritia in aquam translato, primo terminalibus, plerumque ovoideis vel ellipsoideis, hyalinis, tenuibus; plerumque 38–84 \times 27–39 μ , fere 57 \times 33 μ latis, papillis latis, brevibus; conidiis ulterioribus sympodiliter e ramis conidiophori nascentibus, saepe hypha fertile proliferata e conidiis vacuis egredientibus; parietibus conidiorum expulsiōne finita partim collapsis; conidiis in aqua fere zoosporis, sed nonnumquam tubo vel conidiis secundariis germinantibus; conidiis e culturis infirmis vel contaminatis saepe abnormaliter germinantibus. Zoosporis 8–40, plerumque concavoconvexis, ciliis duabus inaequalibus longitudine, lateraliter insertis, mobilibus, motis circa 11 \times 18 μ , quietis 10–11 μ in diametrum, tubo germinantibus. Oosporis non visis.” (219, p. 41.)

(See *phytophthora* root rot, p. 40.)

In 1934 Celino (39) reported a serious blight on *C. calisaya* var. *ledgeriana*, *C. hybrida*, and *C. succirubra* seedlings in the Philippine Islands caused by *Phytophthora faberi* Maub.⁸ Infection takes place in the tips of the young shoots of the seedlings, which suddenly wilt and dry up, and spreads rapidly downward, causing the death of the leaves and petioles (39). Occasionally, infection begins at the tips or margins of the leaves (39). The dead, dry leaves persist on the seedlings, hanging downward (39, 183). If the infection is unchecked, the seedlings die within 5 or 6 days (39).

⁸Tucker (272) recognizes *Phytophthora faberi* Maub. as a synonym of *P. palmivora* Butl. Descriptions of both *P. faberi* and *P. palmivora* are given for the reader's information.

One-year-old *C. ledgeriana* and *C. hybrida* plants were inoculated on the uninjured young shoots and on the petioles of the leaves; by the second day after inoculation the shoots were discolored and wilted and the leaves were slightly wilted and hung downward (39). The causal organism was reisolated 6 days later and again in 10 days (39). Celino reported good control by removing and burning infected parts of the seedlings and spraying twice with standard bordeaux mixture at an interval of 2 weeks (39). Ocfemia (195) also stated that *P. faberi* causes a seedling and shoot blight of cinchona.

Phytophthora faberi Maub.—“Maculis irregulariter brunneis; mycelio ramoso, primum continuo demum septato; sporangiophoris 150–200 μ longis; haustoriis non visis; sporangiis plerumque ovoideis sed magnitudine variabilibus, 42–80 \times 25–30; oogoniis intramatrixalibus; antheridiis nullis v. raris; oosporis plerumque parthenogenetice ortis, cavitatem oogonialem late occupantibus, sphaeroideis, elongatis v. irregularibus 22–45 μ diam.” (242, v. 21, p. 860.)

As early as 1928 a seedling blight of cinchona was reported at the Mungpoo plantations in the Darjeeling District, Bengal, India (177). A species of *Phytophthora*, later identified as *P. palmivora* Butl.,⁸ was isolated and studied (83, 177–180); and in 1935 Kheswalla (152) reported the results of the cultural studies on the pathogen and the inoculation tests. The disease occurs in very wet weather or when the nursery has been overwatered (178). Usually, infection first becomes evident at the root collar, where the tissues become discolored, and then infection extends up the stem (152). The cotyledons wilt; and the leaves turn yellow and curl inward and, in severe cases, are shed (152). Occasionally, seedlings become infected at the tip and the infection spreads downward (152). When the discolored areas of the bark tissues have rotted, the plant droops (152). During damp weather copious mycelial growth may be seen on infected seedlings (152).

At Pusa (temperature about 27° C.) healthy cinchona seedlings 1 foot high were successfully inoculated either at the base of the stem or on the tender leaves and the fungus reisolated (152). Seedlings inoculated at Sureil (temperature about 20° C.) did not become infected (178). Other inoculations indicated that *Phytophthora parasitica* Dast. and *P. colocasiae* Rac. were unable to parasitize cinchona seedlings, while the *P. palmivora* isolate from palm gave only partial infection (152). All cinchona seedlings inoculated with the *P. palmivora* isolate from cinchona succumbed (152).

Phytophthora palmivora Butl.—“Maculas siccas, brunneas, profundas in vaginis fol. generans; mycelio intercellulari, haustoriis vero intra cellulas penetrantibus; hyphis latis, interdum irregulariter inflatis, 7 μ diam., ramosis, haustoriis digitatis; sporangiis obverse piriformibus, raro subrotundis, acrogenis, 50 \times 35; zoosporis 8–10 μ diam.; oosporis sphaericis, 35–45 μ diam., membrana 4 μ cr.” (242, v. 21, p. 855; v. 24, p. 36.) McRae (177) gave the extreme range in sporangia size of the cinchona isolate as 30 μ –79 μ \times 21.6 μ –45 μ .

⁸ Tucker (272) recognizes *Phytophthora faberi* Maub. as a synonym of *P. palmivora* Butl. Descriptions of both *P. faberi* and *P. palmivora* are given for the reader's information.

(See *Phytophthora palmivora*, p. 32.)

Crandall and Davis (49) described a *Phytophthora* disease affecting seedlings of *C. pubescens* Vahl., *C. officinalis* L., *C. officinalis* var. *ledgeriana*, and *Ledgeriana* grafted on *C. pubescens* in Guatemala and on *C. officinalis* var. *ledgeriana* in Peru and Puerto Rico. Infection appears to take place in the petiole region of the tender young leaves or in the succulent stem tissue. Brown necrotic spots with indefinite margins are formed on the leaves in the early stages of the disease. The fungus grows downward through the petiole and into the cambial region of the stem, and the seedling dies. A *Phytophthora*, tentatively identified as *P. parasitica* Dast., was isolated from Peruvian seedling, leaf, petiole, and stem material. Inoculation studies indicated that infection may take place through wounds on the stem or on uninjured bud tips or mature leaves. In Peru some control was obtained by removing the diseased plants from the beds. Applying a copper spray with sticker at regular intervals was suggested as a possible control.

Phytophthora parasitica Dast.—“Maculis amphigenis, orbicularibus, centro areolatis, dilute umbrinis, annulatis, solitariis vel confluentibus; mycelio intermatricali, ex hyphis inter- et intracellularibus, primo continuis, tandem septatis, 3–9 μ crassis, constantibus; haustoriis sparsis, digitatis vel subglobosis, raro ramosis, sporangiophoris 100–300 μ longis, sporangis terminalibus, interdum intercalaribus vel lateralibus, plerumque ovoideis, subinde globosis 25–50 \times 20–40 μ ; zoosporis 8–12 \times 5–8 μ ; sporis perdurantibus globosis, flavidis, 20–60 μ , membrana crassa, levi; oogoniis et antheridiis in vitro cultis; oogoniis intercalaribus vel lateralibus, globosis, levibus vel rugosis, melleis, 18–25 μ (23.8 μ), pedicellis per antheridia penetrantibus; oosporis globosis, 15–20 μ (18.6 μ), membrana crassa, mellea, levi.” (27%, p. 168.)

(See *Phytophthora parasitica*, p. 33.)

A species of *Phytophthora* is said to cause severe damage in some instances as a postemergence damping-off organism in Guatemala.⁹ The diseased plants are sprayed with bordeaux or Yellow Cuprocide for control.⁹

In 1945 Pinkus (206) reported that a species of *Phytophthora* has been identified by A. S. Müller as the cause of a disease affecting grafted cinchona nursery stock in Guatemala. The loss from this disease was said to be 3.75 percent in 1943, and the disease was of comparable prevalence in 1944. Small coffee-colored spots occur on the leaves, and the infection spreads downward through the midrib to the main stem, eventually killing the grafts. Nurseries exposed to the morning sun and on good soil sustain less damage than those less favorably situated. Marked variations in susceptibility of individual clones have been noted. Control has been effected by removing and burning diseased foliage bi-weekly during the rainy season and weekly during the dry season.

Reinking⁹ described a dieback and canker disease affecting *C. ledgeriana* and *C. calisaya* nursery stock in Guatemala. The young growing tips of affected plants wilt and darken, and the disease may move down the shoots and eventually kill the seedlings.

⁹ See footnote 5, p. 4.

Frequently, the graft region is infected. Often when plants are attacked at the bases of leaf petioles, sunken stem cankers are produced that may girdle and kill the upper parts of the plants. Reinking stated that Dr. Albert Müller isolated a species of *Phytophthora* from diseased plants and attributes the disease to this fungus. Control is attained by good sanitary and cultural practices, pruning diseased parts well below point of infection and spraying the most severely attacked plants with bordeaux mixture, burning diseased plants and prunings, watering nursery by flooding rather than spraying, and spacing plants in nursery to allow good aeration.

(See phytophthora cankers, p. 32, and phytophthora root rots, p. 40.)

STEM CANKERS

Olpidiaceae

In 1910 Commelin (43) described a disease that affected the collar region of *Cinchona succirubra* seedlings in Java. Microscopic examination of sections through the affected region showed the presence of a fungus belonging to the Olpidiaceae of the Chytridiales (43). The affected plants were small, having a stem diameter of about 2 mm., and were growing at a high elevation (43). The bark had dried at the soil line for a distance of 1 to 2 or more centimeters, but the roots remained in a healthy condition (43). In the early stages of attack, i. e., before the stem was completely girdled, the leaves were green (43). In the later stages of the disease, however, the leaves faded and withered and the plant tissues turned brown (43). Corky tissues occurred occasionally in the diseased bark (43). Adventitious roots frequently formed on the stem above the diseased parts (43). The sporangia of the Chytrid measured about 0.02 mm. in diameter and were found in various stages of development in the cells of the bark and woody tissues (43). Blücher (13) suggested that the fungus probably gained entrance through hoe wounds. Leersum (170) stated that experiments in controlling the disease by cutting out the diseased spots and treating the wounds with carbolineum, slaked lime, or chloride of lime produced no results in the majority of cases. The same or a similar Chytrid was found in the dead areas of the outer bark of cinchona trees affected with "trunk canker" (43, 227).

Physiological

Owen (198) reported a physiological canker affecting the collar region of small cinchona plants in nursery beds in India. The disease was not found generally in the seedbeds but occurred in nursery beds situated above the mist line in damp locations. He suggested placing the beds at lower elevations and putting in a better drainage system.

Undetermined

In 1904 Zimmermann (289) reported very severe damage to *Cinchona succirubra* and *C. ledgeriana* hybrid seedlings at Amani,

German East Africa (Tanganyika since 1919). The leaves of the affected plants wilted and became discolored, and the stems usually turned brown near the root collar, while the root systems remained in a healthy condition (289). Sometimes only the upper part of the stems turned brown and then only the higher leaves withered and became discolored (289). Occasionally the leaves alone showed brown discolorations, principally along the midrib (289). A fungus mycelium having characteristically lobed haustoria was observed in microscopic sections of diseased leaf and stem material; however, fructifications of this fungus were not observed (289). The disease diminished considerably with the advent of dry weather, and transplanting the seedlings to the plantation brought the disease to a standstill (289). Control measures were unnecessary at the time (289). Zimmermann (289) isolated a number of fungi that developed on parts of the diseased stems when left in damp soil for a time and from these he described the following new species: *Calosphaeria cinchonae* Zimm., *Nectria amaniensis* Zimm., *N. cinchonae* Zimm., and *Pestalotzia cinchonae* Zimm. *Nectria coffeicola* Zimm. was also isolated from the diseased stems (289). Inoculations with these fungi failed to produce the disease (192, 289). In 1906 Koorders (154) stated that, of the fungi described by Zimmermann, *Calosphaeria cinchonae*, *Nectria cinchonae*, and *Pestalotzia cinchonae* had not been reported in Java.

In the report of the Government cinchona plantation at Tjindiroean, Java, for 1917 (56, 104), a disease of unknown cause, which is said to correspond in every way with the collar disease described by Zimmermann, occurred in several seedbeds where a great deal of sprinkling had been done. The disease noticeably appeared in spots where the young plants stood very close together (56).

ROOT ROTS

Rhizoctonia

In 1940 a root rot was reported on *Cinchona ledgeriana* plants in the nurseries at Mayaguez, Castañer, El Semil, and Las Mesas, P. R., and also to some extent on some plants at Maricao (216). The fungus responsible for the disease has been identified as a species of *Rhizoctonia* (216).

Heubel (127) reported a root collar disease in the seedbeds in South Sumatra caused by *Rhizoctonia*, the fungus also causing the "mopo" disease.

(See rhizoctonia damping-off, p. 5.)

Sclerotium

Sclerotium rolfsii Sacc. has been reported by Rhind (235) on cinchona seedlings in the nursery of the Government cinchona plantation in the Tenasserim District, Burma, and by Crandall¹⁰

¹⁰ CRANDALL, B. S. NURSERY ROOT AND COLLAR ROT OF CINCHONA AND ROOT ROT OF CANAVALLIA CAUSED BY SCLEROTIUM ROLFSII. U. S. Bur. Plant Indus., Soils, and Agr. Engin., Plant Dis. Rptr. 29: 536-537. 1945. [Processed.]

on *Ledgeriana* transplants at the agricultural experiment station, Tingo Maria, Peru. Crandall observed the disease in isolated patches in the nursery following the rainy season, where it caused a root and collar rot. He isolated the fungus from diseased collar tissue and found the sclerotia of the fungus in the soil within which diseased plants were growing.¹¹ Eradication of diseased and adjacent healthy plants and the removal of a part of the soil in the bed are said to have checked the spread of the disease in Peru.¹¹ The extent of damage was not serious in Burma (235). Weber (281) listed *Cinchona* sp. as one of the hosts of *S. rolfsii*.

Sclerotium rolfsii Sacc.—“Superficiale, subrotundum v. horizontaliter ellipsoideum, 0.5–0.8 mm. diam., facile secedens, levigatum, nitidulum, roseum, demum fulvescenti-brunneum, carnosum-firmulum, intus pallidum; cellulis e globoso polyedricis 6–8 μ diam. interdum sinuosis, subhyalinis, ad peripheriam sclerotii brunneolis; fructificatione nulla observata.” (243, p. 257.)

Sporodesmium

In 1906 and 1907 Koorders (154, 155) reported a new species, *Sporodesmium cinchonae* Koord., as a wound parasite on 1- to 2-year-old cinchona hybrid seedlings in the Kedoe Residency in Middle Java. Most of the seedlings had been rather severely damaged by the nematode *Heterodera marioni* (Cornu) Goodey; but nematode-free seedlings, which were from a different plantation and were suffering from a root disease, were found to be infected with the same species of *Sporodesmium* (155). Koorders (154) also isolated *Sporodesmium* from diseased plants sent from the Government cinchona plantation in the Preanger; therefore, he considered the nursery bed root disease in the Preanger to be identical with the disease in the Kedoe.

Leersum (162), who had reported the disease in the Preanger in 1891, stated that *Cinchona ledgeriana* seedlings quickly succumbed to the disease, while *C. succirubra* plants were not so quickly killed by it. The bark of young plants cracked at the root collar (167). The tops of the plants appeared healthy for a while after the roots were found to be seriously infected; later the leaves turned red, and the plants withered and died (162). Leersum (162) considered the white mycelium that he found between the root bark and wood to be characteristic of the disease. The wood of the infected roots was found to be brown and the bark rotten (162). In 1903 there was considerable disease in nurseries on soil rather impermeable to water; 50,000 plants were lost in old nursery beds at Kawah-Tjiwidei, while the newly laid-out nursery beds remained free of the disease (2, 154). Using new nursery beds composed of rich humus, building up the beds for drainage, and transplanting to field positions when the plants are 2 years old were measures suggested for control (154).

Sporodesmium cinchonae Koord.—“Parasiticum, conidiis acrogenis sub-solitaris, in radicis cellulis inordinate dispositis, primo juvenute hyalinis vel

¹¹ CRANDALL, B. S. NURSERY ROOT AND COLLAR ROT OF CINCHONA AND ROOT ROT OF CANAVALLA CAUSED BY SCLEROTIUM ROLFSII. U. S. Bur. Plant Indus., Soils, and Agr. Engin., Plant Dis. Rptr. 29: 536–537. 1945. [Processed.]

pallide olivaceis v. fuscis, continuis, dein aterrimis, oblongis v. irregularibus, plerumque $30-45 \times 14-21\mu$ usque ad $50-65 \times 15-18\mu$, multicellularibus." (155, pp. 234-235; also described in 227 and in 242, v. 22, p. 1402.)

Stigeosporium

In March 1941 Marañon and Bartlett (183) reported on a canker disease of cinchona in the Philippine Islands from information and data given in an unpublished manuscript by Servando S. Madarang. All of the cinchona species grown at Impalutao and Kaatoan, namely, *Cinchona ledgeriana*, *C. succirubra*, *C. officinalis*, and *C. hybrida*, were infected. The bark canker is said to have appeared in such epidemic proportions between 1929 and 1934 as to threaten to destroy the plantation at Impalutao. From July to December 1939, 22.4 percent of the 1,916,639 plants raised in the Kaatoan nursery were eradicated because they showed the canker symptoms. In the first 10 months of 1940, 62 percent of the 1,067,592 nursery plants were discarded from the beds and of this group 94 percent were cankered. The disease occurs as a leaf and stem blight, root necrosis, and a canker of seedlings and nursery plants. "The root necrosis is said to be the initial stage of infection, which later manifests itself as seedling stem and leaf blight and finally as tree canker." (183, p. 125.) Illustrations of diseased older trees show the canker near the soil line.

The causal organism of this disease complex is said to be an undescribed soil-borne fungus and was to have been published as *Stigeosporium cinchonae* Roldan n. sp. Infected plants up to 3 years are grubbed out and destroyed. Madarang advised leaving older trees, if not too badly infected, until at least 6 years of age, when they can be harvested. After diseased branches are pruned, the wounds are painted with coal tar. Resistant trees are to be used in future breeding and selection work.

NEMATODE ROOT INFECTIONS

Heterodera

Plant-parasitic nematodes (eelworms, roundworms, or threadworms) are minute organisms of complex structure. The most widely known species, *Heterodera marioni* (Cornu) Goodey, the root knot nematode, has been reported on seedlings of *Cinchona* spp. in southern India (8, 87, 133, 156, 191) and cinchona hybrids in Java (12, 100, 154, 155, 184, 230).

Koorders (154) found large numbers of this nematode in the roots of cinchona seedlings that were diseased and stunted in growth. Galls were not numerous and only 0.5 to 1 mm. in size at the most (154). Swollen females were frequently found in the bark of roots that showed no external evidence of their presence (154). They were present in both the dead rotten bark and the living bark of the cinchona roots (154). Koorders (154) isolated a species of the fungus *Sporodesmium* from the roots of most of the infected plants as well as from the roots of diseased plants that were not infected by nematodes, and he was not able to state conclusively which of the organisms involved had caused the

injury to the seedlings. Rant (226) reported in 1913 that *Cinchona ledgeriana* plants had been successfully inoculated with the nematode *Heterodera marioni*. The extent of damage done to cinchona in both India and Java is said to be unimportant (191, 230).

Dorylaimus

In Java Koorders (154) observed another species of nematode in the rotten bark of a cinchona seedling infected with *Heterodera marioni* and the fungus *Sporodesmium*. He tentatively identified the nematode as *Dorylaimus* sp.

Tylenchorhynchus

Tylenchus alatus Cobb was described originally from root specimens, said to have been *Cinchona* sp., from Africa (42). Goodey (87) stated that it was a root of *C. succirubra* Pav. from Belgian Congo. *Tylenchus alatus* Cobb was placed by Filipjev into the genus *Tylenchorhynchus*, and therefore it is called *Tylenchorhynchus alatus* (Cobb) Filipjev (79).

Tylenchus

In 1941 Fluiter and Mulholland (80) reported *Cinchona succirubra* as a host of the nematode *Tylenchus coffeae* Zimm. in Java.

PLANTATION DISEASES

LEAF SPOTS

Cephaleuros and other algae

An algal leaf spot, caused by *Cephaleuros virescens* O. Kunze, has been reported by Hendrickx (122) on *Cinchona* spp. in the Mulungu region, Belgian Congo; by Reinking¹² on cinchona in Guatemala; and by Crandall and Davis¹³ on *C. succirubra* in Guatemala, on *C. pubescens* in Colombia, and on the Ledgeriana form of *C. officinalis* in Guatemala and Peru. The circular and raised spots are light reddish brown with some light-green coloring and occur mostly on the upper leaf surfaces.¹² In Guatemala the disease is usually found at higher altitudes where the humidity is high.¹² The algal leaf spot in the Mulungu region was not serious enough to warrant a program of control (122), nor was it considered to be of economic importance in the Americas.^{12 13}

Cephaleuros virescens O. Kunze.—Thallus a roundish disk, more or less notched at the edge, generally uninterruptedly complete, red yellow or red brown, up to 1 cm. broad, 1- to many-layered, subcuticular or subepidermal, composed of radial dichotomously branched, radiate, closely connected filaments attached to the leaf by a moderate number of short, 1- to 3-celled rhizoids; cells brown-walled, polygonal-rounded, varying between 4 μ to 12 μ in diameter, 1 to 3 times longer; hairs absent in young thalli, sterile and fertile hairs abundant on the older ones and forming a furry covering that

¹² See footnote 5, p. 4.

¹³ CRANDALL, B. S., and DAVIS, W. C. CEPHALEUROS VIRESCENS ON CINCHONA IN CENTRAL AND SOUTH AMERICA. U. S. Bur. Plant Indus., Soils, and Agr. Engin., Plant Dis. Rptr. 28: 926. 1944. [Processed.]

penetrates the cuticle; sterile hairs simple, stiff, consisting of several cells, the apical cell pointed; fertile filaments branched, capitately swollen, bearing 4 to 8 lateral or terminal clustered sporangia with 1 oval or ellipsoidal sporangium mother cell, 25μ to $30\mu \times 38\mu$ to 40μ , appearing particularly during the rainy period; zoospores biciliate, elongated ovoid, 5μ to 8μ long; globular sporangia occurring in the thallus, chiefly near the edge or terminal on the creeping filaments, borne on the same thallus as the cluster sporangia, oval-ellipsoid, of the same size as the mother cell of the cluster sporangia. The thalli can live and grow on the leaves for years (213).

Martin and Crandall (186) reported a green algal leaf spot on *Ledgeriana cinchona* trees in Peru.

Hunger (132) reported that specimens of *C. ledgeriana* leaves, which were preserved in the station museum at Malang, Java, showed injury caused by an alga belonging to the Chroolepidaceae.

In 1944 Martin and Crandall (186) observed a brown algal leaf spot on a cinchona tree in Peru.

Cercospora

In 1887 Ellis and Everhart (74) described a new species of *Cercospora* collected on cultivated cinchona in the United States of America. Hansford (113, 115, 116) also reported *C. cinchonae* on cinchona in Uganda. This species of *Cercospora* was listed by Lieneman (173).

Cercospora cinchonae Ell. and Ev.—“Spots amphigenous, definite, nearly black above, with a narrow, slightly raised margin, brownish-black below, 2-3 millim. in diam.; hyphae epiphyllous in small, scattered, sphaeriaeform tufts, very short; conidia cylindrical, yellowish-hyaline, nearly straight, becoming faintly 3-septate, $25-35 \times 2\frac{1}{2}\mu$.” (74, p. 17; also described in 227 and in 242, v. 10, p. 645.)

Gloeosporium

An undetermined species of *Gloeosporium* was observed by Koorders (154) in 1905 on mature leaves of a 2-year-old cinchona hybrid in Middle Java. The characteristics of the fungus are said to correspond to some extent with the genus *Zythia* Fr. (*Phomopsis* Sacc.).

Gloeosporium sp.—Sori round or irregular; spots large, straw-colored, on both the upper and under sides of mature leaves, about 120μ in diameter; spores oblong, straight or slightly bent, 1-celled, hyaline, without appendages, 8μ to $10\mu \times 3\frac{1}{4}\mu$ (154; also described in 227).

An undetermined species of *Gloeosporium* was isolated from brown circular spots on cinchona leaves in the Darjeeling District of Bengal, India (84, 136, 199).

(See gloeosporium leaf spot, p. 9.)

Phyllosticta

In 1892 Patouillard and Lagerheim (201) described a fungus causing a leaf spot on *Cinchona* sp., to which fungus the name *Phyllosticta cinchonae* was assigned. It was collected at San Nicolas during a trip to the coastal and mountainous regions of Ecuador (201). The fungus causes angular, dull-yellow spots on the living leaves (259). *P. cinchonae* has been reported to cause brown leaf spots on the leaves of *C. succirubra* in the Transcaucasian region of Russia (U.S.S.R.) (210).

Phyllosticta cinchonae Pat.—“P. maculis exaridis, sparsis vel confluentibus, 4–8 millim. latis, quadrangularibus aut irregulariter 5–6-gonis, ochraceis, augusti brunneo-marginatis; peritheciis epiphyllis, 80–100 μ latis, brunneis; sporulis hyalinis, ovoideis, biguttulatis, $7 \times 2\mu$.” (201, p. 135; also described in 242, v. 11, p. 473.)

Koorders (155) also described a species of *Phyllosticta* under the name *P. cinchonae*. His material was obtained from the leaves of young high-percentage cinchona hybrids in Middle Java at an altitude of 1,600 meters (155). The irregular leaf spots were straw-colored, not bordered, often 0.5 cm. in size, appearing both on the upper and lower surfaces of the leaves (155). The fungus was said to cause little damage at the time and had attacked only a few plants (155). Because of the confusion with Patouillard's fungus, Rant (227) renamed the species *P. cinchonicola*. Later, Stevenson (259) stated that the name *Phyllosticta cinchonae* Koord. is untenable and that *P. cinchonae* Pat. should have priority.

Phyllosticta cinchonicola Rant (*P. cinchonae* Koord. non Pat.).—Pycnidia scattered, straw-colored, thin-walled, flattened globose or ovoid, about 120 μ in diameter; conidia cylindrical, hyaline, 1-celled, oblong, straight or nearly straight, rounded at both ends, 8 μ to 10 $\mu \times 3.25\mu$, generally 10 μ long, not infrequently containing a few large drops of oil, sometimes broader at the tip than the base (227; also described in 155 and in 242, v. 22, p. 846.)

(See *Phyllostictina*, p. 10, and *Phyllosticta* spp., p. 11.)

Prillieuxina

Stevenson (260) reported a species of *Prillieuxina* occurring on living leaves of *Cinchona pubescens* Vahl in Costa Rica. The fungus caused definite leaf spots, and infected leaves turned red and fell prematurely. The diseased tree was standing in a forest near a commercial planting of cinchona.

Prillieuxina cinchonae Stevenson.—“Hypophylla, plagulas 1 cm. latas formans; mycelio anastomosanti 3–3.5 μ crasso; hyphopodiis nullis; thyriotheciis hemisphaerico-lenticularibus, 150–300 μ diam.; ascis subglobosis usque ovatis, aparaphysatis, 30–45 \times 25–35 μ , octosporis; sporis medio 1-septatis et constrictis, fulvis, 24–27 \times 12–15 μ .” (260, p. 632.)

Sclerotinia

In 1904 Lutz (176) reported a leaf disease caused by *Sclerotinia fuckeliana* D By. on cinchona plants cultivated in the greenhouses of the Paris School of Pharmacy, France. Small, semitransparent spots appeared first on the leaves, and these spots turned brown in 1 or 2 days (176). The causal organism continued to spread from the original spots in concentric zones until the entire leaf blade was covered in 5 or 6 days (176). The leaves withered, shriveled, and finally fell from the plants (176). A light-gray mycelial growth appeared on the leaf spots on either the upper or lower sides of the leaves, which, when isolated and cultured, produced the conidia of *S. fuckeliana* (176). Healthy plants that were inoculated with the disease organism showed the leaf spots after 2 days and necrosis followed in 24 hours (176). Control consisted of the removal of seriously affected leaves, repeated spray-

ings with neutral bordeaux mixture, and repotting the plants (176).

Rant (227) stated in his 1914 publication that the disease had not been reported from Java. While investigating the mopog disease of cinchona in Java, Rant (229) inoculated leaves of 4- to 6-inch cinchona hybrid seedlings with *Botrytis cinerea* and obtained leaf spots similar to the spots caused by *S. fuckeliana* as described by Lutz. There was also abundant spore production (229). Rant's further tests, however, indicated that *B. cinerea* was able to produce the leaf symptoms described by Lutz only at the points of inoculation (229).

Sclerotinia fuckeliana D By.—"Stipitata, minuta, patellaris, $\frac{1}{2}$ –3 mm. lata, 5–10 mm. alta, flavo-brunnea ex *Sclerotio echinato* Fuck. nascens; ascis teretivatis, 130×12 –13; sporidiis monostichis, ovoideis, hyalinis, 10 – 11×6 –7." (242, v. 8, p. 196.)

Uredo

In 1902 Hennings (125) described a new species of *Uredo* collected by Zimmermann on *Cinchona* sp. growing in an experimental garden in Java. The fungus formed brown-rust-colored pustules on the leaves (259).

Uredo cinchonae P. Henn.—"Maculis fusciculis vel obsoletis, soris epiphyllis, sparsis, minutissimis, pulvinatis, diutius tectis, brunneolis, ca. 0.3 mm. diametro; sporis ovoideis vel ellipsoideis, intus flavidis, 13 – 18×10 – 15μ episporio hyalino, granulato-verrucoso." (125, p. 140; also described in 227 and in 242, v. 17, p. 439.)

In 1924 P. and H. Sydow (266) stated that Hennings' species is not valid, because examination of the original material showed no fungus formation in the light-brown emarginate spots and the "spores" described by Hennings seemed to be the cells of the matrix of the host leaves.

Undetermined

Reinking¹⁴ has reported a leaf spot of undetermined cause affecting *Cinchona ledgeriana*, especially the narrow-leaved strains, and to a less extent *C. succirubra*, *C. calisaya*, and hybrids on certain plantations in Guatemala. The disease is characterized in the earliest stage by minute water-soaked spots, which later have dark red-brown borders and sunken centers that turn ash gray eventually. The spotting begins at the leaf tip and edge and moves inward. Severely attacked young leaves are wrinkled, and the margins are curled under. Defoliation may occur, although apparently normal trees may have severe spotting without defoliation. The disease was observed on *Ledgeriana* seedlings in seedbeds and on grafted *Ledgeriana* plants grown in plots with and without coffee plant shade. Specimens examined for a causal organism failed to show a fungus or bacterium, and it is suggested that the disease might be of nutritional origin.

¹⁴ See footnote 5, p. 4.

OTHER LEAF DISEASES

Capnodium and other molds

Sooty mold, which is found on a number of tropical and subtropical plants, has been reported on the leaves and stems of *Cinchona ledgeriana* in Java (167, 220, 227) and on *Cinchona* spp. in Indochina (24), Uganda (250), and Guatemala.¹⁵ The characteristic black sooty covering on the affected parts is made up of the hyphae of fungi that live on the "honeydew" excreted by scale and other insects (24, 220). Although a large number of the fungi causing sooty mold belong mainly to the Perisporiaceae of the Ascomycetes (19), *Capnodium brasiliensis* Putt. was the organism reported from Indochina (24) and Uganda (250). Heavy growth of the mold may prevent the proper functioning of the covered leaf,¹⁵ but damage to the host plant is caused primarily by the feeding of the insects (227)¹⁵ and is comparatively unimportant on cinchona (24, 220). Control consists of the destruction of the insects (24, 167).

(See hormiscium sooty mold, p. 27.)

Mineral deficiencies

Two distinct types of chlorophyll deficiency are reported on some cinchona trees at Maricao and Las Mesas, P. R. (117). One type is said to be caused by iron deficiency. In the other type the leaves develop asymmetrically, causing the midrib to be curved. A light-pink color predominates over the natural green in some parts of these leaves, and the leaves vary widely in shape and size. The causes of these disturbances were not determined.

Reinking¹⁵ reported a leaf mottle or chlorosis on cinchona in Guatemala, believed to be a nutritional trouble. The yellow-green mottling usually occurred between the main lateral veins.

Harper (117) in 1943 reported an abnormality of the leaves of some cinchona trees at Maricao and Las Mesas, P. R., probably caused by nutrient deficiencies or by the mineral toxicity of the soil. The young leaves stop growing at the margins, with the result that mature leaves have an inverted-cup shape and are wrinkled, with dark-red mottling. Leaves of the fruiting branches are smaller, thicker, and flatter than other leaves. Generally, sterile flowers develop with only miniature corollas.

Mosaic

In 1909 a mosaic disease was reported to occur occasionally on otherwise normal *Cinchona ledgeriana* plants in the Java plantations (167). One entirely variegated plant of *C. succirubra* was found. It was stated that this symptom is not to be confused with variegation of the edges, which is probably an indication of weakness.

In 1931 Petri (204) reported that the leaves of *Cinchona* sp. at Faghena, Eritrea, were affected with mosaic.

¹⁵ See footnote 5, p. 4.

Pellicularia and other thread blights

Davis and Crandall¹⁶ reported a thread blight caused by *Corticium koleroga* (Cke.) Hoehn. (= *Pellicularia koleroga* Cke.) on 10-year-old trees of *Ledgeriana* stock in Guatemala. The mycelial threads of the fungus extended as a web over the trunk, branches, and leaves, and the dead leaves hung in clusters, suspended from the branches by the fungal threads.¹⁶

Horse hair blight (*Marasmius* sp.) was observed by Davis and Crandall¹⁶ on a few wild trees of *Cinchona barbacoensis* Karst. in Colombia. The fungus strands were found on trunks, branches, and leaves.¹⁶

Hunger (131) reported a "cobweb" disease on the twigs and leaves of *C. ledgeriana* in the Soekaboemi area in western Java. White fungus threads crept along the branches and forked at the side branches, twigs, and leaves (131). The threads grew over the leaf petioles and split into a many-branched system of fine threads on the leaf blade to form the "cobweb" (131). The leaves died and became detached from the twig but hung from it by the fungus thread (131). The disease is said to occur quite infrequently in the cinchona plantations in Java, and cutting and burning the affected branches is recommended as a means of control (131, 227).

Rant (222, 227) reported another thread blight that he found on *C. succirubra* and on a young cinchona hybrid planted in a damp shady spot. The mycelial threads were honey yellow to brown and crept along the trunk for more than 120 cm. (222). The threads were 1 to 1.5 mm. wide and did no damage to the tissues of the plant until the leaf blades were reached (222, 227). The leaf blades died finally and hung from the twigs by the fungus filaments (222). Rant found a white to brownish powdery fruiting stage on the under surface of some of the leaves, but he did not observe a "cobweb" formation (222, 227).

Scab

Jenkins (138) reported a scab caused by a species of *Elsinoë* affecting the leaves, stems, and fruits of cinchona in South America. The disease occurs on *Cinchona pubescens* Vahl and rarely on *C. officinalis* L. in Colombia and Peru and rarely on *C. delessertiana* Standley in Peru. The fungus causes spots on the upper or lower surfaces of the leaves, and occasionally the spots fall out, leaving perforations or tissue network. Raised cankers are formed on the young stems and rachis and branches of the inflorescence. Diseased fruit capsules are frequently curved or otherwise bent and distorted and may also be dwarfed. Lesions on the capsules are often numerous and usually more conspicuous than on leaves and stems. The possible economic importance is not known as yet; but it is suggested that, since the leaves and young stems

¹⁶ DAVIS, W. C., and CRANDALL, B. S. SOME CINCHONA DISEASES IN THE WESTERN HEMISPHERE. U. S. Bur. Plant Indus., Soils, and Agr. Engin., Plant Dis. Rptr. 28: 996-997. 1944. [Processed.]

are affected, it might prove destructive under favorable conditions in the nursery. Jenkins described the causal fungus under the name *Elsinoë cinchonae* Jenkins.

Elsinoë cinchonae Jenkins.—“Maculae plerumque numerosissimae, conspersae, circulares, subcirculares, usque ellipticae, interdum elongatae, elevatae, centro saepe plus minusve apiculiformi, interdum aggregatae vel confluentes, in foliis amphigenae, interdum nervisequentibus, usque 1.5 mm, rare 2 mm, superne conspicuiores et saepe cinnamonea-griseae, margine nigro-vinaceo-brunneo circumdatae, inferne vinaceo-brunneae; cancri in caulibus generaliter elliptici, usque 4×5 mm, avellanei, vel discolores, in capsulis usque 3 mm diameter, avellanei vel discolores; ascomata plus minusve numerosa, in maculis foliorum epigena conspicuoria, rotunda usque elliptica, pulvinata, exposita, usque 300μ diameter, 75μ crassa, superficialiter nigro-brunnea; epithecium fuscum, 10μ crassum; asci numerosi, sub-epithecio in regione stromatica hyaline distributi, globosi usque ellipsoidei, apice incrassati, $18-20\mu$ diameter; ascospores immaturae, 1-3-septatae, hyalinae, $15 \times 15\mu$; status conidiophorus (*Sphaceloma*) in maculis foliorum epigenus prominens; conidiophora in palum compactum, expositum superficialiter nigro-brunneum, plus minusve continuum, ex stromate hyaline oriundum, fructificatione tota 30μ crassa, vel marginem maculae versus usque 50μ ; conidiophora cylindrica apice acuminato, generaliter continua vel uniseptata, $3.5-5 \times 5-15\mu$; conidia rare visa, brunnea, elliptica, $4-5 \times 8-10\mu$.” (138, pp. 348, 350.)

Undetermined

Owen (198) reported a disease on cinchona leaves in Ballanagodde, India, which is controlled by an application of lime and sulfur.

In 1909 an abnormal leaf condition of otherwise healthy cinchona trees was reported in Java (167). Corky growths were formed in spots on the young leaves; and as these spots were unable to keep pace with the rapid growth of the surrounding healthy parts of the leaf blades, the tissues tore around the spots. Frequently the corky material dropped out, leaving holes in the blades.

In 1942 Kevorkian (151) reported a virus disease of cinchona in Puerto Rico.

BARK DISEASES

Hormiscium

In 1888 Fürth (82) and Gorkom (95) reported the occurrence of a black sooty fungus on the trunks and branches of cinchona, some of which were *Cinchona succirubra*, in a few plantations in Java. The spots were jet black and soft in the earlier stages, drying out and becoming firmly attached to the bark later (82), and thereby making the bark unfit for pharmaceutical use (95). Gorkom (95) stated that the fungus belonged to the genus *Hormiscium*, to which he gave the specific name *pannosum* without publishing a description.

The fungus multiplies by conidia and spreads very rapidly (95). It penetrates the outer dead “cork cells” but does not enter the living tissues; however, Gorkom believed that the thick fungus growth interfered with the normal functions of the bark, in addition to making the bark unfit for use (95). Very crowded nurseries, high humidity, and shade were thought to be favorable to the growth of the fungus (95).

Rant (227) stated that he observed a species of *Hormiscium*, which he thought was probably *H. pannosum*, on living, sound trunks of old *C. succirubra* trees at the Government cinchona plantation in Java. He described the fungus as a saprophyte, forming rather thick, black, irregular coverings, which varied in thickness and which bore dark olive-green conidia having a diameter of 6μ to 17.5μ and underdeveloped conidiophores (227). Gorkom (95) recommended removing the fungus growth by mechanical means and burning and washing the infected spots with milk of lime or limewater. He advised against the use of copper sulfate or carbon bisulfide solutions on the spots, fearing that the poisonous component might affect the bark unfavorably (95).

(See capnodium sooty mold, p. 25.)

Septobasidium

The sporadic occurrence of species of *Septobasidium* in the cinchona plantations in the Netherlands Indies has been reported for a period of about 35 years (131, 160, 167, and others). According to the classification by Couch (46) there are three species of *Septobasidium* that have been reported on cinchona, namely *S. bogoriense* Pat., *S. cinchonae* Rac., and *S. lichenicolum* (Berk. and Br.) Petch. *S. bogoriense*, the gray dadap fungus, has been reported on *Cinchona* spp. in Java and Sumatra (11, 16, 17, 46, 66, 67, 85, 86, 88, 121, 157, 236, 252-254); *S. cinchonae*, on young branches of *Cinchona* sp. in Java (16, 46, 218, 227, 253); and *S. lichenicolum*, on *Cinchona* sp. in Java (17, 46). Attacks of *S. bogoriense* are said to be especially heavy on young cinchona grafts and seedlings after heavy blooming (67). *S. cinchonae* has been found on cinchona at elevations of 2,300 to 2,700 meters above sea level and occurs more frequently in young nurseries where the plants are low and produce leaves close to the ground than in older plantations where the trees are widely spaced and the circulation of the air takes place more freely (16). While the *Septobasidium* species on cinchona are generally considered to be epiphytic, they are capable of causing indirect damage to the host plants by providing a damp atmosphere and shelter for other fungus organisms, such as the disease-producing *Corticium salmonicolor* Berk. and Br., and by harboring the scale insects associated with *Septobasidium*, which live at the expense of the host plant (11, 211, 253). Although the plants recover from attack, the death of a large number of branches in the plantation represents a considerable crop loss at the time of harvesting (11, 236). Steinmann (253) advised spraying for the control of scale insects, the insecticide used depending on the type of insect present.

Tulasnella

Raciborski (218) reported a species of *Tulasnella* as being common on young branches of *Cinchona ledgeriana*, *C. officinalis*, and *C. succirubra* in the Prèanger, Java. Although the fungus

hyphae grow only on the surface, the young bark underneath the mycelium is killed and later the tips of such affected branches die even though the fungus has not extended on to them (218). Stevenson (259) reported the fungus as occurring in thin layers on lower leaf surfaces. Leaves that are infected are promptly killed (218). *T. cinchonae* is said to spread in the plantations and to be injurious (218, 227).

Tulasnella cinchonae Rac.—“Effusa, applanata, tenuissima, caules et paginam inf. foliorum obducens, pallide rosea; hyphis hyalinis septatis usque ad 60μ long., depressis sursum copiose ramulosis; basidiis in apice ramulorum nascentibus, anguste ovoideis, $20-24 \times 8-11$, apice rotundatis; sporis nunc apice nunc etiam lateraliter ad basidia quaternis globosis v. breviter ovoideis, hyalinis, tenuiter tunicatis, $7-9\mu$ diam., non secedentibus.” (242, v. 21, p. 452; also described in 218.)

Rogers (238) examined the type specimens from Raciborski's herbarium and stated that he did not believe that the fungus described was a species of *Tulasnella*, being somewhat like a species of *Septobasidium*.

Undetermined

Martin and Crandall (186) reported a few cases of rough bark on *Ledgeriana* trees in Peru. The condition was said to be so rare at the time as not to be alarming.

Owens (198) reported that “hide-bound” trees are sometimes found in the cinchona plantations in India. The condition is not infrequently mistaken for canker. A vertical slit in the bark relieves the condition.

SYSTEMIC DISEASES

Undetermined

In 1939 Hendrickx (122) reported a tracheomycosis disease at Kalonge, Belgian Congo, on *Cinchona ledgeriana* and *C. succirubra*. Stoffels (262) stated that the disease is not limited to the wood, but also causes a bark necrosis. The trees succumb quickly (122). A species of *Fusarium* of the section “Elegans” and a species of *Verticillium* have been isolated and inoculations made (135).

A disease affecting *C. robusta* and occasionally *C. succirubra* has been reported in Java, in which large trees begin to die about a meter from the ground, with the necrosis continuing upward, while the lower part of the tree remains alive and can send out new shoots (167). About 30 trees were found to be affected by the disease in one locality. Inoculation tests with fungi isolated from the diseased trees were negative.

TRUNK AND BRANCH CANKERS

The word “canker” has been used so variously in connection with cinchona diseases that it is difficult to judge from the literature the actual number of different canker diseases involved. Among the earlier workers in the Netherlands Indies who wrote about the canker diseases of cinchona were Eekhout (73), Kessler (146), Warburg (279, 280), and Wurth (as reported by Hunger, 131). Since their discussions are rather general, however, and

do not contribute greatly toward an accurate understanding of the symptoms and causal organisms involved in the various canker diseases, they will not be reviewed at length here. A similar general treatment is found likewise in some of the early reports by the Government cinchona station in Java, where the terms "stem canker" and "stem rust" were used interchangeably to designate one disease (54-59), but which were later separately used for two diseases (60-62, 64, 65, 67, 70). Numerous other reports have been made of the occurrence of stem, trunk, or branch canker in Java and Sumatra (2, 88-92, 98, 100, 106-109, 112, 129, 131, 150, 154, 157-161, 167, 168, 170, 214, 227, 240, 252, 263). Zimmermann (290) stated that canker is the most common disease of cinchona. Some of the fungi observed on or believed to be connected with cinchona canker diseases are *Botryodiplodia* (161), *Nectria* (154, 214, 227), *Peziza* (279), and *Tubercularia* (227).

Corticium

Pink disease, called "djamoer oepas" in Java, is caused by *Corticium salmonicolor* Berk. and Br. (synonymous with *C. javanicum* Zimm.) (227). It occurs on more than 140 different species of plants in the tropical rain-forest regions of the world (249), including such economic plants as citrus species, coffee, tea, rubber, cacao, cinnamon, and green-manure plants (225). Butler (27) stated that *C. salmonicolor* probably has a wider range of recorded hosts, belonging to the most diverse families, than any other tropical fungus known. It has been reported on cinchona in India and Ceylon (29, 40, 41, 178, 187, 264), Burma (48, 235), Java and Sumatra (1, 2, 13, 14, 20, 21, 27, 52-70, 71, 80, 88-93, 100-112, 121, 127, 131, 139-145, 147, 150, 154, 157-162, 164, 165, 167-170, 203, 205, 207, 210, 221, 223, 225-227, 239, 240, 244, 249, 251, 252, 259, 262, 263, 271, 275, 282, 287), Belgian Congo (122, 134), Guatemala,¹⁷ Colombia,¹⁷ and Peru (186).¹⁷ Rant (225) stated that he has observed the djamoer oepas on *Cinchona ledgeriana*, *C. robusta*, *C. succirubra*, *C. officinalis*, *C. pahudiana*, and hybrids in Java.

Davis and Crandall¹⁷ reported that pink disease occurs on cinchona in both plantations and wild stands and is possibly one of the primary enemies of cinchona in the Western Hemisphere. Symptoms of the pink disease on cinchona are the characteristic pink or whitish crusts on the bark, ringed or cracked bark, and occasionally calluses on the affected branch or trunk (225). The leaves turn red and dry up, and they remain attached to the diseased twig if the twig is young and dies suddenly (225). Frequently brown rust spots, caused by the bleeding and drying of the sap that exudes from the diseased areas, are found on the affected branches (225). Zehntner (287) stated that the symptoms of callus formation, cracking, and peeling in the absence of the fruiting stages of pink disease are sometimes mistaken for the symptoms of other stem and branch cankers by the cinchona growers in Java. Davis and Crandall¹⁷ stated that when the attack is most severe,

¹⁷ See footnote 16, p. 26.

the cankers girdle and kill the branches or trunks on which they occur. Rant (223) reported that by the time the fungus fruits on cinchona the attacked branch is almost or quite dead.

Rant (223, 225, 227) described four forms of the fungus on cinchona. In the first stage the mycelium, consisting of fine, shining white hyphae, grows over the bark and is known as the "cobweb mycelium" (225). The fungus is believed to exist saprophytically in this form. In the second stage the mycelium forms white nodules composed of dense coils of fine, colorless hyphae (225). These nodules occur chiefly in the lenticels and connect with the cobweb mycelium on the bark (27). Rant (225) believed that the hyphae of these nodules enter the lenticels and infect the host tissues. The *Corticium*, or third, form is a fruiting stage (237). It appears as a light-pink or faded-white crust, often somewhat cracked, on the under surfaces of branches and twigs and occasionally on the leaves (225). Usually the bark underneath these crusts is killed (225). *Necator decretus* Mass., which is said to be an imperfect form of *Corticium salmonicolor*, is also a fruiting stage, and its orange-red fruiting bodies occur usually on the upper surfaces of branches and twigs (225). These fruiting bodies are produced in the cracks of the bark or superficially on the bark (225). The *Necator* stage is usually found with the perfect stage or on the same cinchona plant on which the *Corticium* is fruiting elsewhere (225).

Rant (225) states that the hyphae of the fungus penetrate the bark and wood of affected cinchona plants, and in the tender succulent twigs of *Cinchona robusta* he found the pith to be infected in advance of the surrounding wood. Rant's inoculations proved that the disease organism can pass readily from one host to another, cinchona having been successfully infected from tea, coffee, hevea, loquat, and others (225). There is no evidence of biological races (27, 225).

Inoculations made by Rant (225) show that *Corticium salmonicolor* is able to infect the sound bark of *Cinchona ledgeriana* twigs when the atmospheric humidity is high. The moisture conditions favoring the attack of the fungus depend not only on the weather but also upon the density of the planting and the habit of the plant. *C. ledgeriana* makes a rather dense, bushy growth and suffers severely from pink disease, while *C. succirubra* makes a more open growth through which the air can circulate freely and is rarely injured (27). The disease attacks are said to diminish during the dry season, but they increase soon after the rains begin (157).

The extent of injury to cinchona varies: Young plants are frequently killed by the disease; but old trees are seldom killed by it, although the branches are badly stunted (100). In 1905 all the plants in low-lying nurseries on one Government plantation in Java were attacked and had to be replaced by grafts (2). Rant stated that the use of fungicides alone will not control the disease (223); the usual method recommended in Java is to cut and burn the affected branches during dry weather (223, 287).

Corticium salmonicolor Berk. and Br.—“Fructifications broadly effused, thin, adnate, membranaceous-soft, separable when moist, pale ochraceous buff to orange-pink when fresh, fading in the herbarium to pale olive-buff and cartridge-buff, pulverulent, even, cracking a little in drying, the margin thinning out; in section 100–200 μ thick, composed of hyphae running longitudinally over the substratum and bearing a broad layer of suberect, branching, loosely interwoven hyphae 4–7 μ in diameter, not incrustated, not nodose-septate; no gloecystidia; basidiospores hyaline, even, 9–12 \times 6–8 μ . The conidia of the imperfect *Necator* stage are catenulate, 14–18 \times 7–8 μ , according to Masee.” (26, pp. 227–228; also in 242, v. 6, p. 620.)

Dasyscypha

Hennings (123, 124) reported that *Dasyscypha warburgiana* P. Henn. causes cankerous overgrowths on the branches of *Cinchona ledgeriana* in the Preanger, Java. He stated that according to Warburg’s observations this fungus is probably the cause of “a disease” that occurs in the cinchona plantations in Java and has killed numerous trees for years (124). Engler and Prantl (75) and Stevenson (259) stated that it causes a serious canker disease in cinchona plantations.

Dasyscypha warburgiana P. Henn.—“Ascomatibus sparsis, brevissime stipitatis vel sessilibus, hemisphaerico-cupuliformibus, extus flavo-villoso, disco aurantio-flavo, margine integro, inflexo, $\frac{1}{2}$ –1 mm. diametro; ascis fusiformibus vel cylindraceo subclavatis carnescentibus dein hyalinis, octosporis 50 \times 70 und 7–9 μ ; sporidiis elongato-ellipsoideis vel subfusoides, hyalino-subcarnescentibus 5–10 \times 3–4 μ ; paraphysibus filiformibus.” (123, p. 226; also described in 75, 227, and in 242, v. 11, p. 413.)

Phytophthora

Although “stripe canker” of cinchona has been reported from Java and Sumatra only within recent years (44, 45, 88, 90, 93, 147–150, 258), it has probably been prevalent there for many years. In 1886 Eekhout (73) described cinchona canker as being easily recognized by the vertical furrows in the bark and the death of the wood underneath the furrows. Stripe canker is caused by *Phytophthora palmivora* Butl. and is characterized by the nearly parallel vertical depressions in the bark of the stem that precede a cankerous condition, the death of the tissues underneath the strips and extending into the root, rust-colored spots on the bark resulting from the bleeding and drying of the exuding sap, and leaf fall (150). For many years conspicuous rust-colored spots on the bark of diseased or dying cinchona trees were observed by the planters in the Netherlands Indies and the condition was reported as “stem rust” (60–62, 64–67, 70, 88–93, 106–109, 112, 126–129, 131, 148, 157–161, 227, 252).

Since diseases other than stripe canker or injuries to the bark of the tree might cause bleeding and drying of the sap, it is not possible to state with certainty that all the reports of stem rust refer to the stripe canker; however, when rust spots are accompanied by dying of the tissues in strips, as reported by some authors (131, 166, 227), then it is probably the stripe canker disease. Goot (90) reported that in 1933 stripe canker was of common occurrence in the plantations of South and West Sumatra. It is said to be especially injurious to some grafts and in *Cinchona*

succirubra seedling nurseries (149). Keuchenius reported that the stripe canker disease is the most serious of the cinchona trunk diseases in Sumatra (147), especially in damp, unterraced plantations and where pruning and thinning of the groves have not been done early (148). Failure to practice crop rotation in the seedbeds and mature stands was observed on plantations in Sumatra reporting damage to *C. ledgeriana* (9). Pruning the diseased tissue down to the wood and applying 5-percent carbolineum in Socony grease was found to be an effective control (9). Stoffels (262) recommended using 15 percent "carbolineum plantarum" and Coster (44) advised the selection of resistant clones.

(See *Phytophthora palmivora*, p. 15.)

Crandall and Davis (49) and Martin and Crandall (186) have reported a stem canker and top wilt on *Cinchona* spp. in Central America and South America. The disease was observed on *C. officinalis* and *C. pubescens* in the plantations and on *C. officinalis* var. *ledgeriana* grafted on *C. pubescens* in the nurseries and plantations in Guatemala, on *C. ledgeriana* plantation stock in Peru, and on *C. pubescens* and *C. pitayensis* growing wild in forest stands in Colombia (49). The same disease symptoms were reported on seedling cinchona stock in Guatemala and Puerto Rico (49). Inoculations of 18-inch potted *Ledgeriana* seedlings with the causal organism, tentatively identified as *Phytophthora parasitica* Dast., resulted in girdling stem cankers, dieback, and leaf infections (49). Considerable evidence of host resistance of individuals and of certain clonal lines was observed wherever the disease occurred (49). In Guatemala control of the disease in the plantations was obtained apparently by cutting back the tops of infected grafted stock (49).

In 1942 Kevorkian (151) reported a stem canker of cinchona in Puerto Rico. *C. pubescens* is particularly susceptible, but *C. officinalis*, *C. ledgeriana*, and the hybrids *C. pubescens* \times *C. officinalis* and *C. officinalis* \times *C. pubescens* are also attacked. The disease is especially prevalent on young plants (151). Harper and Winters (118) stated later that symptoms of the canker disease in Puerto Rico were similar to those of the disease in Guatemala and Peru caused by *Phytophthora parasitica*.

(See *Phytophthora parasitica*, p. 16.)

Undetermined

King (153) and Owen (198) reported a canker disease in the cinchona plantations in India. Dark, shriveled, brittle patches were found on the stems and branches (153). The patches varied in size and were not numerous on one tree, often being confined to one branch (153). The tissue of the bark was destroyed, adhering firmly to the wood of the tree, and a canker was produced (198). When a large spot occurred on a small tree, involving the bark almost around the stem, death of the tree resulted (153). If a tree were cut back, healthy shoots would be produced (153). The disease was most prevalent during the rainy season and

usually was not fatal (153). Owen (198) stated that fungus filaments were found in the decayed bark, occasionally penetrating the living tissue; however, he did not consider the fungus to be the cause of the disease. Pruning the diseased parts was recommended (198).

In 1879 Hooker (130) reported that large *Cinchona succirubra* trees in Jamaica were affected by a disease of the trunks in which an area of a few inches to a few feet in the middle of the trunk was said to be "contracted." The leaves showed a slight discoloration, and the bark and sapwood in the affected areas were dead. All the parts above the ringed areas died eventually, but the lower trunks and roots remained healthy. Suckers formed when the trees were cut back. The number of trees attacked was comparatively small. The mycelium of a fungus was believed to be responsible.

In 1930 and 1931 a destructive canker disease that girdled the branches or trunks of cinchona trees was reported from the Philippine Islands by Brown (22, 23). Trees up to 7 years of age were attacked and many were killed (22). Efforts to isolate a causal organism failed (23). Eradication methods were recommended for field control (23).

Martin and Crandall (186) reported a few cases of a canker of unknown cause on cinchona trees in Peru. The numerous cankers on both trunk and branches were about 1 inch in diameter and closely resembled the cat's-eye cankers caused by *Nectria* sp. No fungus or fruiting bodies were observed.

In 1945 Reinking¹⁸ reported a stem and crown canker affecting Ledgeriana grafts on *C. succirubra* stock in Guatemala. The grafts were dead at the point of union with the stock. Frequently a species of *Nectria* was observed on the dead areas above the graft, but it was not considered to be the cause of the disease. The disease was not epidemic, apparently; and faulty grafting methods and incompatibility of stock and scion were suggested as contributing factors.

Mitra (187) and Rhind (235) have reported a diseased condition of young cinchona plants in the Government cinchona plantation in Lower Burma. *C. succirubra* and a hybrid are affected, and *C. ledgeriana* is especially injured (235). The collar of an affected plant swells to about three times its normal diameter, and the bark turns dark brown to black and has many small longitudinal fissures (187). The lower leaves fall, and the leaves that remain on the extremities of the branches are subnormal in size (187). Badly affected plants usually die within a short time (187). Most of the abnormal size in the swollen collar region is caused by the thickening of the bark (187). The brown discoloration extends into the wood (187).

Microscopic examination failed to reveal the presence of either bacteria or fungi in any of the affected parts, and the disease was thought to be physiological (187). The plantation is located

¹⁸ See footnote 5, p. 4.

on a low hill with the top exposed to the southwest monsoon, while the sides are sheltered (187). Eighteen percent of the plants died on one-fourth acre on top of the hill, but only 3 percent died on the whole field of 20 acres (187). Too low an elevation (300 to 500 feet), too high temperatures, insufficient drainage, and over-exposure during the monsoon season are thought to be the causes of the abnormal development and disease in the plantation (187). Although a causal organism was not found, saprophytic fungi are said to invade the tissues of dead plants (187). *Botryodiplodia theobromae* Pat. (= *Diplodia theobromae* (Pat.) Nowell) was isolated from a few plants (29, 187).

(See *Diplodia theobromae*, p. 12.)

In 1930 McRae (178) reported a "bleeding disease" affecting cinchona trees at the Mungpoo and Munsong plantations in Bengal, India. The disease is said to be worse in plantations between 2,500 and 3,500 feet in elevation (178). Narrow red streaks occur on the surface of the bark near the base of dying trees (178). The bark under these patches is brown and dead to a varying depth, sometimes to the cambium (178). Bleeding takes place on the trunk and occasionally on the branches, chiefly during the hot weather during April (178). The "bleeding fluid" was found to be soluble in acetic acid and alcohol, leaving a residue of collapsed cells (178). Considerable areas of rust-brown stain are also present sometimes, especially toward the base of the stem, under which the bark is dead to the cambium and sometimes even the wood is stained brown (178).

It is stated that in the plantation the bleeding and rust stains are considered part of the same disease, although both symptoms are not always found together at the same time (178). Usually no hyphae are to be found in the bark tissues observed in the sections of the bark of either of these two types of stained bark (178). The bark of a few dying trees, however, which had the rust spots near the ground line but few obvious symptoms of bleeding, had cracked in longitudinal fissures several inches in length, exposing the wood (178). A layer of white to pale-yellow septate hyphae 3.5μ to 5.5μ in diameter was found between the dead bark and the wood of one tree (178). The hyaline to brown hyphae of another fungus, measuring 4.7μ to 7μ in diameter and sparingly septate, were found in the wood and sparingly in the bark of another tree (178). These fungi were not identified, although a later report by Mitra (188) stated that attempts were being made to isolate a fungus from diseased material. Padwick (199) in 1939 reported that a species of *Rhizoctonia* had been isolated from a plant with the "splitting disease" symptoms.

A disease on *C. succirubra* and on *Ledgeriana* grafts was reported in Java, in which the leaves turned yellow and the entire plant withered (132). Brown, sharply outlined spots were found in the region of the graft union. Several plants died from the disease.

Rutgers (240) reported a disease of unknown cause that affected the trunks of 2- to 4-year-old grafts in Java. Numerous cracks

appeared in the bark just above the root collar, and, although the roots were entirely sound, the wood in the collar region was killed. The disease appeared when the rains came and attacked plants on poor soils.

ROOT AND COLLAR ROTS

Frequently accurate diagnosis of root diseases is difficult because the symptoms on the upper parts of the tree may be similar to those of other diseases and because secondary organisms may have entered the wholly or partially destroyed root system by the time the visible symptoms are evident (19). Furthermore, unfavorable climatic and soil conditions, as impermeable subsoils and high or low soil-moisture content, may so weaken the plants as to afford easy entrance for soil-inhabiting fungi that would not attack vigorous, living roots (19). Consequently, the isolation of a particular fungus species from the roots of a dying tree does not indicate always that the isolate is the sole causative agent. It would be equally misleading to consider the death of less vigorous plants growing on unfavorable sites and invaded by weakly parasitic organisms as caused by physiological causes alone.

The terms "root rot," "root disease," "root canker," "root collar canker," "collar disease," "collar canker," and "collar rot" were used indiscriminately in the earlier literature from the Netherlands Indies in describing root and collar diseases of cinchona. The causal agents were unknown, and frequently such meager information was given on the symptoms as to make it difficult to associate with certainty these reports with the works of later investigators who described specific root diseases and fungi. Consequently, the contributions of such writers as Berkhout (10), Eekhout (73), Kessler (146), and Warburg (280) are of historical interest mainly and will not be reviewed here.

Many references have been made in the Dutch literature to the "root diseases" or "root fungi" of cinchona (14, 54-56, 58, 72, 96, 108, 126, 170, 240, 282, and others). Wurth (as reported by Hunger, 131) stated that diseases of the root system are rather widespread and are the most dangerous diseases of cinchona in Java. The symptoms are not recognized usually until the disease is too far advanced for control. Also, control is difficult because the roots are not easily accessible (131). Cases of root diseases generally occur locally (157), and attacked plants are uprooted and harvested (56, 257). *Cinchona succirubra* and its hybrids are less susceptible than *C. ledgeriana* (205), and grafted stock is said to show much greater resistance than seedling stock to root fungi (59). As a result, in Java the vacancies left by uprooting diseased plants are filled usually with grafts on *C. succirubra* stocks or hybrids (56, 205). In some nurseries where the loss from root diseases has been high, the number of grafted trees used as replacements is said to exceed the number of original seedling stock still left in the planting (57) and this interplanted grafted stock usually remains vigorous and healthy (56).

Armillaria

Armillaria mellea Vahl ex Fr., the "white root fungus" or honey agaric, causes a root rot of cinchona in the nurseries and plantations in Java and Sumatra (13, 28, 52, 60-62, 64-67, 69, 88-93, 100, 109-112, 131, 144, 147, 148, 150, 157-160, 169, 171, 210, 231, 251, 252, 257, 262, 263, 271), in Belgian Congo (122, 135), in Nyasaland (20, 278), and in Uganda (268). *Cinchona ledgeriana* is more susceptible than *C. succirubra* (227); and 1- to 2-year-old plants in the nurseries are most frequently attacked (131), although older trees may be attacked when grown in partially cleared fields in which there are many standing stumps and deadwood to serve as sources of infection (227). The presence of the disease is not apparent on the parts of the tree above ground until the fungus is well established in the roots (227). By the time the leaves begin to turn and fall, the main roots have become infected and the plant dies (131). Thin, rather tough white mycelium may be seen between the bark and wood of infected trees, and black rhizomorphs are sometimes found on the outside of the roots (227). Stoffels (262) states that the mycelial elements on cinchona are phosphorescent. Diseased plants should be grubbed out and the roots burned (131). If the area is to be replanted, all broken pieces of roots should be removed and well-slaked lime applied (131). Diseased stock in the nurseries in Java is replaced by *Ledgeriana* grafted on *C. succirubra* rootstocks (227).

Armillaria mellea Vahl ex Fr.—"Pileo carnoso, tenui, explanato, squamoso-piloso; margine expanso, striato; stipite spongioso-farcto; annulo floccoso patente; lamellis adnatis, dente decurrentibus, subdistantibus, pallidis, demum subrufescenti-maculatis, farinosis." (242, v. 5, p. 80.)

Fomes

In 1928 Sundararaman (265) reported *Fomes lamoensis* as causing a collar rot associated with *Rosellinia* sp. on 3-year-old cinchona trees at the Government cinchona plantation, Anamalai, India, and in 1931 Butler and Bisby (29) listed *F. lamoensis* (Murr.) Sacc. and Trott. as occurring on cinchona in India. In 1939 Keuchenius (150) reported a single case of *F. noxius* on cinchona in West Sumatra. According to Sharples (249), the brown root disease of rubber, tea, and other tropical plants was believed to be caused by *Hymenochaete noxia* until 1917. He reported that in 1917 Petch had found fructifications of a fungus on tea and hevea that had been killed by the brown root disease, and this fungus had been identified by Lloyd as *F. lamoensis* Murr. (249). Sharples (249) stated further that Corner pointed out in 1932 that two similar fungi had been confused: The first, *F. lamoensis*, is a harmless saprophyte, and the second, to which Corner assigned the name *F. noxius*, is a facultative parasite and the true cause of the brown root disease of *Hevea brasiliensis*.

In view of the above information it seems probable that the authors reporting *F. lamoensis* on cinchona in India were recording the occurrence of *F. noxius* Corner on this host. In reference to the occurrence of the disease at Anamalai, Sundararaman

(265) stated that the sanitation conditions on the plantation were very poor, since the ground was littered with dead trunks and rotting stumps and there was a lack of adequate surface and sub-soil drainage. He advised the installation of a good drainage system and the removal of the trees and stumps of species likely to harbor the fungus (265).

Fomes noxius Corner.—Effuso-reflexed, pileus applanate, dimidiate, slightly ascending, up to 13.5 cm. radius, 25 cm. wide, resupinate part spreading up to 35 cm. wide, upper surface rapidly glabrescent, growing margin white, creamy white, or pale ochraceous; flesh 6 to 19 mm. thick at base, rarely up to 5 cm. thick, 1.5 to 12 mm. at 5 mm. from margin, 0.5 to 2 mm. in the resupinate part, crust 0.5 to 1 mm. thick, with black crustaceous lines; tubes short in the first season, 2 to 5 mm. at the base, 0.3 to 1.5 mm. at 5 mm. from margin, developing 2 to 5 layers, with a total thickness up to 11 mm., carbonaceous; pores 80μ to 110μ wide, dissepiments 40μ to 100μ thick; spores 3.5μ to $4.5\mu \times 3.0\mu$ to 3.5μ ; cystidia sparse; hymenial setae none; extrahymenial setae in the flesh up to $600\mu \times 4\mu$ to 10μ , in the dissepiments up to $100\mu \times 9\mu$ to 16μ ; generative hyphae 2.5μ to 5μ wide (249).

Fusarium

Barat (6, 7) and Bugnicourt (24) have reported a serious root and collar disease on cinchona species in Indochina. Ledgeriana, Malabar, and *C. succirubra* plants are affected and also grafts (24); and the disease is found throughout the cinchona plantations, causing high mortality in seedbeds and plantations at an approximate elevation of 1,100 meters (7). Hendrickx (122) stated that cinchonas that have been planted too deeply are attacked by the disease. A species of *Fusarium*, possibly *F. moniliforme* Sheldon or an odorless variety of *F. vasinfectum* Atk., was isolated from the stems and roots of diseased plants (7). *Nectria perithecia* obtained from branches also produced *Fusarium* in culture (24).

The fungus attacks the inner bark of the stem, root, collar, and large roots (7) and forms cracks and cankers on the large roots extending up through the collar region (24). The cankers gradually become deep and circular in shape (24). The disease is initiated in the nurseries, and infected nursery plants showing the characteristic cracks and cankers wither soon after being transplanted and the leaves turn red (24). The growth of the plants is slowed down or stopped, and the plants die very slowly (7). The alkaloids in the bark are reduced very rapidly; however, the damage at elevations less than 1,100 meters has not been so great but that most of the trees could be exploited in 5 to 10 years (7). Better care and maintenance of the plantations is said to retard the disease (7).

The following control measures were suggested: Disinfection of the seed, use of soil disinfected with carbon disulfide or formaldehyde in the seed and nursery beds, discarding at transplanting time all plants showing symptoms, and destruction of all diseased plants in the nurseries and plantations (24). Barat (7) suggests that the disease might be overcome by planting at higher elevations, if these were available in Indochina, or by breeding disease-resistant varieties.

Fusarium sp.—Pustules on roots and stems whitish, projecting, round, isolated or in groups; spores hyaline, curved, 3- to 5-septate, 32μ to $42\mu \times 4\mu$ to 4.5μ ; *Nectria* perithecia spherical, reddish orange, isolated or in groups, with distinctly red papilla; asci 8-spored, $14\mu \times 6\mu$; ascospores 2-celled, hyaline, rounded at both ends, constricted (24).

In 1939 Bugnicourt (25) listed *Fusarium vasinfectum* Atk. as occurring on the stems and roots of *C. ledgeriana* in Central and South Annam, Indochina.

(See fusarium damping-off, p. 4, and fusarium stem blight, p. 12.)

Ganoderma

Ganoderma pseudoferreum (Wakef.) v. Over. and Steinm. has been reported on cinchona in Sumatra and Java (13, 89, 90, 158-161, 257) and Belgian Congo (134). In 1933 Leefmans (160) reported that a great deal of damage was done in the older Ledgeriana nurseries by the "root fungi" (including *Armillaria mellea*, *Rosellinia* sp., and *Ganoderma pseudoferreum*). In 1935 Goot (90) stated that in Sumatra the extent of damage caused by the red root fungus was exceeded only by that caused by *Rosellinia* spp., both in the nurseries and on plants up to 4 years old. The red root fungus, however, is said to remain thoroughly localized (160). Steinmann (257) stated that burning diseased plants, digging isolation ditches around infected areas, clear-cutting, and removing stumps from forest lands should be adequate for control.

Ganoderma pseudoferreum (Wakef.) v. Over. and Steinm.—Rhizomorphs light red becoming dark wine red to violet black in older specimens, flattened, branching, closely adpressed to surface of host roots, sometimes forming extensive networks or coverings, white mycelia from rhizomorphs penetrate host roots to cause a rot; sporophores solitary or only a few together, with more or less clearly developed stalk and cap, resupinate when on lower surface of exposed roots; cap more or less round, up to 30 cm. in diameter, upper surface rough in center, brown to brown black or brown red with violet or greenish tinge, margin zonate, light red brown; stalks when present same color as center of cap; inner tissue of cap black brown when fresh, cinnamon-colored with black-brown radial stripes when dry, composed of very thick-walled, branching hyphae, 4.5μ to 8.5μ in diameter; tubes darker than tissue, drying to a brownish color on the outside, barely 1 mm. long at edge of cap but up to 10 mm. at base of cap, not stratified; pore surface dirty yellowish white when dry, gray violet when drying, pores 90μ to 140μ in diameter; basidia colorless, 4-spored, short, about $20\mu \times 7\mu$; spores ellipsoid, hyaline at first, later with yellow-brown inner wall, slightly roughened, 6μ to $9.5\mu \times 2.8\mu$ to 4μ (197).

Physiological

King (153) reported a physiological disease affecting cinchona in the plantations in Sikkim and the Nilgiris, India, in 1871-72. The disease was confined to trees in damp soils (153). The roots of the trees were attacked first, and the cortical and woody tissues gradually shriveled from the root upward (153). The leaves became discolored and fell, and the plants usually died (153). Owen (198) stated that the shriveled appearance of the young plants, the shrinking of the leaves, and the stunted growth of the primary branches were noticeable for some time before the roots

decayed. Draining the soil and harvesting the diseased trees before the bark became worthless were recommended (198).

Calder (36, 37) and Cowan (47, 48) reported a collar canker disease on cinchona in the Burma plantations in 1924-27. Only a limited amount of canker was present in 1923-24 (36); but in 1926-27 approximately 21,000 plants were killed, while many more were affected by the disease (48). The first symptoms were "flagging" of the whole plant, and the bark at the soil level and extending a few inches above became thick and dark and cracked longitudinally (37). Calder (37) stated that the symptoms increased rapidly during the rains in 1924-25. Trees in a state of collapse were observed on the sides of ridges, but were much more prevalent on the flat damp wind-swept crests (37).

Microscopic and cultural investigations failed to reveal a fungus in the affected plant tissues (37). The disease was believed to be a physiological disorder caused by planting too deeply, combined with wind action (37). When the young trees were swayed by the wind, hollow cone-shaped depressions were formed in the soil at the collar of the stems, in which water collected and was retained (37). Plants with the root systems near the soil level and with the collar completely free seldom exhibited the symptoms (37). Removing the earth from around the collar, shallow planting, good drainage (37), and the use of windbreak hedgerows were found to minimize the trouble (47).

The occurrence of root rot in cinchona plantings in Puerto Rico is believed to be favored by soil unsuitable for cinchona (117). The plantings are located on Nipe clay, a ferruginous laterite, which has a very low air content and becomes waterlogged during the rainy season. The taproot and some of the deeper roots become diseased and die. A mat of new roots is formed at the base of the diseased roots near the surface of the soil. This condition results in a check in the vigor of the trees, and some of the trees attain a pathological maturity after one or two seasons.

Phytophthora

Thompson (269) reported a root rot and dieback of *Cinchona ledgeriana* trees in the Cameron Highlands, Malaya. The affected trees were growing on a combination of shallow, sandy topsoil and a stiff subsoil into which the roots did not penetrate. The main root system partly decayed, and the bark just above and below the soil line was cankered or split. The cortex was soft and mottled with red-brown lines. *Phytophthora cinnamomi* Rands was isolated from the roots but not from the cankered tissue above the soil line. Often a species of *Phomopsis* was found in the affected cortex. Inoculation of roots and stems of trees and seedlings, however, indicated that the *Phytophthora* was a slow-acting, weak parasite and that the *Phomopsis* was probably a secondary organism. The *Phytophthora* did not reproduce the typical bark canker symptoms and advanced only a slight distance in the cortex in 2 months; and 2 to 5 months elapsed before wound-inoculated seedlings died. Thompson advised planting

Ledgeriana grafted on *C. succirubra* rootstocks, unless good rich virgin soil was available for Ledgeriana seedlings.

(See phytophthora stem blights, p. 13, and phytophthora cankers, p. 32.)

In 1944 Crandall and Davis¹⁹ reported high mortality from a root disease affecting the calisaya form of *C. officinalis* in field plantings located in the valleys of the eastern slopes of the Andes in Bolivia. The infected plants had been grown from locally collected seed of wild trees. In several cases an entire planting had been destroyed. Faulty field-planting practices were said to be contributing factors. The young plants had been placed in the bottoms of holes about 1½ feet deep, and the drowning of the roots during the rainy season and the inability of the roots to make normal growth in the impervious clay subsoil apparently allowed rot fungi to gain entry. A fungus with the hyphal characteristics of a *Phytophthora* was isolated from infected roots. Wild plants of the same cinchona species were found with rot lesions on the roots but otherwise were in healthy condition. The selection of root-rot-resistant plants and the modification of field-planting methods were suggested as means of cutting down plantation losses.

In 1944 Martin and Crandall (186) reported a root disease on Ledgeriana variety of *C. officinalis* at an elevation of about 3,700 feet in Peru. A species of *Phytophthora* was said to be the probable cause. Part of the planting was located on a well-drained hillside, where the death rate was about 35 percent, but the planting in the poorer drained bottom land had a mortality rate of about 56 percent. Infection appeared to be in the collar region, and death resulted when girdling took place at or above the collar within a year or two after infection. A root and collar rot in a newer planting was considered to be caused by the same fungus, although planting too deeply was thought to be partly responsible for the disease.

Rosellinia

Species of *Rosellinia* cause the black or gray root rots of many tropical plants (230). In 1935 Heusden reported that "root fungi (*Rosellinia* sp.)" were the principal cause of disease in old plantations of cinchona seedlings in South Sumatra (128). *Rosellinia* sp. has been recorded on *Cinchona* spp. in India (265), Belgian Congo (122, 135), Indochina (6), and Java and Sumatra (45, 60-62, 64-67, 69-71, 88, 89, 92, 93, 100, 101, 108-112, 128, 129, 144, 145, 157, 158, 160, 171, 172, 202, 210, 227, 231, 251, 252, 263, 271).

Rosellinia arcuata Petch, frequently called the "gray root fungus" in the Dutch cinchona literature, has been reported on cinchona in Java and Sumatra (13, 45, 90-93, 147, 148, 150, 159, 161, 230, 257, 262). In 1931 Leefmans (159) wrote that *R.*

¹⁹ CRANDALL, B. S., and DAVIS, W. C. OCCURRENCE OF CINCHONA ROOT ROTS IN THE AMERICAS. U. S. Bur. Plant Indus., Soils, and Agr. Engin., Plant Dis. Rptr. 28: 926-929. 1944. [Processed.]

arcuata occurred quite generally throughout South Sumatra and was rather serious on some plantations. Keuchenius (147) reported that in 1935 trees were still dying regularly from *Rosellinia* attack on an estate in West Sumatra where the young cinchona plantation had been injured by fire in 1930. Rant (230) states that the gray root fungus attacks young plants of *Ledgeriana*, *C. succirubra*, *C. robusta*, and hybrids in the nurseries and also occurs in plantations on the roots of older trees of the same species. It has been reported to affect seedlings in the germination beds occasionally (54). *C. succirubra* is more resistant than *C. ledgeriana* (257).

Since the fungus spreads by means of aerial-borne conidia or by root contact, diseased plants may occur in the plantations as isolated specimens or roughly grouped in circles (257). The fungus attacks the tips of primary and secondary roots; and the mycelium, which spreads over the surface of the root, is a gray-white color at first, later turning black (257). The white mycelium found between the bark and wood is characteristically arranged in small fan-shaped, starlike figurations about 1.5 cm. in diameter (257). The external mycelium may grow up over the collar and the base of the trunk under favorable conditions of moisture (262). Rant (230) successfully inoculated *Ledgeriana* seedlings with the fungus. He states that good drainage and cultivation of the soil in the nurseries are desirable and that diseased plants should be dug out and burned immediately (230). When the disease occurs in the plantation, the diseased trees should be removed and the area of contaminated soil should be isolated by drains and ditches (230). The use of resistant plants, as *Ledgeriana* grafts on *C. succirubra* rootstocks, is recommended by Steinmann (257).

Rosellinia arcuata Petch.—“Peritheciis gregariis, primum in mycelio purpureo-brunneo immersis, fusco-brunneis, dein nigris, liberis, carbonaceis, globosis, leniter depressis, 1.5–2.4 mm. diam., levibus, ostiolo conico 0.1 mm. alto, basi 0.4 mm. cr.; ascis cylindraceis 300×8 ; sporis oblique monostichis; paraphysisibus 2μ circ. cr., ascos aequantibus; sporis nigris, cymbiformibus, apicibus acutis et saepe mox contractis, $30\text{--}47 \times 5\text{--}7$.” (242, v. 24, p. 834.)

Rosellinia bunodes (Berk. and Br.) Sacc. has been reported on *Cinchona* spp. in Java and Sumatra (90, 121, 147, 148, 150). Goot (90) stated that *R. bunodes* and *R. arcuata* were first in importance among the root fungi causing losses in the nurseries and on older plants (up to 4 years old) in the South and West Sumatra plantations in 1933.

Rosellinia bunodes (Berk. and Br.) Sacc.—“Peritheciis magnis, 2 mill. diam., globosis verrucosis e strato strigoso oriundis; sporidiis maximis fusiformibus, apicibus elongatis acutissimis, multinucleatis . . . Mycelium ex hyphis fasciculatis fuscis, articulatis, ramosis, sursum pallidioribus constans; conidiis moniliformi-catenulatis $4\text{--}5\mu$ long.; asci luminis apice capitato-foveolati.” (242, v. 1, p. 254.)

Thompson (270) reported a fatal root and collar rot of two young cinchona hybrid trees in Malaya in 1940. The disease was caused by a species of *Rosellinia*.

Kevorkian (151) and Harper and Winters (118) have reported a root disease of cinchona in Puerto Rico caused by a fungus that appears to be similar to the "gray root fungus" of Java (the *Graphium* or imperfect stage of a species of *Rosellinia*). The disease is not detected until the trees, which appear to be healthy but making poor growth, suddenly wilt (151). The leaves dry out and remain attached for some time before falling (151). The main roots are found to be rotted off, leaving short stubs (151). When the adventitious roots have likewise rotted, the trees wilt and eventually die (151). Kevorkian stated that the prevalence of the disease in Puerto Rico may be caused in part by the fact that the plantation is on poorly drained clay soil (151).

Reinking²⁰ reported a root rot disease killing *Ledgeriana* plants on their own roots in Guatemala. In one case several trees in one row had been killed, while *C. succirubra* plants in the next row were not affected. Although a *Rosellinia*-like fungus was found on the roots of dead trees in one planting, the type of soil in which the trees were planted was also considered as a possible contributing factor.

Undetermined

Crandall and Davis²¹ observed a dry root rot in a wild stand of *Cinchona pubescens* in Colombia, but the disease was not considered to be of economic importance.

Crandall and Davis²¹ reported a root rot and collar canker disease on *C. officinalis* var. *ledgeriana* grafted on *C. succirubra* in Guatemala. The foliage of infected trees was thin, and the leaves were reddish and somewhat smaller than normal. A very narrow streak of infected cambial tissue was found to extend from a root lesion in the rootstock to a girdling lesion in the *Ledgeriana* scion, connecting through the graft union. These canker lesions on the scion extended up the trunk as much as 3 feet before complete girdling occurred just above the graft union. Some plants of *Ledgeriana* growing on their own roots were also attacked by the disease. This disease is thought to be caused by a soil-borne parasite that is probably also an active root rot on susceptible rootstocks.

The occurrence of a root disease in a 10-year-old planting of *Ledgeriana* in Peru has been reported by Crandall and Davis²¹. The mortality rate is about 20 percent, and in some cases as many as three replants have been killed by the disease. The infection apparently starts in the collar region below the soil line, either on the main roots or at the base of the trunk. Cambial tissue is said to be invaded as high as 10 inches above the soil line before the tree is girdled and dies.

²⁰ See footnote 5, p. 4.

²¹ See footnote 19, p. 41.

INJURIES ATTRIBUTED TO ENVIRONMENTAL FACTORS OTHER THAN SOIL CONDITIONS

DROUGHT

Calder (38) reported an unusual reaction by cinchona plants to drought injury in the plantations in South Burma in 1928-29. During a long period of dry weather a large area of cinchonas, which had been planted shallow in order to minimize the risk of collar disease, became quite dry and the plants lost almost all of their leaves. Within a week after the rains began the plants had leafed out and made healthy growth.

Rant (226) reported drought-killing of cinchona seedlings in seedbeds in Java. The humus, before being placed in the seedbed, had become rather impermeable to water, owing to overexposure to sunlight. Consequently, when the beds were watered, the moisture stood on the surface of the bed while the underlayer of soil remained quite dry.

FROST OR FREEZING

Leersum (163) reported severe frost injury to cinchona plants in old and young nurseries following a drought in Java. When the trees are badly hit by frost, the bark splits from the wood and remains loosely attached. The wood is brown and the bark is hard and brittle and fibrous. As thawing takes place rapidly in the Tropics, the plants quickly wilt and turn black. The sap exudes as a dark-brown liquid and runs down the trunk, drying to a rust-brown color. The alkaloidal content is not appreciably lowered if the bark is harvested within a short time.

LIGHT

Tests at the Puerto Rico Agricultural Experiment Station, at Mayaguez, showed that cinchona seedlings with poor root development caused by excess moisture in the seedbed are more sensitive to light than plants with good root development.²² Seedlings that are exposed to a slight excess of light become weak and the leaves develop a pinkish-red color, while overexposure to direct sun rays results in severe leaf burning (217). Reinking²³ has reported a leaf scorch characterized by red-brown scorched areas extending from the margin inward and thought to be caused by sunburning of weakened leaves, as occurring in Guatemala. Augusto (5) stated that lack of adequate shade results in a stunted condition and subnormal size of the leaves of *Cinchona calisaya*. Severe injury can result in the death of the seedlings (117), and Popenoe (208) stated that seedlings in the seedbed may be killed within a few hours if exposed to the direct rays of the sun. Cowan (48) reported that in 1926-27 the exceedingly hot sun at Mergui, Burma, caused more deaths in the cinchona plantation than did insects and fungus diseases.

²² HARPER, R. E. CINCHONA PRODUCTION. Rpt. Activities of Puerto Rico (Mayaguez) [Fed.] Expt. Sta. for months of Jan., Feb., and Mar., 1943: 31-35. 1943. [Processed.]

²³ See footnote 5, p. 4.

LIGHTNING

Gorkom (97) reported lightning injury to *Cinchona succirubra* and hybrid trees in Java. The trees were stripped of their leaves, and the bark was brown and difficult to peel (97). Rant (223, 225) stated that the sap exudes and runs down the trunk, drying in rust-brown spots.

RAIN AND HAIL

Popenoe (208) stated that during the early months in the nursery many of the young cinchona plants may die if not protected from heavy rains. Washing of the soil in the seedbed may kill germinated seedlings by mechanical injury (202). Hail, if heavy, is said to strip a plantation of its leaves, but the check in growth is only temporary usually, as new leaves appear speedily (153). In 1932 Calder (33) reported considerable damage to the Munsong cinchona plantation in India by a severe hailstorm in May 1931.

SHADE

Tests at the Puerto Rico Agricultural Experiment Station, at Mayaguez, have shown that undesirable etiolation occurs in excessive shade (217).

SMOKE

Pring (212) reported that all the leaves on *Cinchona officinalis* growing in the City Garden, St. Louis, Mo., U. S. A., showed leaf burn after the cloud of smoke of December 17-18, 1931. The plants died.

Reinking²⁴ reported black blotching of cinchona leaves in Guatemala, caused by the rubbing of leaves covered with a layer of pumice from an active volcano.

WIND

Gorkom (94) quoted from the 1870 report on the cinchona crop in Java the statement that "the wind remains" the worst enemy of the cinchona plants. King (153) stated that excessive and frequent wind appears to do considerable and permanent damage, especially to *Cinchona succirubra*, the leaves of which are large and tender. In 1887 some cinchona trees in Jamaica were reported to have died, owing to wind action during a hurricane (137). The bark was injured at the collar region, and the subsequent invasion of the root bark by fungal mycelium resulted in the bark rotting (137).

MISCELLANEOUS FUNGI AND LICHENS

A number of fungi have been reported on cinchona that are not listed in this publication with those causing the diseases of cinchona, either because the reports were brief and failed to indicate diseased conditions caused by the fungal organisms or because the fungi are usually considered nonparasitic, although frequently found on plants dying from other causes. Brief discussions of these reports follow.

²⁴ See footnote 5, p. 4.

Reichert and Hellinger (234), in reporting a tip end rot of banana fruits and flowers, listed cinchona as one of the many hosts attacked by *Botrytis cinerea* Pers.

A species of *Chaetomium* has been reported on dead twigs of cinchona in Uganda by Hansford (114).

In 1907 Koorders (155) reported a parasitic species of *Diplodia*, which he described under the name of *D. cinchonae* Koord., on the bark of old branches of *Cinchona succirubra* trees 1,500 meters above sea level in Middle Java. These branches were also attacked at the same time by *Corticium salmonicolor* Berk. and Br. and its so-called imperfect stage, *Necator decretus* Mass. (155). The *Diplodia* was scattered sparingly in the plantations (155). Rant (227) also collected *D. cinchonae* on dead branches of *Cinchona* sp. in Java and considered it to be saprophytic.

Diplodia theobromae (Pat.) Nowell has been reported as one of the fungi associated with root diseases on cinchona in Belgian Congo (135, 262).

An unidentified species of *Diplopeltis* was observed by Koorders (154) in 1905 in the bark of dying branches of 30-year-old *C. succirubra* trees in Middle Java. Numerous pycnidia with spores in various stages of development were found. Koorders (154) also found *Diplopeltis* on a 1-year-old cinchona hybrid that had died primarily because of a root disease. A few hyaline 2-celled spores were found on the later collection, but no pycnidia.

A species of *Fusoma* was observed by Koorders (154) in Java on the bark of a 1-year-old cinchona hybrid that had died primarily because of a root disease.

Glomerella sp. is listed as occurring on *C. ledgeriana* in India in 1936 (136).

In 1936 Keuchenius (147) reported *Helicobasidium compactum* Boed. as a root fungus on cinchona in West Sumatra.

A species of *Helicobasidium* has been reported on cinchona in Sumatra (160) and in Belgian Congo (135). It is said to be associated with root diseases (135, 262).

Ocfemia and Celino (196) and Wollenweber (284, 285) listed cinchona as one of the hosts of *Hypomyces haematococcus* (Berk. and Br.) Wr.

Hypomyces ipomoeae (Halst.) Wr. has been recorded by Wollenweber (285) as occurring on cinchona.

Hansford (113, 115, 116) has reported *Rhizoctonia lamellifera* Small on *Cinchona* sp. in Uganda. The fungus form with the large sclerotia was the type observed on cinchona (115). In 1929 Leefmans (158) reported *R. bataticola* (Taub.) Butl. on *Cinchona* sp. in South Sumatra. *R. bataticola* and *R. lamellifera* are synonyms of *Macrophomina phaseoli* (Maub.) Ashby (4).

Koorders (154) reported a species of *Pestalozzia* on mature living leaves of old *C. succirubra* trees and on the dead stem of a 1-year-old cinchona hybrid in Middle Java. He stated that perhaps the species was identical with *P. cinchonae* Zimm.

Wallace (276, 277) reported *Phomopsis* sp. on twigs of *Cinchona* sp. in Tanganyika.

Polyporus rubidus Berk., the pink root fungus, has been reported on *Cinchona* sp. in West Sumatra (93, 148, 150).

Seymour (248) listed *Polystictus fimbriatus* Fr. (= *Polyporus fimbriatus* Fr.) on *Cinchona* sp.

Keuchenius (148, 150) has reported *Poria* sp., "the brick red root fungus," on cinchona in West Sumatra.

Rant (227) and Leersum (167) reported the occurrence of *Stilbum minutula* Penz. and Sacc. on dying branches of young Ledgeriana grafts and seedlings and on branches of *C. succirubra* and hybrids in Java. The fungus, which is usually a saprophyte on deadwood and stems, forms red fruiting bodies on the cinchona branches (167). On cinchona branches that have been weakened from other causes, however, the fungus can invade the living host tissues and kill them (227). Although Rant considered the fungus identical to *Stilbum minutula* Penz. and Sacc., he preferred to call it *Stilbella* (227).

The following fungi were reported on commercial cinchona barks: *Cystodium coccineum* Fée on bark of *Cinchona lancifolia* Mutis in South America (77); *Gloniella chinicola* Rehm and *Gloniopsis regia* Rehm on bark offered for sale as "cinchona regia" (227, 232, 242); *Himantia cinchonarum* Fée on cinchona trees in Peru (76); *Hysterium enteroleucum* (Ach.) Fr. on cinchona trade bark from South America (227, 242); *Myriangium cinchonae* Rehm on bark of "cinchona regia" offered for sale in India (29, 233, 242); *Rhabdospora thallicola* Tassi in a lichen on cinchona bark in Brazil (242, 267); *Rhizomorpha cinchonarum* Roth on bark of cinchona trees in "America" (76, 227, 242); *R. crinum* Fée on bark of *C. cordifolia* Mutis from South America (76, 227, 242); *Stilbospora fumosa* Fée on bark on *C. floribunda* Sw. from South America (76, 227, 242); *Syncephalastrum elegans* El. Marchal on bark of "cinchona rubra" offered for sale at Brussels (227, 242); *Thelephora aurea* Zenker on "china rubra" in South America (288); *T. cyanescens* Fée on cinchona bark in Lima, Peru (77); and *T. lactea* Fr. (= *Corticium lacteum* Fr.) on several species of *Cinchona* in South America (288).

Lichens, which are usually classed as a distinct group of plants, are composed of a fungus and an alga living in a symbiotic relationship (19). They are found on rocks and the bark of living or dead trees (19). The presence of lichens on a tree causes little or no damage to the host, although an abundant growth of lichens may cause the death of small twigs or small branches by smothering them and thus interfering with the normal plant processes (19).

The lichens in the following lists were reported by Fée (76, 77) and Zenker (288) as collected from trade barks of *Cinchona* species.

On *Cinchona glandulifera* R. and Pav.: *Graphis afzelii* Ach., *G. duplicata* Ach., *Opegrapha congesta* Fée, *O. subimmersa* Fée.

On *Cinchona laccifera* R. and Pav.: *Chiodecton paradoxum* Fée, *Graphis interrupta* Fée.

On *Cinchona officinalis* L.: *Arthonia gregaria* Fée, *A. leucocheila* Fée, *A. leucocheila* var. *pallida* Fée, *A. obtrita* Fée, *A. patellula*

Fée, *A. rugosa* Fée, *A. sinensigrapha* Fée, *A. sulfurea* Fée, *Chiodecton depressum* Fée, *C. meratii* Fée, *C. sphaerale* Ach., *C. umbratum*, *Collema diaphanum* Ach., *Fissurina dumastii* Fée, *Glyphis favulosa* Ach., *Graphis balbisii* Fée, *G. cinnabarina* Fée, *G. cleitops* Fée, *G. frumentaria* Fée, *G. fulgurata* Fée, *G. laubertiana* Fée, *G. oryzaeformis* Fée, *G. plagiocarpa* Fée, *G. rubiginosa* Fée, *Hypochnus albidus* Fée, *Lecanora atra* Ach., *L. punicea* Ach., *L. russula* Fée, *Lecidea aurigera* Fée, *L. brebissonii* Fée, *L. carenola* var. *arceutina* Ach., *L. condamineana* Fée, *L. conspersa*, *L. cuticula* Fée, *L. disjuncta* Fée, *L. glaucotheca* Fée, *L. grisea* Zenker, *L. hypoxantha* Fée, *L. leucoxantha* Spreng., *L. mutabilis* Fée, *L. punctulata* Fée, *L. tremelloidea* Fée, *Opegrapha condaminea* Fée, *O. globosa* Fée, *O. inaequalis* Fée, *O. nana* Fée, *O. ovata* Fée, *O. rhabdotis* Fée, *O. rhizocola* Fée, *O. scaphella* Ach., *O. subimmersa* Fée, *Parmelia appressa* Zenker, *P. goebelii* Zenker, *P. melano-leuca* Zenker, *Porina chiodectonoides* Fée, *P. marginata* Fée, *P. pertusaria*, *P. sclerotium* Fée, *P. tetrathalamia* Fée, *Pyrenodium clandestinum* Fée, *Pyrenula annularis* Fée, *P. clandestina* Fée, *P. discolor* Ach., *P. epappilata* Fée, *P. mollis* Fée, *P. myriocarpa* Fée, *P. pinguis*, *Sarcographa cinchonarum* Fée, *Thelotrema concretum* Fée, *T. terebratum* Ach., *T. urceolare* Ach., *Triclinium cinchonarum* Fée, *Trypethelium inconspicuum* Meissn., *Usnea barbata* Ach., *Variolaria amara* Ach., *Verrucaria acharii* Fée, *V. amara*, *V. catervaria* Fée, *V. pyrenuloides* Fée.

On *Cinchona pubescens* Vahl: *Arthonia sinensigrapha* Fée, *Chiodecton effusum* Fée, *Graphis cinerea* Fée, *G. elongata* Zenker, *G. intricata* Fée, *G. subbifida* Zenker, *Hypochnus rubrocinctus* Ehr., *Lecanora melano-xantha* Zenker, *Lecidea chloroplaca* Fée, *L. rubrica* Zenker, *L. tuberculosa* Fée, *Opegrapha conglomerata* Fée, *O. peruviana* Fée, *O. rugulosa* Fée, *O. scaphella* Ach., *Parmelia tiliacea* Ach., *Pyrenula viridescens* Fée, *Trypethelium clandestinum* Fée, *Verrucaria pyrenuloides* Fée.

On *Cinchona* spp.: *Arthonia confluens* Fée, *A. divergens* Fée, *A. glyphoides* Fée, *A. granulosa*, *A. lecanoroides* Fée, *Ascidium cinchonarum* Fée, *A. cinchonarum* var. *verrucosum* Fée, *Asterisca cinchonarum* Spreng., *Chiodecton monostichum* Fée, *C. sphaerale* Ach., *Fissurina incrustans* Fée, *F. irregularis* Fée, *Graphis acharii* Fée, *G. atrosanguinea* Zenker, *G. aurantiaca* Zenker, *G. canaliculata* Fée, *G. chlorocarpa* Fée, *G. cinnabarina* var. *distans* Fée, *G. cometia* Fée, *G. conferta* Zenker, *G. cooperta* Zenker, *G. fulminatrix* Zenker, *G. grammitis* Fée, *G. haematites* Fée, *G. inconspicua* Fée, *G. interrupta* Fée, *G. poitiaei* Fée, *G. reniformis* Fée, *G. sordida* Fée, *G. subcurva* Zenker, *Hypochnus nigrocinctus* Ehr., *Lecidea biformis* Fée, *L. brunneo-atra* Zenker, *L. luteola* var. *americana* Fée, *L. parasema* Ach., *L. parasema* var. *americana* Fée, *L. patellula* Fée, *L. sanguineo-macularis* Zenker, *L. thelotrematis* Fée, *L. vernalis* Ach., *L. versicolor* Zahlbr., *Leptra farinosa* Zenker, *L. flava*, *Ocellularia thelotrematoides* Zenker, *O. urceolaris* Spreng., *Opegrapha agelaea* Fée, *O. endochroma* Fée, *O. hiascens* Fée, *O. rigida* Fée, *O. tumidula* Fée, *O. vernicosa* Fée, *Parmentaria cinchonarum* Fée, *Porina americana* Fée, *P. depressa* Fée, *P. desquamascens* Fée, *P. marginata* Fée, *P. tetrathalamia* Fée, *P. verrucosa* Fée, *Porophora rufescens* Zenker, *Pyrenodium clandestinum* Fée, *P. macrocarpon* Fée, *Pyrenula aggregata* Fée, *P. cartilaginea* Fée, *P. ceratina* Fée, *P. cinchonae* Fée, *P. kunthii* Fée, *P. marcida* Fée, *P. myriocarpa* Fée, *P. nitens* Fée, *P. porinoides* Fée, *P. pupula* Ach., *P. subfarinosa* Fée, *Sarcographa inquinans* Fée, *Sticta aurata* Ach., *S. macrophylla* Delise, *Thelotrema calvescens* Fée, *Trypethelium erumpens* Fée, *T. inaequale* Fée, *Urceolaria cinchonarum* Fée, *Variolaria depressa* Zenker, *V. fulva* Fée, *V. globulifera* Ach., *V. microcephala* Fée, *Verrucaria decolorata* Fée, *V. diluta* Fée, *V. exasperata* Zenker, *V. macrozoma* Fée, *V. myriococca* Spreng., *V. nitida*, *V. parasema* Zenker, *V. pustulosa* Zenker, *V. salebrosa* Fée, *V. sinapisperma* Fée, *V. socialis* Zenker.

MYCORHIZAE

Since the investigations of Frank in 1885, it has been known that the roots of many plants live symbiotically with certain kinds of fungi (255). Ludowyke (175) stated that there are two

schools of thought regarding the significance of the presence of mycorrhizal fungi in the roots of plants. On the one hand, it is believed that the association is of mutual benefit to the host and fungus, and on the other, that the condition is one of suppressed parasitism and that the host plant receives no benefit from harboring the fungus (175). It is possible that there is a range of conditions from the one extreme to the other (175). If the condition is one of suppressed parasitism, then any factor that reduces the vitality of the plant will lessen its resistance to the fungus, with a consequent increase in degree of parasitism and the possible death of the plant (175). Ectotropic mycorrhizae form a sheath around the root and endotropic mycorrhizae live within the host root cells (255).

Steinmann (255, 256) reported the presence of endotropic mycorrhizae in the roots of healthy plants of *Cinchona succirubra* in Java. The mycorrhizae are found in the thin capillary roots as well as in smaller roots 2 to 3 mm. in diameter (255). The fungus is not present throughout the parenchymatous tissue of the root cortex but only in certain places (255). These places may be recognized sometimes from the exterior of the root by irregular and sometimes nodular swellings on some parts of the root (255). Sometimes the fungi are found several cell layers deep in the cortex, but do not penetrate the endodermis (255). The hyphae are mostly brown, inter- and intra-cellular, branched, and generally 1.7μ to 3.5μ thick (255). In some cases the hyphae unite into bundles, which change into rough yellow nodular masses (255). In other cases dark-gray to brown fungal tissue of pseudoparenchymatic structure is found, which strongly resembles sclerotia (255). Steinmann found the mycorrhizae in cinchona roots in February and April on estates at very high elevation where the pH of the soil was 5.1 to 6.2 (255).

LITERATURE CITED

- (1) ANONYMOUS.
1904. HET MEEST WETENSWAARDIGE UIT HET JAARVERSLAG DER GOVERNEMENTS KINA-ONDERNEMING. Cultuurgids 5: 967-973.
- (2) ————
1905. ZIEKTEN EN PLAGEN IN DE GOVERNEMENTS KINA TUINEN. Cultuurgids 6: 1012-1013.
- (3) ARENS, P.
1910. HET INSTERVEN VAN HEVEA. AANTEEKENINGEN OVER LASIODIPLODIA THEOBROMAE, GRIFF. ET MAUBL. (BOTRYODIPLODIA ELASTICAE PETCH.). Cultuurgids 12: 276-281.
- (4) ASHBY, S. F.
1927. MACROPHOMINA PHASEOLI (MAUBL.) COMB. NOV. THE PYCNIDIAL STAGE OF RHIZOCTONIA BATATICOLA (TAUBL.) BUTL. Brit. Mycol. Soc. Trans. 12: 141-147, illus.
- (5) AUGUSTO, H.
1943. LA QUINA EN EL PERÚ. II. B. CULTIVO. Vida Agr. 20: 299-301, 303-308, illus.
- (6) BARAT, H.
1931. II. LABORATOIRE DE CRYPTOLOGIE. ÉTUDES DE LA DIVISION DE PHYTOPATHOLOGIE (SECTION SUD-INDOCHINOISE DE L'INSTITUT DES RECHERCHES AGRONOMIQUES) AU COURS DE L'ANNÉE 1930. Bul. Écon. de l'Indochine 34: 779B-796B.

- (7) BARAT, H.
1937. LA SÉLECTION DU CINCHONA LEDGERIANA EN INDOCHINE. Agron. Colon. 26 (239): 138-150.
- (8) BARBER, C. A.
1901. A TEA-EELWORM DISEASE IN SOUTH INDIA. [Madras] Dept. Land and Agr. Branch Bul. 2 (45): 227-234, illus.
- (9) BATAVIA, LANDBOUW SYNDICAAT.
1941. VERSLAGEN PROEFSTATIONS. Verslag. Landb. Synd., Batavia 1940: 154-282. [Abstract in Rev. Appl. Mycol. 23: 289-291. 1944.]
- (10) BERKHOUT, [A. H.]
1887. NASCHRIFT. Tijdschr. v. Nijv. Landb. Nederland. Indië 35: 213-216.
- (11) BERNARD, C.
1925. SEPTOBASIDIUM BOGORIENSE OP JONGE KINABOOMEN. Cinchona 2: 46-48, illus.
- (12) BESSEY, E. A.
1911. ROOT-KNOT AND ITS CONTROL. U. S. Dept. Agr. Bur. Plant Indus. Bul. 217, 89 pp., illus.
- (13) BLÜCHER, G. L. A. VON.
1938. ANBAU DER CHINARINDE (CINCHONA) IN NIEDERLÄNDISCH-INDIEN. Tropenpflanzer 41: 231-245, illus. [Transl. by Burns, W., Cinchona Cultivation in the Netherlands-Indies. Indian Farming 1: 311-317. 1940.]
- (14) BOECOP, M. VAN.
1929. SCHADUW IN KINA-PLANTSOENEN. Bergcultures 3: 1532-1533.
- (15) B[OECOP], [M.] v[AN].
1930. WORTELKRAAG-ZIEKTE OP SUCCIRUBRA-ZAADBEDDEN. Bergcultures 4: 218.
- (16) BOEDIJN, K. B., and STEINMANN, A.
1930. OVER DE OP THEE EN ANDERE CULTUURPLANTEN IN NED.-INDIË OPTREDENDE HELICOBASIDIUM- EN SEPTOBASIDIUM-SOORTEN. Arch. v. Theecult. Nederland. Indië 4(1): 5-59, illus. [English translation, pp. 37-59.]
- (17) ——— and STEINMANN, A.
1931. LES ESPÈCES DES GENRES HELICOBASIDIUM ET SEPTOBASIDIUM DES INDIES NÉERLANDAISES. Buitenzorg Jard. Bot. Bul. (ser. 3) 11: 165-219, illus.
- (18) BONDARZEWA-MONTEVERDE, V. N., GUTNER, L. S., and NOVOSSELOVA, E. D.
1936. DIE PARASITÄREN PILZE IN DEN GEWÄCHSHÄUSERN DES BOTANISCHEN INSTITUTES DER AKADEMIE DER WISSENSCHAFTEN DER USSR. Acta Inst. Bot. Acad. Sci. URSS ser. 2. Plantae Cryptogamae, fasc. 3, pp. 715-802, illus. [In Russian. German summary, p. 802.]
- (19) BOYCE, J. S.
1938. FOREST PATHOLOGY. 600 pp., illus. New York and London.
- (20) BRITON-JONES, H. R.
1934. THE DISEASES AND CURING OF CACAO. 161 pp., illus. London.
- (21) BROOKS, F. T.
1928. PLANT DISEASES. 386 pp., illus. London.
- (22) BROWN, W. H.
1930. RESEARCH AND FIELD WORK. Philippine Dept. Agr. and Nat. Resources Bur. Sci. Ann. Rpt. Dir. (1929) 28: 26-29.
- (23) ———
1931. RESEARCH AND FIELD WORK. Philippine Dept. Agr. and Nat. Resources Bur. Sci. Ann. Rpt. Dir. (1930) 29: 821-824.
- (24) BUGNICOURT, F.
1932. TRAVAUX DE CRYPTO GAMIE—RAPPORT SUR LE FONCTIONNEMENT DE LA DIVISION DE PHYTOPATHOLOGIE PENDANT L'ANNÉE 1931. (SECTION SUD-INDOCHINOISE DE L'INSTITUT DES RECHERCHES AGRONOMIQUES). Bul. Écon. de l'Indochine 35: 476B-514B.
- (25) ———
1939. LES FUSARIUM ET CYLINDROCARPON DE L'INDOCHINE. Encyclopédie Mycologique, v. 11, 206 pp., illus. Paris.

- (26) BURT, E. A.
1926. THE THELEPORACEAE OF NORTH AMERICA. XV. Mo. Bot. Gard. Ann. 13: 173-354, illus.
- (27) BUTLER, E. J.
1918. FUNGI AND DISEASE IN PLANTS. 547 pp., illus. Calcutta.
- (28) ———
1928. REPORT ON SOME DISEASES OF TEA AND TOBACCO IN NYASALAND. 30 pp., illus. Zomba.
- (29) ——— and BISBY, G. R.
1931. THE FUNGI OF INDIA. [India] Imp. Council Agr. Res. Sci. Monog. 1, 237 pp.
- (30) CALDER, C. C.
1928. SIXTY-SIXTH ANNUAL REPORT OF THE GOVERNMENT CINCHONA PLANTATIONS AND FACTORY IN BENGAL FOR THE YEARS 1927-28. 22 pp. Calcutta.
- (31) ———
1929. SIXTY-SEVENTH ANNUAL REPORT OF THE GOVERNMENT CINCHONA PLANTATIONS AND FACTORY IN BENGAL FOR THE YEAR 1928-29. 23 pp. Calcutta.
- (32) ———
1930. SIXTY-EIGHTH ANNUAL REPORT OF THE GOVERNMENT CINCHONA PLANTATIONS AND FACTORY IN BENGAL FOR THE YEAR 1929-30. 23 pp. Calcutta.
- (33) ———
1932. SEVENTIETH ANNUAL REPORT OF THE GOVERNMENT CINCHONA PLANTATIONS AND FACTORY IN BENGAL FOR THE YEAR 1931-32. 23 pp. Calcutta.
- (34) ———
1934. SEVENTY-FIRST ANNUAL REPORT OF THE GOVERNMENT CINCHONA PLANTATIONS AND FACTORY IN BENGAL FOR THE YEAR 1932-33. 23 pp. Calcutta.
- (35) ———
1936. SEVENTY-FOURTH ANNUAL REPORT OF THE GOVERNMENT CINCHONA PLANTATIONS AND FACTORY IN BENGAL FOR THE YEAR 1935-36. 25 pp. Calcutta.
- (36) ———
[c.1924.] REPORT OF THE BOTANICAL SURVEY OF INDIA FOR 1923-24. 9 pp. Calcutta.
- (37) ———
[c.1925.] REPORT OF THE BOTANICAL SURVEY OF INDIA FOR 1924-25. 10 pp. Calcutta.
- (38) ———
[c.1929.] REPORT OF THE BOTANICAL SURVEY OF INDIA FOR 1928-29. 9 pp. Calcutta.
- (39) CELINO, M. S.
1934-35. BLIGHT OF CINCHONA SEEDLINGS. Philippine Agr. 23: 111-127, illus.
- (40) CEYLON DEPARTMENT OF AGRICULTURE, DIVISION OF BOTANY AND MYCOLOGY.
1922. A PRELIMINARY LIST OF THE DISEASES OF CULTIVATED PLANTS IN CEYLON. Ceylon Dept. Agr. Bul. 52, 24 pp.
- (41) CEYLON DEPARTMENT OF AGRICULTURE, DIVISION OF MYCOLOGY.
1928. A LIST OF THE FUNGI ASSOCIATED WITH DISEASES OF CULTIVATED PLANTS IN CEYLON. Ceylon Dept. Agr. Bul. 83, 41 pp.
- (42) COBB, N. A.
1930. *TYLENCHUS ALATUS* N. SP. In Report of the Harvard African Expedition: The African Republic of Liberia and the Belgian Congo. Harv. Univ. Inst. Trop. Biol. and Med. Contrib. 5, pt. 1: 487-489. Cambridge.
- (43) COMMELIN, J. W.
1910. ZIEKTEN IN KINA-KWEEKBEDDEN. Cultuurgids 12: 106-110.
- (44) COSTER, C.
1941. HET WERK VAN HET PROEFSTATION WEST-JAVA IN 1940. Bergcultures 15: 1124-1133.

- (45) COSTER, C.
1942. THE WORK OF THE WEST-JAVA RESEARCH INSTITUTE, BUITENZORG, 1938-41. *Empire Jour. Expt. Agr.* 10: 22-30. [Reprinted in Honig, P., and Verdoorn, F., ed., *Science and Scientists in the Netherlands Indies*. *Natuurk. Tijdschr. V. Nederland Indië*. Sp. Sup. 102: 55-59. 1945.]
- (46) COUCH, J. N.
1938. THE GENUS SEPTOBASIDIUM. 480 pp., illus. Chapel Hill.
- (47) COWAN, J. M.
[1926.] REPORT OF THE BOTANICAL SURVEY OF INDIA FOR 1925-26. 9 pp. Calcutta.
- (48) ———
[1927.] REPORT OF THE BOTANICAL SURVEY OF INDIA FOR 1926-27. 10 pp. Calcutta.
- (49) CRANDALL, B. S., and DAVIS, W. C.
1945. PHYTOPHTHORA WILT AND STEM CANCER OF CINCHONA. *Phytopathology* 35: 138-140, illus.
- (50) DAVIS, W. C., WRIGHT, E., and HARTLEY, C.
1942. DISEASES OF FOREST-TREE NURSERY STOCK. U. S. Fed. Security Agency, Civ. Conserv. Corps Forestry Pub. 9, 79 pp., illus.
- (51) DUGGAR, B. M.
1915. RHIZOCTONIA CROCORUM (PERS.) DC. AND R. SOLANI KÜHN (CORTICIUM VAGUM B. AND C.), WITH NOTES ON OTHER SPECIES. *Mo. Bot. Gard. Ann.* 2: 403-458, illus.
- (52) DU PASQUIER, R.
1933. PRINCIPALES MALADIES PARASITAIRES DU THÉIER ET DU CAFÉIER EN EXTRÊ-ORIENT. *Bul. Econ. de l'Indochine* (n. s.) 36: 1-144, illus.
- (53) [DUTCH EAST INDIES] DEPARTEMENT VAN LANDBOUW, NIJVERHEID, EN HANDEL.
1911. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies] Dept. van Landb., Nijv. en Handel Jaarb. 1910: 176-227, illus.
- (54) ———
1916. VERSLAG VAN DE GOUVERNEMENTS KINA-ONDERNEMING TE TJINJIROEAN (BANDOENG) OVER 1915. 50 pp., illus. Bandoeng.
- (55) ———
[1917.] VERSLAG VAN DE GOUVERNEMENTS KINA-ONDERNEMING TE TJINJIROEAN (BANDOENG) OVER 1916. 75 pp., illus. Bandoeng.
- (56) ———
1918. VERSLAG VAN DE GOUVERNEMENTS KINA-ONDERNEMING TE TJINJIROEAN (BANDOENG) OVER 1917. 56 pp., illus. Bandoeng.
- (57) ———
1919. VERSLAG OMTRENT DE GOUVERNEMENTS KINA-ONDERNEMING TE TJINJIROEAN (BANDOENG) OVER 1918. 54 pp., illus. Bandoeng.
- (58) ———
1920. VERSLAG OMTRENT DE GOUVERNEMENTS KINA-ONDERNEMING TE TJINJIROEAN (BANDOENG) OVER 1919. 56 pp., illus. Bandoeng.
- (59) ———
1922. VERSLAG OMTRENT DE GOUVERNEMENTS KINA-ONDERNEMING TE TJINJIROEAN (BANDOENG) OVER 1920. 56 pp., illus. Bandoeng.
- (60) ———
1922. VERSLAG OMTRENT DE GOUVERNEMENTS KINA-ONDERNEMING TE TJINJIROEAN (PENGALENGAN) OVER 1921. 47 pp., illus. Bandoeng.
- (61) ———
1923. VERSLAG OMTRENT DE GOUVERNEMENTS KINA-ONDERNEMING TE TJINJIROEAN (PENGALENGAN) OVER 1922. 53 pp., illus. Bandoeng.

- (62) [Dutch East Indies] DEPARTEMENT VAN LANDBOUW, NIJVERHEID, EN
HANDEL.
1924. VERSLAG OMTRENT DE GOUVERNEMENTS KINA-ONDERNEMING TE
TJINJIROEAN (PENGALENGAN) OVER 1923. 44 pp. Wel-
tevreden.
- (63) ———
1925. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies]
Dept. van Landb., Nijv. en Handel Jaarb. 1924: 128-131.
- (64) ———
1925. VERSLAG OMTRENT DE GOUVERNEMENTS KINA-ONDERNEMING TE
TJINJIROEAN (PENGALENGAN) OVER 1924. 48 pp. Wel-
tevreden.
- (65) ———
1926. VERSLAG OMTRENT DE GOUVERNEMENTS KINA-ONDERNEMING TE
TJINJIROEAN (PENGALENGAN) OVER 1925. 48 pp. Wel-
tevreden.
- (66) ———
1927. GOUVERNEMENTS KINA- EN THEE-ONDERNEMING. [Dutch East
Indies] Dept. van Landb., Nijv. en Handel Jaarb. 1926:
102-108.
- (67) ———
1927. VERSLAG OMTRENT DE GOUVERNEMENTS KINA-ONDERNEMING TE
TJINJIROEAN (PENGALENGAN) OVER 1926. 49 pp. Wel-
tevreden.
- (68) ———
1928. 'S-LANDS KINA- EN THEE-ONDERNEMING. [Dutch East Indies]
Dept. van Landb., Nijv. en Handel Jaarb. 1927: 124-132.
- (69) ———
1928. VERSLAG OMTRENT DE GOUVERNEMENTS KINA EN THEE-ONDERNEM-
ING TE TJINJIROEAN (PENGALENGAN) OVER 1927. 51 pp.
Weltevreden.
- (70) ———
1929. VERSLAG OMTRENT DE GOUVERNEMENTS KINA- EN THEE-
ONDERNEMING TE TJINJIROEAN (PENGALENGAN) OVER 1928.
49 pp. Bandoeng.
- (71) ———
1931. VERSLAG OMTRENT 'S LANDS KINA- EN THEE-ONDERNEMING TE
TJINJIROEAN (PENGALENGAN) OVER 1929. 49 pp. Bandoeng.
- (72) ———
1931. VERSLAG OMTRENT 'S LANDS KINA- EN THEE-BEDRIJF TE
TJINJIROEAN (PENGALENGAN) OVER 1930. 62 pp. Bandoeng.
- (73) EEKHOUT, G. W.
1886. DE HEER KESSLER OVER KINA. Tijdschr. v. Nijv. Landb. Neder-
land. Indië 33: 539-542.
- (74) ELLIS, J. B., and EVERHART, B. M.
1887. ADDITIONS TO CERCOSPORA, GLOEOSPORIUM AND CYLINDROSPORIUM.
Jour. Mycol. 3: 13-22.
- (75) ENGLER, A., and PRANTL, K.
1897. DIE NATÜRLICHEN PFLANZENFAMILIEN. v. 1, pt. 1, 513 pp., illus.
Leipzig.
- (76) FÉE, A. L. A.
1824. ESSAI SUR LES CRYPTOGRAPHES DES ÉCORCES EXOTIQUES OFFICINALES.
167 pp., illus. Paris.
- (77) ———
[1837.] SUPPLÉMENT A L'ESSAI SUR LES CRYPTOGRAPHES DES ÉCORCES
EXOTIQUES OFFICINALES. 178 pp., illus. Paris.
- (78) FEILDEN, G. S. C., and GARNER, R. J.
1940. VEGETATIVE PROPAGATION OF TROPICAL AND SUB-TROPICAL
PLANTATION CROPS. Imp. Bur. Hort. and Plantation Crops
Tech. Commun. 13, 99 pp., illus.
- (79) FILIPJEV, I. N.
1936. ON THE CLASSIFICATION OF THE TYLENCHINAE. Helminthol.
Soc. Wash. Proc. 3: 80-82.

- (80) FLUITER, H. J. DE, and MULHOLLAND, J. J.
1941. GEGEVENS, VERKREGEN BIJ HET ONDERZOEK NAAR DE WAARD-
PLANTEN VAN TYLENCHUS COFFEAEE. *Bergcultures* 15: 1588-
1593.
- (81) FOSBERG, F. R.
1946. GIFTS OF THE AMERICAS—QUININE. *U. S. For. Agr. Relat., Agr.*
Amer. 6: [91].
- (82) FÜRTH, J.
1888. DE ZWARTE SCHIMMEL OP SUCCIRUBRABOOMEN; EENE STUDIE.
Tijdschr. v. Nijv. Landb. Nederland. Indië 37: 673-678.
- (83) GALLOWAY, L. D.
1936. REPORT OF THE IMPERIAL MYCOLOGIST. *Pusa Imp. Inst. Agr.*
Res. Sci. Rpts. 1934-35: 120-140.
- (84) ———
1937. REPORT OF THE IMPERIAL MYCOLOGIST. *New Delhi Imp. Inst.*
Agr. Res. Sci. Rpts. 1936: 105-111.
- (85) GÄUMANN, E. A.
1922. ENKELE GEGEVENS OMTRENT DE GRIJZE DADAPSCHIMMEL. *De Thee*
3: 48-50, illus.
- (86) ———
1922. ÜBER DAS SEPTOBASIDIUM BOGORIENSE PAT. *Ann. Mycol.* 20:
160-173, illus.
- (87) GOODEY, T.
1933. PLANT PARASITIC NEMATODES AND THE DISEASES THEY CAUSE.
306 pp., illus. London.
- (88) GOOT, P. VAN DER.
1928. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-
INDIË IN 1927. *Buitenzorg Inst. v. Plantenziekten Meded.*
74, 85 pp.
- (89) ———
1934. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-
INDIË IN 1932. *Buitenzorg Inst. v. Plantenziekten Meded.*
83, 80 pp.
- (90) ———
1935. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-
INDIË IN 1933. *Buitenzorg Inst. v. Plantenziekten Meded.*
84, 79 pp.
- (91) ———
1935. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-
INDIË IN 1934. *Buitenzorg Inst. v. Plantenziekten Meded.*
85, 94 pp.
- (92) ———
1936. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-
INDIË IN 1935. *Buitenzorg Inst. v. Plantenziekten Meded.*
87, 106 pp.
- (93) ———
1937. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-
INDIË IN 1936. *Buitenzorg Inst. v. Plantenziekten Meded.*
89, 104 pp.
- (94) GORKOM, K. W. VAN.
1878. DE ZIEKTE DER KINA-PLANT OP JAVA. *K. Akad. van Wetensch. te*
Amsterdam, Afd. Natuurk., Verslag. en Meded. 13 (reeks
2): 25-38.
- (95) ———
1888. EEN NIEUWE PLAAG DER KINABOOMEN. *Indische Mercur* 11:
355. [For corrections see *Indische Mercur* 11: 371. 1888.]
- (96) ———
1891. DE WORTELZIEKTE DER KINA EN DE GOUVERNEMENTS—RAPPORTEN.
Indische Mercur 14: 507.
- (97) ———
1908. SCHEIKUNDIGE BIJDAGEN TOT DE KENNIS DER JAVA-KINA 1872-
1907. 136 pp. Amsterdam.

- (98) GORKOM, K. W. VAN.
1925. KINA. *His Koloniaal Museum. Afdeeling: Voortbrengselen van de Groote Cultuur in Nederlandsch Oostindië* 5, 156 pp. Haarlem.
- (99) [GREAT BRITAIN IMPERIAL INSTITUTE.]
1939. THE WORLD'S CINCHONA BARK INDUSTRY, I AND II. [Gt. Brit.] *Imp. Inst. Bul.* 37: 18-31, 183-196.
- (100) GROOTHOFF, A.
1925. DE KINACULTUUR. *Onze Kol. Landb.; Twaalf Pop. Handb. over Nederland-Indische Landb.-Producten* 3, ed. 3, 122 pp., illus. Haarlem.
- (101) GUILLAUME, M.
1924. LE QUINQUINA À JAVA. *Agron Colon. (n. s.)* 11: 113-117, 137-146, 179-184.
- (102) HALL, C. J. J. VAN.
1916. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1915. *Buitenzorg Inst. v. Plantenziekten Meded.* 20, 47 pp.
- (103) ———
1917. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1916. *Buitenzorg Inst. v. Plantenziekten Meded.* 29, 37 pp.
- (104) ———
1918. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1917. *Buitenzorg Inst. v. Plantenziekten Meded.* 33, 42 pp.
- (105) ———
1919. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1918. *Buitenzorg Inst. v. Plantenziekten Meded.* 36, 49 pp.
- (106) ———
1920. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1919. *Buitenzorg Inst. v. Plantenziekten Meded.* 39, 50 pp.
- (107) ———
1921. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1920. *Buitenzorg Inst. v. Plantenziekten Meded.* 46, 50 pp.
- (108) ———
1922. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1921. *Buitenzorg Inst. v. Plantenziekten Meded.* 53, 46 pp.
- (109) ———
[1923.] ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1922. *Buitenzorg Inst. v. Plantenziekten Meded.* 58, 42 pp.
- (110) ———
1924. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1923. *Buitenzorg Inst. v. Plantenziekten Meded.* 64, 47 pp.
- (111) ———
1925. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1924. *Buitenzorg Inst. v. Plantenziekten Meded.* 67, 53 pp.
- (112) ———
1926. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1925. *Buitenzorg Inst. v. Plantenziekten Meded.* 70, 51 pp.
- (113) HANSFORD, C. G.
1937. ANNOTATED HOST LIST OF UGANDA PARASITIC FUNGI AND PLANT DISEASES.—PART IV. *East African Agr. Jour.* 3: 235-240.

- (114) HANSFORD, C. G.
1941. CONTRIBUTIONS TOWARDS THE FUNGUS FLORA OF UGANDA—III. SOME UGANDA ASCOMYCETES. Linn. Soc. London Proc. (1940-41) 153(1): 4-52, illus.
- (115) ———
1943. CONTRIBUTIONS TOWARDS THE FUNGUS FLORA OF UGANDA—V. FUNGI IMPERFECTI. Linn. Soc. London Proc. (1942-43) 155(1): 34-67, illus.
- (116) ———
1943. HOST LIST OF THE PARASITIC FUNGI OF UGANDA. PART II. East African Agr. Jour. 9: 50-55.
- (117) HARPER, R. E.
1943. CINCHONA PRODUCTION. Puerto Rico (Mayaguez) [Fed.] Expt. Sta. Rpt. 1942: 3-7.
- (118) ——— and WINTERS, H. F.
1946. CINCHONA INVESTIGATIONS IN PUERTO RICO. U. S. For. Agr. Relat., Agr. Amer. 6: 30-32, 37, illus.
- (119) HARTLEY, C.
1921. DAMPING-OFF IN FOREST NURSERIES. U. S. Dept. Agr. Dept. Bul. 934, 99 pp., illus.
- (120) HEIM, R.
1940. UN AGARIC RHIZOMORPHIQUE PARASITE DES SEMIS DE QUINQUINA EN HAUTE-GUINÉE. Rev. de Bot. Appl. et d'Agr. Trop. 20: 77-87, illus.
- (121) HENDRICKX, F. L.
1935-36. LISTE ANNOTÉE DES ORGANISMES VÉGÉTAUX SIGNALÉS SUR LE GENRE COFFEA. EN ANNEXE: MALADIES PHYSIOLOGIQUES OU D'ORIGINE MAL CONNUE. Ann. de Gembloux 41: 408-419, 446-465; 42: 20-25.
- (122) ———
1939. OBSERVATIONS PHYTOPATHOLOGIQUES A LA STATION DE MULUNGU EN 1938. Inst. Natl. pour l'Étude Agron. Congo Belge Rap. Ann. pour l'Exercice 1938 (pt. 2) Hors Ser.: 117-128.
- (123) HENNINGS, P.
1893. FUNGI WARBURGIANI. Hedwigia 32: 216-227, illus.
- (124) ———
1900. FUNGI. Monsunia [Leipzig] 1: 1-38, illus.
- (125) ———
1902. FUNGI JAVANICI NOVI A CL. PROF. DR. ZIMMERMANN COLLECTI. Hedwigia 41: 140-149.
- (126) HEUBEL, G. A.
1938. BEKNOPT OVERZICHT VAN DE ONDERNEMINGSCULTURES IN HET RAYON ZUID-SUMATRA GEDURENDE 1937. Bergcultures 12: 678-689.
- (127) ———
1939. BEKNOPT OVERZICHT VAN DE ONDERNEMINGSCULTURES IN HET RAYON ZUID-SUMATRA GEDURENDE 1938. Bergcultures 13: 768-782, illus.
- (128) HEUSDEN, W. C. VAN.
1935. OVERZICHT VAN DE MEERJARIGE CULTURES IN HET RAYON ZUID-SUMATRA GEDURENDE 1934. Bergcultures 9: 214-222.
- (129) ———
1936. BEKNOPT OVERZICHT VAN DE MEERJARIGE CULTURES IN HET RAYON ZUID-SUMATRA GEDURENDE 1935. Bergcultures 10: 688-696.
- (130) HOOKER, J. D.
1879. CINCHONA-DISEASE IN JAMAICA. Kew Roy. Bot. Gard. Rpt. 1878: 31.
- (131) HUNGER, F. W. T.
1907. JAARVERSLAG VAN DEN DIRECTEUR VAN HET ALGEMEEN-PROEFSTATION OVER 1906. [Dutch East Indies] Alg.-Proefsta. te Salatiga Verslag 1906: 13-93.

- (132) HUNGER, F. W. T.
[c.1908.] JAARVERSLAG VAN DEN DIRECTEUR VAN HET ALGEMEEN-
PROEFSTATION OVER 1907. [Dutch East Indies] Alg.-
Proefsta. te Salatiga Verslag 1907: 13-171, illus.
- (133) IMPERIAL BUREAU OF AGRICULTURAL PARASITOLOGY.
1931. THE ROOT-INFESTING EELWORMS OF THE GENUS HETERODERA. 99
pp., illus. St. Albans, England.
- (134) INSTITUT NATIONAL POUR L'ÉTUDE AGRONOMIQUE DU CONGO BELGE.
1939. RAPPORT ANNUEL POUR L'EXERCICE 1938. Inst. Natl. pour
l'Étude Agron. Congo Belge Pubs. (pt. 1), Hors ser., 269
pp., illus.
- (135) ————
1943. RAPPORT POUR LES EXERCICES 1940 AND 1941. Inst. Natl. pour
l'Étude Agron. Congo Belge Pubs., Hors ser., 152 pp.
- (136) INTERNATIONAL INSTITUTE OF AGRICULTURE.
1937. DISCOVERIES AND CURRENT EVENTS. Internatl. Inst. Agr., In-
ternatl. Bul. Plant Protect. 11: 85M-87M.
- (137) JAMAICA BOTANICAL DEPARTMENT.
1887. DISEASES OR INJURIES IN*PLANTS. Jamaica Bot. Dept. Bul. 3,
pp. 4-5.
- (138) JENKINS, A. E.
1945. SCAB OF CINCHONA IN SOUTH AMERICA CAUSED BY ELSINOË.
Wash. Acad. Sci. Jour. 35: 344-352, illus.
- (139) KERBOSCH, M.
1917. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies]
Dept. van Landb., Nijv. en Handel Jaarb. 1915: 167-172.
- (140) ————
1918. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies]
Dept. van Landb., Nijv. en Handel Jaarb. 1916: 83-88.
- (141) ————
1919. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies]
Dept. van Landb., Nijv. en Handel Jaarb. 1917: 92-97.
- (142) ————
1920. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies]
Dept. van Landb., Nijv. en Handel Jaarb. 1918: 204-207.
- (143) ————
1920. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies]
Dept. van Landb., Nijv. en Handel Jaarb. 1919: 200-203.
- (144) ————
1923. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies]
Dept. van Landb., Nijv. en Handel Jaarb. 1922: 122-125.
- (145) ————
1929. 'S-LANDS KINA- EN THEE-ONDERNEMING. [Dutch East Indies]
Dept. van Landb., Nijv. en Handel Jaarb. 1928: 148-156.
- (146) KESSLER, A.
1886. EEN EN ANDER OVER KINA. Tijdschr. v. Nijv. Landb. Nederland.
Indië 33: 259-275.
- (147) KEUCHENIUS, A. A. M. N.
1936. OVERZICHT VAN DE MEERJARIGE CULTURES IN HET RAYON WEST-
SUMATRA GEDURENDE 1935. Bergcultures 10: 432-434.
- (148) ————
1937. OVERZICHT VAN DE ONDERNEMINGSCULTURES IN HET RAYON WEST-
SUMATRA GEDURENDE 1936. Bergcultures 11: 510-518.
- (149) ————
1938. OVERZICHT VAN DE ONDERNEMINGSCULTURES IN HET RAYON WEST-
SUMATRA GEDURENDE 1937. Bergcultures 12: 268-275.
- (150) ————
1939. OVERZICHT VAN DE ONDERNEMINGSCULTURES IN HET RAYON WEST-
SUMATRA GEDURENDE 1938. Bergcultures 13: 569-576.
- (151) KEVORKIAN, A. G.
1942. CINCHONA PRODUCTION. Puerto Rico (Mayaguez) [Fed.] Expt.
Sta. Rpt. 1941: 9-10.

- (152) KHESWALLA, K. F.
1935. SEEDLING BLIGHT OF CINCHONA LEDGERIANA MOENS CAUSED BY PHYTOPHTHORA PALMIVORA BUTL. IN THE DARJEELING DISTRICT. Indian Jour. Agr. Sci. 5: 485-495, illus.
- (153) KING, G.
1880. A MANUAL OF CINCHONA CULTIVATION IN INDIA. Ed. 2, 105 pp., illus. Calcutta.
- (154) KOORDERS, S. H.
1906. RESULTATEN VAN EEN VOORLOOPIG MIKROSKOPISCH ONDERZOEK EENER WORTELZIEKTE VAN JONGE KINAPLANTJES VEROORZAAKT DOOR HETERODERA-AALTJES EN EEN SCHIMMEL. Cultuurgids 7: 901-919, illus.
- (155) ———
1907. BOTANISCHE UNTERSUCHUNGEN ÜBER EINIGE IN JAVA VORKOMMENDE PILZE, BESONDERS ÜBER BLÄTTER BEWOHNENDE, PARASITISCH AUFTRETENDE ARTEN. K. Akad. Wetensch. te Amsterdam, Verhandel. Tweede sectie, deel 13, No. 4, 264 pp., illus.
- (156) KRISHNA AYYAR, P. N.
1933. SOME EXPERIMENTS ON THE CONTROL OF THE ROOT-GALL NEMATODE IN SOUTH INDIA. Madras Agr. Jour. 21: 97-107.
- (157) LEEFMANS, S.
1927. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1926. Buitenzorg Inst. v. Plantenziekten Meded. 73, 60 pp.
- (158) ———
1929. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1928. Buitenzorg Inst. v. Plantenziekten Meded. 75, 96 pp.
- (159) ———
1931. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH OOST-INDIË IN 1929. Buitenzorg Inst. v. Plantenziekten Meded. 79, 100 pp.
- (160) ———
1933. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH OOST-INDIË IN 1930. Buitenzorg Inst. v. Plantenziekten Meded. 81, 84 pp.
- (161) ———
1934. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH OOST-INDIË IN 1931. Buitenzorg Inst. v. Plantenziekten Meded. 82, 92 pp.
- (162) LEERSUM, P. VAN.
1891. IETS OVER DE WORTELZIEKTE DER KINABOOMEN Indische Cult. (Teysmannia) 2: 327-338.
- (163) ———
1891. OVER DEN INVLOED VAN VORST OP HET ALCALOÏD-GEHALTE DER KINABOOMEN. Indische Cult. (Teysmannia) 2: 675-681.
- (164) ———
1907. BERICHT OMTRENT DE GOUVERNEMENTS KINA-ONDERNEMING OVER HET 1° KWARTAAL 1907. Cultuurgids 9: 149-152.
- (165) ———
1908. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies] Dept. van Landb., Nijv. en Handel Jaarb. 1907: 226-298, illus.
- (166) ———
1908. OVER DEN Z. G. "STAMKANKER" BIJ KINE. Indische Cult. (Teysmannia) 19: 494-498.
- (167) ———
1909. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies] Dept. van Landb., Nijv. en Handel Jaarb. 1908: 147-226, illus.

- (168) LEERSUM, P. VAN.
1910. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies] Dept. van Landb., Nijv. en Handel Jaarb. 1909: 216-258, illus.
- (169) ———
1912. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies] Dept. van Landb., Nijv. en Handel Jaarb. 1911: 140-182, illus.
- (170) ———
1913. KINA. In Gorkom, K. W. van, Oost-Indische Cultures, deel 3, pp. 57-154, illus. Amsterdam.
- (171) ———
1914. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies] Dept. van Landb., Nijv. en Handel Jaarb. 1913: 189-195.
- (172) ———
1915. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies] Dept. van Landb., Nijv. en Handel Jaarb. 1914: 161-167.
- (173) LIENEMAN, C.
1929. A HOST INDEX TO THE NORTH AMERICAN SPECIES OF THE GENUS CERCOSPORA. Mo. Bot. Gard. Ann. 16: 1-52.
- (174) LOBLE, [M.]
1919. NOTE SUR LA CULTURE DU QUINQUINA À JAVA. Bul. Écon. de l'Indochine (n. s.) 22: 761-764.
- (175) LUDOWYKE, H. [Translator.]
1929. THE OCCURRENCE OF MYCORRHIZA IN CINCHONA IN JAVA. Trop. Agr. [Ceylon] 72: 281-283. With an addition by the translator, p. 281. [Transl. of Steinmann, A., Over het Optreden van Mycorrhiza bij Kina op Java. Cinchona 5: 27-29, illus. 1928.]
- (176) LUTZ, M. L.
1904. NOTES MYCOLOGIQUES. Soc. Mycol. de France, Bul. Trimest. 20: 211-213.
- (177) McRAE, W.
1928. REPORT OF THE IMPERIAL MYCOLOGIST. Pusa Imp. Inst. Agr. Res. Sci. Rpts. 1927-28: 56-70.
- (178) ———
1930. REPORT OF THE IMPERIAL MYCOLOGIST. Pusa Imp. Inst. Agr. Res. Sci. Rpts. 1928-29: 51-66.
- (179) ———
1932. REPORT OF THE IMPERIAL MYCOLOGIST. Pusa Imp. Inst. Agr. Res. Sci. Rpts. 1930-31: 73-86.
- (180) ———
1933. REPORT OF THE IMPERIAL MYCOLOGIST. Pusa Imp. Inst. Agr. Res. Sci. Rpts. 1931-32: 122-140.
- (181) ———
1934. REPORT OF THE IMPERIAL MYCOLOGIST. Pusa Imp. Inst. Agr. Res. Sci. Rpts. 1932-33: 134-151.
- (182) MADARANG, S. A.
1941. RHIZOCTONIA DAMPING-OFF OF CINCHONA SEEDLINGS. Philippine Jour. Forestry 4: 105-121, illus.
- (183) MARAÑON, J., and BARTLETT, H. H.
1941. CINCHONA CULTIVATION AND THE PRODUCTION OF TOTAQUINA IN THE PHILIPPINES. Philippine Univ. Nat. and Appl. Sci. Bul. 8: 111-142, illus.
- (184) MARCINOWSKI, K.
1909. PARASITISCH UND SEMIPARASITISCH AN PFLANZEN LEBENDE NEMATODEN. K. Biol. Anst. f. Land u. Forstw. Arb. 7 (1): 1-192, illus.
- (185) MARKHAM, C. R.
1880. PERUVIAN BARK. A POPULAR ACCOUNT OF THE INTRODUCTION OF CHINCHONA CULTIVATION INTO BRITISH INDIA. 550 pp., illus. London.

- (186) MARTIN, W. E., and CRANDALL, B. S.
1944. OBSERVACIONES SOBRE EL CULTIVO Y ENFERMEDADES DE LA CINCHONA EN LAS PLANTACIONES DE PUÑIZAS Y SU AVALUCIÓN, COMO FUENTE DE SEMILLA Y MATERIAL DE PROPAGACIÓN. Peru. Dir. de Asuntos Orientales, Colon. y Terrenos de Oriente. Colon. y Foresta 1: 4-15.
- (187) MITRA, M.
1925. REPORT OF THE IMPERIAL MYCOLOGIST. Pusa Imp. Inst. Agr. Res. Sci. Rpts. 1924-25: 45-57.
- (188) ———
1931. REPORT OF THE IMPERIAL MYCOLOGIST. Pusa Imp. Inst. Agr. Res. Sci. Rpts. 1929-30: 58-71.
- (189) MOENS, J. C. B.
1882. DE KINACULTUR IN AZIË, 1854-1882. 393 pp., illus. Batavia.
- (190) MOSER, C. K.
1942. CINCHONA FOR QUININE: VITAL NETHERLANDS INDIES PRODUCT. U. S. Dept. Com., For. Com. Weekly 6 (2): 6-7, 29, illus.
- (191) NOACK, F.
1903. KURZE MITTEILUNGEN ÜBER KRANKHEITEN TROPISCHER NUTZPFLANZEN. Ztschr. f. Pflanzenkrank. 13: 225-230.
- (192) ———
1907. KRANKHEITEN TROPISCHER NUTZPFLANZEN. Ztschr. f. Pflanzenkrank. 16: 90-100.
- (193) ———
1908. KRANKHEITEN TROPISCHER NUTZPFLANZEN. II. Ztschr. f. Pflanzenkrank. 17: 345-355.
- (194) NOWELL, W.
[n. d.] DISEASES OF CROP-PLANTS IN THE LESSER ANTILLES. 383 pp., illus. London.
- (195) OCFEMIA, G. O.
1934-35. BUD ROT OF COCONUT. Philippine Agr. 23: 4-10, illus.
- (196) ——— and CELINO, M. S.
1933. A BROWN BARK ROT OF CACAO TRUNK. Philippine Agr. 21: 665-673, illus.
- (197) OVEREEM, C. VAN.
1925. BEITRÄGE ZUR PILZFLORA VON NIEDERLÄNDISCH INDIEN. II. 13. UEBER DEN ROTEN WURZELPILZ. Buitenzorg Jard. Bot. Bul. (ser. 3) 7: 436-443, illus.
- (198) OWEN, T. C.
1881. THE CINCHONA PLANTER'S MANUAL. 203 pp. Colombo.
- (199) PADWICK, G. W.
1939. REPORT OF THE IMPERIAL MYCOLOGIST. New Delhi Imp. Inst. Agr. Res. Sci. Rpts. 1938: 105-112, illus.
- (200) ———
1940. REPORT OF THE IMPERIAL MYCOLOGIST. New Delhi Imp. Inst. Agr. Res. Sci. Rpts. 1939: 103-115.
- (201) PATOUILLARD, N., and LAGERHEIM, G. DE.
1892. CHAMPIGNONS DE L'EQUATEUR (PUBILLUS II). Soc. Mycol. de France, Bul. Trimest. 8: 113-140, illus.
- (202) PENNOCK, W.
1945. NOTES ON CINCHONA CULTURE. In Verdoorn, F., ed., Plants and Plant Science in Latin America, pp. 202-205. Waltham, Mass.
- (203) PETCH, T.
1912. PINK DISEASE IN JAVA. Trop. Agr. [Ceylon] 39: 44-45. [Review of Rant, A., Ueber die djamoer—oepas-krankheit und ueber das Corticium javanicum Zimm. Buitenzorg Jard. Bot. Bul., ser. 2, No. 4, 50 pp., illus. 1912.]
- (204) PETRI, L.
1931. RASSEGNA DEI CASI FITOPATHOLOGICI OSSERVATI NEL 1930. [Roma] R. Staz. di Patol. Veg. Bol. (n. s.) 11: 1-50.

- (205) PHILLIPE, G.
1919. RAPPORT SUR LA CULTURE DES ARBRES À QUINQUINA. 79 pp.
Lons-le-Saunier.
- (206) PINKUS, R.
1945. EL INJERTO DE LA CHINCHONA (CONTINUA). Rev. de Agr.
[Guatemala] 22: 135-140, illus. [Abstract in Rev. Appl.
Mycol. 24: 387. 1945.]
- (207) POPENOE, W.
1941. CULTIVO DE LA QUINA (CINCHONA) IN GUATEMALA. 39 pp., illus.
[Guatemala City.]
- (208) ———
1942. QUININE FROM THE "FEVER-TREE." U. S. For. Agr. Relat., Agr.
Amer. 2: 43-47, illus.
- (209) ———
1943. CINCHONA. Rev. del Inst. de Defensa del Café de Costa Rica
13: 379-399, illus.
- (210) POSNJAK, A. D.
1936. CINCHONA AND ITS CULTIVATION. [Leningrad] Inst. Zashch.
Rast. (Lenin Acad. Agr. Sci., U.S.S.R., Inst. Plant Protect.)
Sup. 81 to the Bul. of Appl. Bot., Genet., and Plant Breed-
ing. 172 pp., illus. [In Russian.]
- (211) PRILLWITZ, P. M.
1926. VERGELIJKENDE BESCHOUWINGEN OVER DE GRIJZE DADAPSSCHIM-
MEL EN HARE BESTRIJDING. De Thee 7: 91-94, illus.
- (212) P[RING], G. H.
1932. EFFECT OF CITY SMOKE ON PLANTS. Mo. Bot. Gard. Bul. 20:
43-45.
- (213) PRINTZ, H.
1940. VORARBEITEN ZU EINER MONOGRAPHIE DER TRENTÉPOHLIACEEN.
NYT Mag. 80: 137-210, illus.
- (214) PRUDHOMME, E.
1902. LE QUINQUINA, CULTURE, PRÉPARATION, COMMERCE. 84 pp., illus.
Paris.
- (215) PUERTO RICO (MAYAGUEZ) [FEDERAL] EXPERIMENT STATION.
1939. STUDIES OF QUININE PRODUCTION. Puerto Rico (Mayaguez)
[Fed.] Expt. Sta. Rpt. 1938: 26-29, illus.
- (216) ———
1940. INVESTIGATIONS OF DRUG AND SPICE PLANTS. Puerto Rico
(Mayaguez) [Fed.] Expt. Sta. Rpt. 1939: 40-44.
- (217) ———
1941. INVESTIGATIONS OF DRUG PLANTS. Puerto Rico (Mayaguez)
[Fed.] Expt. Sta. Rpt. 1940: 18-22, illus.
- (218) RACIBORSKI, M.
1909. NALISTNE I PARORZYTNE GRZYBY JAWY.—PARASTISCHE UND
EPIPHYTISCHE PILZE JAVA'S. Krakow. Akad. Umiejtnosci
Bul. Internatl. Cl. des Sci., Math. et Naturel., Ser. B, Sci.
Nuturel 1909: 346-394, illus.
- (219) RANDS, R. D.
1922. STREEPKANKER VAN KANEEL, VEROORZAAKT DOOR PHYTOPHTHORA
CINNAMOMI N. SP. Buitenzorg Inst. v. Plantenziekten
Meded. 54, 53 pp., illus.
- (220) RANT, A.
1908. KORTE AANTEKENINGEN OVER KINA. I. Indische Cult. (Teys-
mannia) 19: 431-435.
- (221) ———
1909. KORTE AANTEKENINGEN OVER KINA. III. Indische Cult. (Teys-
mannia) 20: 409-417, illus.
- (222) ———
1910. KORTE AANTEKENINGEN OVER KINA. IV. Indische Cult. (Teys-
mannia) 21: 777-780.

- (223) RANT, A.
1911. DE DJAMOER OEPAS-ZIEKTE IN HET ALGEMEEN EN BIJ KINA IN HET BIJZONDER. [Dutch East Indies] Dept. van Landb., Nijv. en Handel Meded. 13, 38 pp., illus. Batavia.
- (224) ———
1912. KORTE AANTEKENINGEN OVER KINA. v. Indische Cult. (Teysmannia) 23: 610-612, illus.
- (225) ———
1912. ÜBER DIE DJAMOER—OEPAS-KRANKHEIT UND ÜBER DAS CORTICIUM JAVANICUM ZIMM. Buitenzorg Jard. Bot. Bul., ser. 2, No. 4, 50 pp., illus.
- (226) ———
1913. BOTANISCHE ONDERZOEKINGEN. [Dutch East Indies] Dept. van Landb., Nijv. en Handel Jaarb. 1912: 152-153.
- (227) ———
1914. DE ZIEKTEN EN SCHIMMELS DER KINA. [Dutch East Indies] Kina Proefsta. Meded. 2, 47 pp., illus.
- (228) ———
1915. KORTE AANTEKENINGEN OVER KINA. VI. Indische Cult. (Teysmannia) 26: 54-57.
- (229) ———
1915. ÜBER DIE MOPOKRANKHEIT JUNGER CINCHONAPFLANZEN UND ÜBER DEN JAVANISCHEN VERMEHRUNGSPILZE. Buitenzorg Jard. Bot. Bul., ser. 2, No. 18, 21 pp., illus.
- (230) ———
1916. DER GRAUE WURZELPILZ VON CINCHONA. Buitenzorg Jard. Bot. Bul., ser. 2, No. 22, 22 pp., illus.
- (231) ———
1917. THE WHITE ROOT-FUNGUS OF CINCHONA. Rec. des Trav. Bot. Néerland. 14: 143-148, illus.
- (232) REHM, H.
1903. ASCOMYCETEN-STUDIEN. I. Hedwigia 42 (Beiblatt): (172)-(176).
- (233) ———
1908. ASCOMYCETES NOVI. Ann. Mycol. 6: 313-325.
- (234) REICHERT, I., and HELLINGER, E.
1932. ON BOTRYTIS TIP-END ROT OF BANANA FRUITS IN PALESTINE. Hadar 5: 162-163, illus.
- (235) RHIND, D.
1927. ANNUAL REPORT OF THE MYCOLOGIST, BURMA, FOR THE YEAR ENDING THE 30TH JUNE 1926. 7 pp. Rangoon.
- (236) RITZEMA BOS, J.
1926. BEKNOPTE AANTEKENINGEN OP PLANTENZIEKTENKUNDIG GEBIED. 56. SEPTOBASIDIUM BOGORIENSE PAT. OP JONGE KINABOOMEN. Tijdschr. over Plantenziekten 32: 303-304.
- (237) ROEPKE, W.
1911. VERDERE BIJDRAGEN TOT DE KENNIS VAN DE KINA-INSEKTEN. Cultuurgids 13: 1-14, illus.
- (238) ROGERS, D. P.
1933. TAXONOMIC NOTES ON THE TULASNELLACEAE. Iowa Univ. Studies in Nat. Hist. 15(3): 31-32, illus.
- (239) ROGGEN, M. A. VAN.
1922. GOUVERNEMENTS KINA-ONDERNEMING. [Dutch East Indies] Dept. van Landb., Nijv. en Handel Jaarb. 1920: 110-115.
- (240) RUTGERS, A. A. L.
1915. ZIEKTEN EN PLAGEN DER CULTUURGEWASSEN IN NEDERLANDSCH-INDIË IN 1914. Buitenzorg Inst. v. Plantenziekten 15, 45 pp.
- (241) S., J. L.
1877. CULTURE DU CINCHONA ET DU JALAP, À LA JAMAÏQUE. Jour. de Pharm. et de Chim. (ser. 4) 26: 215-216.

- (242) SACCARDO, P. A.
1882-1931. SYLLOGE FUNGORUM OMNIUM HUCUSQUE COGNITORUM. 25 vols.
- (243) ———
1911. NOTAE MYCOLOGICAE. Ann. Mycol. 9: 249-257.
- (244) SANDS, W. N.
1922. THE CINCHONA (QUININE) INDUSTRY IN JAVA. Trop. Agr. [Ceylon] 59: 352-370.
- (245) SAWADA, K.
1936. PHYTOPHTHORA BLIGHT OF CINCHONA SEEDLINGS OCCURRING IN FORMOSA, CAUSED BY PHYTOPHTHORA CINCHONAE SAW. N. SP. Formosan Agr. Rev. 32 (354): 326-346, illus. [In Japanese. Abstract in Rev. Appl. Mycol. 16: 408. 1937.]
- (246) SEN, S. C.
1935. SEVENTY-THIRD ANNUAL REPORT OF THE GOVERNMENT CINCHONA PLANTATIONS AND FACTORY IN BENGAL FOR THE YEAR 1934-35. 25 pp. Calcutta.
- (247) ———
1938. SEVENTY-SIXTH ANNUAL REPORT OF THE GOVERNMENT CINCHONA PLANTATIONS AND FACTORY IN BENGAL FOR THE YEAR 1937-38. 30 pp. Calcutta.
- (248) SEYMOUR, A. B.
1929. HOST INDEX OF THE FUNGI OF NORTH AMERICA. 732 pp. Cambridge.
- (249) SHARPLES, A.
1936. DISEASES AND PESTS OF THE RUBBER TREE. 480 pp., illus. London.
- (250) SMALL, W.
1925. ANNUAL REPORT OF THE GOVERNMENT MYCOLOGIST. Uganda Dept. Agr. Ann. Rpt. 1924: 18-20.
- (251) SPRUIT, C.
1923. CHINARINDEN (CINCHONA). In Fruwirth, K., Handbuch der Landwirtschaftlichen Pflanzenzüchtung. Aufl. 2, Bd. 5. Die Züchtung Kolonialer Gewächse, pp. 256-268, illus. Berlin.
- (252) ———
1927. VERSLAG OMTRENT HET OPTREDEN VAN ZIEKTEN EN PLAGEN IN DE KINACULTUUR GEDURENDE 1926. Cinchona 4: 10-13.
- (253) STEINMANN, A.
1926. EENIGE BESCHOUWINGEN OMTRENT VOORKOMEN EN BESTRIJDING VAN DE OP KINA OPTREDENDE SEPTOBASIDIUM-SOORTEN. Cinchona 3: 123-128, illus.
- (254) ———
1926. VOORLOOPIGE MEDEDEELING OMTRENT HET OPTREDEN VAN SEPTOBASIDIUM BOGORIENSE EN RUBIGINOSUM OP THEE. De Thee 7: 52-53, illus.
- (255) ———
1928. OVER HET OPTREDEN VAN MYCORRHIZA BIJ KINA OP JAVA. Cinchona 5: 27-29, illus.
- (256) ———
1928. VOORLOOPIGE MEDEDEELING OMTRENT HET OPTREDEN VAN RHIZOCTONIA BATATICOLA (TAUB.) BUTLER OP JAVA EN SUMATRA. Arch. v. Theecult. Nederland Indië 2: 74-86, illus. [English summary, pp. 79-80.]
- (257) ———
1929. BIJDRAGE TOT DE KENNIS DER WORTELZIEKTEN VAN DE KINA. Cinchona 6: 23-30, illus.
- (258) ———
1931. VERSLAG VAN DEN MYCOLOOG OVER 1930. Arch. v. Theecult. Nederland Indië 5: 161-164.
- (259) STEVENSON, J. A.
1926. FOREIGN PLANT DISEASES. U. S. Dept. Agr., 198 pp.

- (260) STEVENSON, J. A.
1943. FUNGI NOVI DENOMINATI—I. *Mycologia* 35: 629-637.
- (261) STODDARD, D. L.
1945. SEED TREATMENT. Puerto Rico (Mayaguez) [Fed.] Expt. Sta. Rpt. 1944: 28-29.
- (262) STOFFELS, E. H. J.
1945. LE QUINQUINA. Inst. Natl. pour l'Étude Agron. Congo Belge Pubs., Tech. Ser. 24a, 57 pp., illus. Ed. 2.
- (263) STROOMBERG, J.
1929. LA CULTURE DU QUINQUINA AUX INDES NÉERLANDAISES. Rev. Internatl. des Prod. Colon. 4: 290-295, illus.
- (264) SUBBA RAO, M. K.
1936. PINK DISEASE. United Planters' Assoc. South. India Tea Sci. Dept. (Madras) Bul. 10, 25 pp., illus.
- (265) SUNDARARAMAN, S.
1928. ADMINISTRATION REPORT OF THE GOVERNMENT MYCOLOGIST, COIMBATORE, FOR 1927-28. Madras Dept. Agr. Rpts. Sub. Off. 1927-28: 355-372.
- (266) SYDOW, P., and SYDOW, H.
1924. UREDO CINCHONAE. MONOGRAPHIA UREDINEARUM SEU SPECIEFUM OMNIUM AD HUNC USQUE DIEM COGNITARUM DESCRIPTIO ET ADUMBRATIO SYSTEMATICA. v. 4, fasc. 4, p. 561. Leipzig.
- (267) TASSI, F.
1900. NOVAE MICROMYCETUM SPECIES DESCRIPTAE ET ICONIBUS ILLUSTRATAE. Lab. Orto Bot. R. Univ. Studi Siena Bul. 3: 117-132, illus.
- (268) THOMAS, A. S.
1946. CINCHONA IN UGANDA. Empire Jour. Expt. Agr. 14(54): 75-84, illus.
- (269) THOMPSON, A.
1940. NOTES ON PLANT DISEASES IN 1939. Malayan Agr. Jour. 28: 400-407, illus.
- (270) ———
1941. NOTES ON PLANT DISEASES IN 1940. Malayan Agr. Jour. 29: 241-245.
- (271) TSCHIRCH, A.
1923. HANDBUCH DER PHARMAKOLOGIE. Ed. 2, Bd. 3, t. 1, 748 pp. Leipzig.
- (272) TUCKER, C. M.
1931. TAXONOMY OF THE GENUS PHYTOPHTHORA DE BARY. Mo. Agr. Expt. Sta. Res. Bul. 153, 208 pp., illus.
- (273) VINCENS, F.
1921. RAPPORT SOMMAIRE SUR LES TRAVAUX EFFECTUÉS AU LABORATOIRE DE PHYTOPATHOLOGIE DE L'INSTITUT SCIENTIFIQUE DE L'INDOCHINE DU 1^{er} JANVIER 1919 AU 1^{er} JUILLET 1921. Inst. Sci. de Saigon Bul. Agr. 3: 307-323.
- (274) ———
1922. MALADIES DES JEUNES PLANTS ET CHAMPIGNONS MICROSCOPIQUES NOUVEAUX OBSERVÉS SUR CINCHONA EN INDOCHINE. Soc. de Path. Vég. France, Bul. 9: 125-133, illus.
- (275) WALKER, E. M.
1939. "PINK DISEASE" IN RUBBER. ITS SYMPTOMS, EFFECTS AND PREVENTION. Planters' Chron. 34: 664-666.
- (276) [WALLACE, G. B.]
[1929.] DISEASES OF PLANTS. Tanganyika Dept. Agr. Rpt. 1928: 40-42.
- (277) WALLACE, G. B.
1932. III.—PRELIMINARY LIST OF FUNGI OR DISEASES OF ECONOMIC PLANTS IN TANGANYIKA TERRITORY. Kew Roy. Bot. Gard. Bul. Misc. Inform. 1932 (1): 28-40.

- (278) WALLACE, G. B.
1935-36. ARMILLARIA ROOT ROT IN EAST AFRICA. ARMILLARIA MELLEA (VAHL) QUÉL. East African Agr. Jour. 1: 182-192, illus.
- (279) WARBURG, O.
1887. BEITRAG ZUR KENNTNISS DER KREBSKRANKHEIT DER KINA BÄUME AUF JAVA. Ber. über die Sitzungen der Gesell. f. Bot. zu Hamburg, heft 3, pp. 62-72.
- (280) ———
1887. BIJDRAGE TOT DE KENNIS VAN DEN KANKER DER KINABOOMEN. Tijdschr. v. Nijv. Landb. Nederland. Indië 35: 195-213.
- (281) WEBER, G. F.
1931. BLIGHT OF CARROTS CAUSED BY SCLEROTIUM ROLFSSII, WITH GEOGRAPHIC DISTRIBUTION AND HOST RANGE OF THE FUNGUS. Phytopathology 21: 1129-1140, illus.
- (282) WESTERDIJK, J.
1915. PHYTOPATHOLOGY IN THE TROPICS. Mo. Bot. Gard. Ann. 2: 307-313.
- (283) WILSON, C. M.
1943. QUININE—REBORN IN OUR HEMISPHERE. Harper's Mag. 187: 275-280.
- (284) WOLLENWEBER, H. W.
1926. PYRENOAMYCETEN-STUDIEN. II. Angew. Bot. 8: 168-212, illus.
- (285) ——— and REINKING, O. A.
1935. DIE FUSARIEN IHRE BESCHREIBUNG, SCHADWIRKUNG UND BEKÄMPFUNG. 355 pp., illus. Berlin.
- (286) WURTH, T.
1905. OVER EENE OP KINA GEVONDENE SLIJMZWAM. Cultuurgids 7: 626-630, illus.
- (287) ZEHNTNER, L.
1905. EENIGE WAARNEMINGEN OVER DE DJAMOER OEPAS ZIEKTE, VEROORZAAKT DOOR CORTICIUM JAVANICUM ZIMM. Cultuurgids 7: 401-420, illus.
- (288) ZENKER.
1827-29. KRYPTOGAMISCHE PARASITEN AUF OFFICINELLEN RINDEN. In Goebel, F., and Kunze, G., Pharmaceutische Waarenkunde. v. 1, pp. 109-200, illus. Eisenach.
- (289) ZIMMERMANN, A.
1904. UNTERSUCHUNGEN ÜBER TROPISCHE PFLANZENKRANKHEITEN. Deut. Ost Afrika Ber. über Land. u. Forstw. 2: 11-36, illus.
- (290) ———
1905. NEWER METHODS OF CINCHONA CULTIVATION. Drug Cir. 49: 423-426, illus.

INDEX

	Page		Page
Africa	21	<i>Cercospora cinchonae</i>	22
Algal leaf spot:		Ceylon	30
brown	22	<i>Chaetomium</i>	46
<i>Cephaleuros</i>	21	Chloride of lime, control of	
green	22	stem canker	17
<i>Alternaria</i>	3	Chlorophyll deficiency	25
Anthraxnose leaf spot	10	Chlorosis	25
<i>Armillaria</i> —		Chroolepidaceae	22
<i>mellea</i>	37, 39	Chytridiales	17
root rot	37	Cinnamon, host for <i>Corticium</i>	
Aromatic nitrated phenols, control of clitocybe damping-off	4	<i>salmonicolor</i>	30
		Citrus species, hosts for	
		<i>Corticium salmonicolor</i>	30
		<i>Cladosporium herbarum</i>	3
Bark diseases	27	Clitocybe	4
Belgian Congo	3, 7, 8, 21, 29, 30, 37, 39, 41, 46	Coffee, host for <i>Corticium</i>	
Bleeding disease	35	<i>salmonicolor</i>	30, 31
Bolivia	41	Collar and root rot	36
Bordeaux mixture, control of—		<i>Colletotrichum</i> —	
damping-off	7	<i>cinchonae</i>	9
gloeosporium leaf spot	10	leaf spot	9
phyllostictina leaf spot	11	Colombia	21, 26, 30, 33, 43
phytophthora stem blight	13, 15, 16	Commercial barks, fungi on ..	47
sclerotinia leaf disease	24	Copper fungicide, control of	
<i>Botryodiplodia</i> —		<i>cladosporium</i> damping-off ..	3
species	13, 30	Copper spray, control of—	
<i>theobromae</i>	35	nursery leaf disease	11
<i>Botrytis cinerea</i>	3, 5, 24, 46	phytophthora stem blight ..	16
Branch and trunk canker	29	Copper sulfate, control of	
Brazil	47	<i>hormiscium</i> sooty mold	28
Brick red root fungus	47	<i>Coprinus</i>	8
Brown algal leaf spot	22	<i>Corticium</i> —	
Burma	18, 34, 40, 44	<i>javanicum</i>	30
		<i>koleroga</i>	26
		<i>lacteam</i>	47
Cacao, host for <i>Corticium</i>		<i>salmonicolor</i>	30, 31, 32, 46
<i>salmonicolor</i>	30	<i>solani</i>	5
Calcium hypochlorite, control of damping-off	8	<i>vagum</i>	3
<i>Calosphaeria cinchonae</i>	18	Costa Rica	23
Canker:		Cryptonol, control of—	
stem	17	<i>fusarium</i> and <i>helminthosporium</i> stem blight	13
trunk and branch	29	<i>rhizoctonia</i> damping-off	5
Capnodium—		Cuprocide, control of damping-off	7
and other molds	25		
<i>brasiliensis</i>	25	Cuprous oxide, control of	
Carbolic acid, control of myxomycete	8	<i>rhizoctonia</i> damping-off ..	7
Carbolineum, control of—		<i>Cystodium coccineum</i>	47
phytophthora stripe canker ..	33		
stem canker	17	Damping-off:	
Carbolineum plantarum, control of phytophthora stripe canker	33	<i>cladosporium</i>	3
		<i>clitocybe</i>	4
Carbon—		<i>fusarium</i>	4
bisulfide, control of <i>hormiscium</i> sooty mold	28	late	3
disulfide, control of <i>fusarium</i> ..		mopog (mopo)	5
root rot	38	<i>pestalozzia</i>	4
<i>Cephaleuros virescens</i>	21	<i>pythium</i>	4
		<i>rhizoctonia</i>	5
		<i>Dasyscypha warburgiana</i>	32

	Page		Page
Deficiencies:		<i>Fusarium</i> —	
chlorophyll	25	damping-off	4
iron	25	<i>moniliforme</i>	38
nutrient	25	root rot	38
<i>Dendrophoma cinchonae</i>	11	species	3, 4, 13, 29
<i>Diplodia</i> —		stem blight	12
<i>cinchonae</i>	46	<i>vasinfectum</i>	38
species	13	<i>Fusoma</i>	46
stem blight	12		
<i>theobromae</i>	12, 35, 46	<i>Ganoderma</i> —	
<i>Diplopeltis</i>	46	<i>pseudoferreum</i>	39
Djamoeer oepas	30	root rot	39
<i>Dorylaimus</i>	21	German East Africa	18
Drought injury	44	Gloeosporium	9, 22
Ecuador	22	<i>Glomerella</i>	46
Eelworms	20	<i>Glomella chinicola</i>	47
<i>Elsinoë</i> —		<i>Glomopsis regia</i>	47
<i>cinchonae</i>	27	Graphium	43
scab	26	Gray dadap fungus	28
Eritrea	25	Green algal leaf spot	22
<i>Fomes</i> —		Guatemala	4, 7, 16,
<i>lamaoensis</i>	37	21, 24, 26, 30, 33, 34, 43, 45	
<i>lamaoensis</i>	37	<i>Guignardia yersini</i>	11
<i>noxius</i>	37		
root rot	37	Hail and rain injury	45
Formaldehyde, control of—		<i>Helicobasidium compactum</i>	46
damping-off	4, 6, 7	<i>Helminthosporium</i>	13
fusarium root rot	38	<i>Heterodera marioni</i>	19, 20, 21
Formalin, control of phy-		Hevea, host for—	
tophthora stem blight	13	<i>Corticium salmonicolor</i>	31
Formosa	13	<i>Fomes noxius</i>	37
France	23	<i>Hevea brasiliensis</i> , host for	
French Guinea	4	<i>Fomes noxius</i>	37
Frost or freezing injury	44	Hide-bound condition	29
Fungi on commercial barks	47	<i>Himantia cinchonarum</i>	47
Fungicides:		Honey agaric	37
aromatic nitrated phenols	4	<i>Hormiscium</i> —	
bordeaux mixture	7, 10, 11, 13,	<i>pannosum</i>	27
15, 16, 24		sooty mold	27
calcium hypochlorite	8	Horsehair blight	26
carbolic acid	9	<i>Hymenochaete noxia</i>	37
carbolineum	17, 33	<i>Hypomyces</i> —	
carbolineum plantarum	33	<i>haematococcus</i>	46
chloride of lime	17	<i>ipomoeae</i>	46
copper	3	<i>Hysterium enteroleucum</i>	47
copper spray	11, 16		
copper sulfate	28	India	7, 12, 15, 17, 20, 21, 27, 29,
Cryptonol	6, 13	30, 33, 35, 37, 39, 41, 45, 46, 47	
Cuprocide	7	Indochina	4, 9, 10, 11, 25, 38, 39, 41
cuprous oxide	7	Injuries attributed to environ-	
formaldehyde	4, 6, 7, 38	mental factors:	
formalin	13	drought	44
Kerol	13	frost and freezing	44
lime	8, 11, 17, 27, 37	light	44
malachite green	13	lightning	45
mercuric chloride	7	rain and hail	45
Nosperit	6	shade	45
Semesan	7	smoke	45
sulfur	6, 26	wind	45
Superol	6	Irish potato, host for <i>Rhizoc-</i>	
Terbolan	6	<i>tonia solani</i>	6
Yellow Cuprocide	7, 16	Iron deficiency	25

	Page		Page
Jamaica	7, 34, 45	Nematodes	19, 20
Java. See Netherlands Indies.		Netherlands Indies. ²⁵	
Kerol, control of fusarium stem blight	13	Nosperit, control of mopog damping-off	6
Lapp disease	10	Nutrient deficiency	25
<i>Lasiodiplodia theobromae</i>	12	Nyasaland	37
Late damping-off	3	Olpidiaceae	17
Leaf mottle	25	Palm, host for <i>Phytophthora palmivora</i>	15
Leaf scab	10	<i>Pellicularia</i> —	
Leaf spot:		<i>filamentosa</i>	5
algae	21	<i>koleroga</i>	26
cephaleuros and other algae	21	Peru	7, 16, 19, 21, 22, 26, 29, 30, 33, 34, 41, 43, 47
cercospora	22	<i>Pestalozzia</i> —	
colletotrichum	9	<i>cinchonae</i>	18, 46
gloeosporium	9, 22	<i>funerea</i>	3
Lapp disease	10	<i>myristicae</i>	4
phyllosticta	11, 22	<i>Peziza</i>	30
phyllostictina	10	Philippine Islands	6, 14, 20, 34
prillieuxina	23	<i>Phlyctaena cinchonae</i>	11
sclerotinia	23	<i>Phoma cinchonae</i>	11
uredo	24	<i>Phomopsis</i>	22, 40, 46
Lichens	47	<i>Phyllosticta</i> —	
Light injury	44	<i>cinchonae</i>	10, 22, 23
Lightning injury	45	<i>cinchonaecola</i>	11
Lime, control of—		<i>cinchonicola</i>	23
armillaria root rot	37	<i>honbaensis</i>	11
leaf disease	11, 27	leaf spots	11, 22
myxomycete	8	<i>yersini</i>	11, 12
stem canker	17	<i>Phyllostictina</i> leaf spot	10
Loquat, host for <i>Corticium salmonicolor</i>	31	<i>Physalospora cinchonae</i>	12
<i>Macrophomina phaseoli</i>	46	Physiological—	
Malachite green, control of fusarium stem blight	13	stem canker	17
Malaya	14, 40, 42	root rot	39
Marasmius	8, 26	<i>Phytophthora</i> —	
Mechanical injury	8	<i>cactorum</i>	3
Mercuric chloride, control of rhizoctonia damping-off	7	canker	32
Mineral—		<i>cinchonae</i>	13
deficiencies	25	<i>cinnamomi</i>	3, 14, 40
toxicity	25	<i>colocasiae</i>	15
Mold, sooty	25, 27	<i>faberi</i>	14
<i>Moniliopsis aderholdii</i>	5	<i>palmivora</i>	15, 32
Mopog (mopo)	5	<i>parasitica</i>	3, 15, 16, 33
Mosaic	25	root rot	40
Mycorrhizae:		species	16, 17, 41
ectotrophic	49	stem blight	13
endotrophic	49	stripe canker	32
<i>Myrangium cinchonae</i>	47	trunk and branch canker	32
Myxomycete	8	Pink disease	30
<i>Necator decretus</i>	31, 46	Pink root fungus	47
<i>Nectria</i> —		<i>Pinus insularis</i> , host for <i>Rhizoctonia solani</i>	6
<i>amaniana</i>	18	<i>Polyporus</i> —	
<i>cinchonae</i>	18	<i>fimbriatus</i>	47
<i>coffeicola</i>	18	<i>rubidus</i>	47
species	13, 30, 34, 38	<i>Polystictus fimbriatus</i>	47
		<i>Poria</i>	47

²⁵ Most of the diseases discussed in this publication occur in the Netherlands Indies.

	Page		Page
<i>Prillieuxina cinchonae</i>	23	<i>Sporodesmium</i> —	
Puerto Rico	7, 16, 18, 25, 27, 33,	<i>cinchonae</i>	19
	40, 43, 44, 45	root rot	19
<i>Pythium</i>	3, 4, 7	species	20, 21
Rain and hail injury	45	<i>Sporotrichum</i>	13
<i>Rhabdospora thallicola</i>	47	Stem blight:	
<i>Rhizoctonia</i> —		<i>diplodia</i>	12
<i>bataticola</i>	46	<i>fusarium</i>	12
damping-off	5	<i>phytophthora</i>	13
<i>lamellifera</i>	46	Stem canker:	
root rot	18	<i>Olpidiaceae</i>	17
<i>solani</i>	5	physiological	17
species	3, 35	Stem rust	30
<i>Rhizomorpha</i> —		<i>Stemonitis</i>	8
<i>cinchonarum</i>	47	<i>Stigeosporium</i> —	
<i>crinum</i>	47	<i>cinchonae</i>	20
Root and collar rot	36	root rot	20
Root rot:		<i>Stilbella</i>	47
<i>armillaria</i>	37	<i>Stilbospora fumosa</i>	47
fomes	37	<i>Stilbum minutula</i>	47
<i>fusarium</i>	38	Stripe canker	32
<i>ganoderma</i>	39	Sulfur, control of—	
physiological	39	leaf disease	27
<i>phytophthora</i>	40	mopog damping-off	6
<i>rhizoctonia</i>	18	Sumatra. <i>See</i> Netherlands	
<i>rosellinia</i>	41	Indies.	
<i>sclerotium</i>	18	Superol, control of mopog	
<i>sporodesmium</i>	19	damping-off	6
<i>stigeosporium</i>	20	<i>Syncephalastrum elegans</i>	47
<i>Rosellinia</i> —		Systemic diseases	29
<i>arcuata</i>	41	Tanganyika	18, 46
<i>bunodes</i>	42	Tea, host for—	
root rot	41	<i>Corticium salmonicolor</i>	30, 31
species	37, 39	<i>Fomes noxius</i>	37
Rough bark	29	Terbolan, control of mopog	
Roundworms	20	damping-off	6
Rubber tree, host for—		<i>Thelephora</i> —	
<i>Corticium salmonicolor</i>	30	<i>aurea</i>	47
<i>Fomes noxius</i>	37	<i>cyanescens</i>	47
Russia. <i>See</i> U. S. S. R.		<i>lactea</i>	47
Salt, control of myxomycete ...	8	Thread blight	26
Saltpeter, control of myxomy-		Theadworms	20
cete	8	Tomato, host for <i>Rhizoctonia</i>	
Scab	26	<i>solani</i>	6
<i>Sclerotinia</i> —		Tracheomycosis	29
<i>fuekeliana</i>	23	Trunk and branch canker:	
leaf disease	23	<i>corticium</i>	30
<i>Sclerotium</i> —		<i>dasyscypha</i>	32
<i>rolfsii</i>	18	<i>phytophthora</i>	32
root rot	18	<i>Tubercularia</i>	30
Semesan, control of damping-		<i>Tulasnella cinchonae</i>	29
off	7	Turf fungus	9
<i>Septobasidium</i> —		<i>Tylenchorhynchus alatus</i>	21
<i>bogoriense</i>	28	<i>Tylenchus</i> —	
<i>cinchonae</i>	28	<i>alatus</i>	21
<i>lichenicolum</i>	28	<i>coffae</i>	21
Shade injury	45	Uganda	22, 25, 37, 46
Smoke injury	45	United States of America ...	22, 45
Sooty mold:		<i>Uredo</i> —	
<i>capnodium</i>	25	<i>cinchonae</i>	24
<i>hormiscium</i>	27	leaf spot	24

	Page		Page
U. S. S. R.	9, 12, 22	Wind injury	45
Verticillium	13, 29	Withertip	9
Virus disease	27	Yellow Cuprocide	7, 16
White root fungus	37	Zythia	22

☆ U. S. GOVERNMENT PRINTING OFFICE: 1947—748314

